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Primary Human T Lymphocytes Engineered with a Codon-Optimized IL-15 Gene Resist Cytokine Withdrawal-Induced Apoptosis and Persist Long-Term in the Absence of Exogenous Cytokine

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IL-15 is a common γ-chain cytokine that has been shown to be more active than IL-2 in several murine cancer immunotherapy models. Although T lymphocytes do not produce IL-15, murine lymphocytes carrying an IL-15 transgene demonstrated superior antitumor activity in the immunotherapy of B16 melanoma. Thus, we sought to investigate the biological impact of constitutive IL-15 expression by human lymphocytes. In this report we describe the generation of a retroviral vector encoding a codon-optimized IL-15 gene. Alternate codon usage significantly enhanced the translational efficiency of this tightly regulated gene in retroviral vector-transduced cells. Activated human CD4+ and CD8+ human lymphocytes expressed IL-15Re and produced high levels of cytokine upon retroviral transduction with the IL-15 vector. IL-15-transduced lymphocytes remained viable for up to 180 days in the absence of exogenous cytokine. IL-15 vector-transduced T cells showed continued proliferation after cytokine withdrawal and resistance to apoptosis while retaining specific Ag recognition. In the setting of adoptive cell transfer, IL-15-transduced lymphocytes may prolong lymphocyte survival in vivo and could potentially enhance antitumor activity. The Journal of Immunology, 2005, 175: 7226–7234.

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3 Abbreviations used in this paper: CO, codon optimized; 7-AAD, 7-aminoactinomycin; PPL, preprolactin leader.
Materials and Methods

Cell lines

The cell lines used include the rhabdomyosarcoma line TE 671 (American Type Culture Collection (ATCC) HTB-139), the highly transfactable human renal epithelial line 293T (ATCC CRL-11268), the mouse fibroblast line NIH-3T3 (ATCC CRL-1658), the human lymphoid cell line Sup T1 (ATCC CRL-1942), the TAP-deficient lymphoblastoid cell line T2 (17), the PG13 gibbon ape leukemia virus-packaging cell line (ATCC CRL-10686), and the human ectotropic packaging cell line Phoenix Eco (provided by G. Nolan, Stanford University, Stanford, CA). Cell culture medium consisted of RPMI 1640 (Invitrogen Life Technologies) supplemented with 10% FCS, 100 U/ml penicillin, 100 μg/ml streptomycin, 2 mM l-glutamine, and 25 mM HEPES buffer solution (all from Invitrogen Life Technologies). Cells lines were cultured at 37°C in a 5% CO₂ humidified incubator.

Construction of retroviral vector plasmids

The pMSGV1 retroviral vector contains a murine stem cell virus long-terminal repeat and RNA processing signals similar to the MFG class of retroviral vectors. The detailed construction of this vector was recently described (18).

Two retroviral vector plasmids coding for human IL-15 were constructed: pMSGV1 proprealin leader (PPL) IL-15, which carries the native IL-15 sequence linked to the bovine PPL, was assembled by insertion of an NcoI/BamHI fragment of an IL-15 expression construct (provided by Y. Tagaya, National Institutes of Health, Bethesda, MD) (6) into the NcoI/SnaBI sites of pMSGV1. A CO IL-15 gene was synthesized in which 63 codon substitutions were made in the mature protein region of the PPL IL-15 sequence (Blue Heron Biotechnology). These changes in the coding sequence minimized the use of rare codons while maintaining a low free energy as calculated by the Vienna RNA package (19). This synthetic gene was then cloned and inserted into the NcoI/SnaBI sites of pMSGV1 to yield pMSGV1 PPL CO IL-15.

The anti-MART-1 TCR retroviral vector, designated AIB, was used for control transductions. This vector also uses the pMSGV1 retroviral vector backbone (18).

Preparation of retrovirus for comparison of pMSGV1 PPL IL-15 and pMSGV1 PPL CO IL-15

Phoenix Eco packaging cells (5 x 10⁶) were plated into each well of a six-well tissue culture plate. On the following day, the cells were transfected with 2 μg of plasmid DNA from either pMSGV1 PPL IL-15 or pMSGV1 PPL CO IL-15 using the GenePorter reagent (Gene Therapy Systems). Twenty-four hours later, the medium was aspirated and replaced with 2 ml of fresh medium/well. Forty-eight hours post-transfection, the cell culture supernatant containing retrovirus was collected. The transfections were performed in triplicate for each plasmid; thus, three separate preparations of retrovirus were produced from each plasmid.

Retroviral transduction

Nontissue culture-treated 24-well plates (BD Biosciences) were coated with 25 μg/ml recombinant fibronectin fragment (RetroNectin; Takara). Retroviral vector supernatants were added, and the plates were incubated at 32°C for 2–4 h, followed by storage at 4°C overnight. Plates were warmed to room temperature, supernatant was removed, and 0.1–1.0 x 10⁶ cells were added to each well with 1 ml of tissue culture medium/well. The plates were then incubated overnight in a 5% CO₂ humidified incubator at 37°C.

Transduction of NIH-3T3 cells: cytokine production and PCR amplification to detect vector integration

Retroviral supernatants were used to transduce NIH-3T3 cells to assess target cell IL-15 production. Four hundred microliters of undiluted and serially diluted retroviral supernatants were applied to recombinant fibronectin-coated, nontissue culture, 24-well plates as described previously. NIH-3T3 cells (1 x 10⁶) were then transduced in each well of the retrovirus-coated plates. Twenty-four hours after transduction, the wells were washed three times with 2 ml of PBS (Biofluids) and then filled with 2 ml of fresh medium. The cell culture medium was collected 72 h after transduction, and the IL-15 content was analyzed by ELISA (R&D Systems).

PCR amplification used genomic DNA extracted from cell lysates using QuickExtract solution (Epicentre). Oligonucleotide primers flanking the multiple cloning site of the pMSGV1 retroviral vector backbone were synthesized: ggggttgacattcctctaga and accgtcgactgcagaattcg. Oligonucleotide primers for the amplification of β-actin were also used. PCR was performed for 30 cycles at 96°C for 30 s, 60°C for 30 s, and 72°C for 90 s in a PTC-200 Thermal Cycler (Global Medical Instrumentation). The PCR products were run on a 1% agarose gel and subsequently imaged and quantitated with an LAS-1000 luminescent image analyzer system (Fujifilm Medical Systems USA). The ratio of vector insert band intensity compared with the corresponding β-actin band was calculated for each of the cell lysate samples.

Generation of high titer packaging cell clones

Phoenix Eco packaging cells were transfected with pMSGV1 PPL IL-15 or pMSGV1 PPL CO IL-15 using the GenePorter reagent. The retroviral supernatant was harvested 48 h after transfection and transferred to nontissue culture plates coated with recombinant fibronectin (Takara). PG13 cells were then transduced on these plates. Clones were isolated by limiting dilution culture and screened for IL-15 production by ELISA (R&D Systems).

Retroviral supernatants produced by these clones were then applied to RetroNectin (Takara)-coated plates and used to transduce Sup T1 cells. A high titer clone produced from pMSGV1 PPL CO IL-15 was selected on the basis of IL-15 production by transduced Sup T1 cells. Genomic DNA was then extracted from the selected PG13 clone, and a Southern blot was performed to verify vector integrity and evaluate the number of integrations. This clone was used to produce retrovirus for all subsequent experiments in which human lymphocytes were transduced.

Transduction of PBLs and cytokine withdrawal studies

PBLs were obtained by leukapheresis from patients with a history of melanoma who were treated on an adjuvant peptide vaccine protocol at the National Cancer Institute. Lymphocytes were purified by centrifugation on a Ficoll/Hypaque cushion, washed in HBSS, and cultured in medium consisting of AIM-V medium (Invitrogen Life Technologies) supplemented with 300 IU/ml IL-2, 5% human AB serum (Valley Biomedical), 100 U/ml penicillin, 100 μg/ml streptomycin, 2 mM l-glutamine (all from Invitrogen Life Technologies), 55 μM 2-ME, and 25 mM HEPES buffer solution (Invitrogen Life Technologies). Polyclonally activated lymphocytes were generated by adding 50 ng/ml OKT3 on culture day 0. Peptide-reactive lymphocytes were generated from PBLs obtained from patients previously vaccinated with the anchor residue-modified gp100:209–217/210(M) peptide (20); these PBLs were activated by adding 1 μg/ml gp100:209–217/210(M) to the lymphocyte culture medium, in the absence of IL-2, on day 0. On day 1, 300 IU/ml IL-2 was added to the cells. Lymphocytes were cultured at 37°C in a 5% CO₂ humidified incubator. OKT3-activated cells were transduced on culture days 2 and 3, whereas peptide-stimulated cells were transduced between culture days 5 and 7. Each culture was exposed to retrovirus a total of two times, on successive days, by transferring 0.5–1.0 x 10⁶ cells in a volume of 1 ml of medium to each well of a 24-well, nontissue culture plate that had been coated sequentially with RetroNectin (Takara) and then retrovirus.

Studies requiring withdrawal of the cells from exogenous IL-2 were undertaken by harvesting and washing lymphocytes three times in lymphocyte culture medium without IL-2. After the final wash, the medium was tested by IL-2 and/or IL-15 ELISA (Endogen and R&D Systems, respectively) to verify that no residual cytokine was present. The cells were then resuspended in lymphocyte culture medium without IL-2 at a concentration of 1 x 10⁶ cells/ml, returned to tissue culture vessels, and enumerated and assessed for viability by trypan exclusion every 3–7 days. The medium was refreshed every 3–7 days by removing half the spent medium and replacing the volume with lymphocyte culture medium without IL-2.

Lymphocyte IL-15 production assay

Control or IL-15-transduced lymphocytes were washed three times in culture medium. Lymphocytes (2 x 10⁶) were then resuspended in cell culture medium without IL-2 and plated in round-bottom, 96-well plates. Half the wells were precocated with OKT3 Ab (200 μg/well). After 3 days in culture, the cell culture supernatants were harvested for analysis by ELISA (ELISA (R&D Systems).

Lymphocyte proliferation assay

Lymphocytes were washed three times with culture medium and plated at 1 x 10⁶ cells/well in a 96-well, round-bottom microplate in the presence of IL-2 to a total volume of 300 μl. The cells were cultured for 4 days, and in the final 16 h of culture, 1 μCi/well [methyl-3H] thymidine (PerkinElmer Life Sciences) was added to each well. Cellular DNA was harvested and counted by liquid scintillation counting.
Flow cytometry

Cell surface expression of CD3, CD4, CD8, CD27, CD28, CD45RA, CD45RO, CD62L, and CCR7 was measured using FITC-, PE-, or allophycocyanin-conjugated Abs and the corresponding isotype controls (BD Pharmingen). IL-15Ra was detected with a polyclonal biotinylated Ab and a biotinylated isotype-matched control Ab; cell surface-bound Ab was then labeled with streptavidin-PE (R&D Systems). Intracellular proteins Bcl-2 and Bcl-xL were detected using FITC- and PE-conjugated Abs and their respective isotype controls (BD Pharmingen and Southern Biotechnology Associates). Cells were washed twice in staining buffer composed of PBS with 0.5% BSA and stained with cell surface Abs or matched isotype controls, followed by incubation in the dark for 20 min at 4°C. Cells were then washed twice more with staining buffer before analysis. Intracellular staining was performed by fixing and permeabilizing the cells with Cytofix/Cytoperm solution (BD Pharmingen), two washes in Perm/Wash buffer (BD Pharmingen), staining with intracellular Abs or isotype controls, and two additional washes before analysis. Annexin V and 7-aminomethylcoumarin (7-AAD) staining for the detection of apoptotic cells was performed using the Annexin V-PE Apoptosis Kit I (BD Pharmingen). Immunofluorescence was measured using a FACSscan flow cytometer and was analyzed using CellQuest Pro software (BD Biosciences).

Lymphocyte Ag reactivity assay

Target cells were prepared by pulsing T2 cells with 10–1000 ng/ml peptide in cell culture medium for 2 h at 37°C. The following HLA-A2-restricted peptides were used: influenza peptide (GILGFVFTL), MART-1:27–35 and Bcl-xL were detected using FITC- and PE-conjugated Abs and their respective isotype controls (BD Pharmingen and Southern Biotechnology Associates). Cells were washed twice in staining buffer composed of PBS with 0.5% BSA and stained with cell surface Abs or matched isotype controls, followed by incubation in the dark for 20 min at 4°C. Cells were then washed twice more with staining buffer before analysis. Intracellular staining was performed by fixing and permeabilizing the cells with Cytofix/Cytoperm solution (BD Pharmingen), two washes in Perm/Wash buffer (BD Pharmingen), staining with intracellular Abs or isotype controls, and two additional washes before analysis. Annexin V and 7-aminomethylcoumarin (7-AAD) staining for the detection of apoptotic cells was performed using the Annexin V-PE Apoptosis Kit I (BD Pharmingen). Immunofluorescence was measured using a FACSscan flow cytometer and was analyzed using CellQuest Pro software (BD Biosciences).

Results

Generation of wild-type and CO IL-15 vectors

The redundancy of the genetic code is not reflected in a homogeneous distribution of tRNAs for certain amino acids, leading to the potential for rare codons in any given cistron. Analysis of the DNA sequence of the mature IL-15 protein revealed that 29 of the 114 codons (25%) were used with a frequency of <20% in human genes. In comparison, the IL-2 gene contains 20 rare codons in a coding sequence totaling 133 codons (15%). We designed an alternative IL-15 coding sequence taking into consideration: 1) codon frequency and 2) minimalization of nucleic acid folding, as predicted by free energy calculations. The final nucleic acid sequence of the synthetic IL-15 gene includes a total of 63 codon substitutions. Nineteen of these alternate codons replace native codons with utilization frequencies <20%. The amino acid sequence of the optimized IL-15 gene remains identical with that of the wild-type sequence (Fig. 1). The wild-type RNA transcript has a predicted free energy of 76 kcal/mol; the CO molecule has a predicted free energy of 38 kcal/mol. This synthetic gene was subcloned into the wild-type IL-15 retroviral vector, replacing the wild-type mature protein sequence while leaving the bovine PPL sequence intact.

Wild-type and CO IL-15 retroviral vectors have comparable expression in transfected cell lines

We tested the activity of the vectors pMSGV1 PPL IL-15 and pMSGV1 PPL CO IL-15 by transfecting NIH-3T3, TE671, and 293T cell lines in triplicate with serial dilutions of retroviral vector plasmid DNA. The cytokine production of each cell line was assessed by ELISA. Under all conditions tested and in every cell line, IL-15 was produced at similar levels by cells receiving either the native or CO vector DNA (Fig. 2).

To assess whether codon optimization influences the production of retrovirus by packaging cell lines, Phoenix Eco packaging cells were transfected with pMSGV1 PPL IL-15 and pMSGV1 PPL CO IL-15. The resultant retrovirus was titrated by Northern dot blot analysis (21) using a probe common to both vectors. Retroviral supernatants from Phoenix Eco cells transduced with either native or CO vector contained comparable amounts of RNA (data not shown).

Enhanced IL-15 expression in cells transduced with the CO retroviral vector

Retroviral vector preparations described in the previous section were used to transduce NIH-3T3 cells. Twenty-four hours after transduction, the plates were washed to eliminate any trace of IL-15 that could be carried over from the retroviral supernatant. Forty-eight hours later, the cell culture medium was collected and assayed for IL-15 (Fig. 3A). Cells transduced with a retrovirus coding for GFP did not produce detectable levels of IL-15 (data not shown). The cell culture medium from NIH-3T3 transduced with 2-fold serial dilutions of pMSGV1 PPL IL-15 retrovirus contained 436, 205, 98, and 44 pg/ml IL-15. In contrast, cell culture medium from NIH-3T3 transduced with similar dilutions of pMSGV1 PPL CO IL-15 retrovirus contained 1051, 498, 248, and 135 pg/ml IL-15, an ~2.5-fold increase in IL-15 production.

To rule out differences in transduction efficiencies of the wild-type and CO IL-15 retroviral vectors, genomic DNA was isolated and analyzed for vector sequences. Using identical oligonucleotide primers, PCR was performed, and vector-specific amplification was normalized to coamplified β-actin. These studies demonstrated that transduction with either the wild-type or CO retrovirus led to comparable efficiencies of gene transfer (Fig. 3B).

IL-15 vector transduction of human PBLs results in stimulation-dependent cytokine production and persistence of the cells in the absence of exogenous cytokine support

PG13 retroviral packaging cell clones carrying the IL-15 gene were generated using pMSGV1 PPL CO IL-15. A high titer producer cell clone was selected on the basis of its ability to transduce Sup T1 cells to express IL-15. A Southern blot verified vector integrity and demonstrated four sites of vector integration (data not shown). This CO IL-15 retroviral vector-packaging cell was used to produce the retrovirus used in all of the subsequent studies involving IL-15-transduced human lymphocytes. Transduction efficiencies were 50–70% after two sequential transductions, as assessed by semiquantitative PCR and real-time PCR (data not shown).

OKT3-stimulated PBLs from five patients were transduced with the IL-15 vector. Cytokine production from these cells ranged from 251 to 2905 pg/ml (untransduced cells did not produce detectable quantities of IL-15). Stimulation of the cells with plate-bound OKT3
Activated human T cells express IL-15Rα

IL-15- and IL-15Rα-deficient mice manifest similar phenotypes, exhibiting decreased numbers of CD8+ cells and nearly a total lack of memory CD8 cells, suggesting that the high affinity IL-15Rα is critical to the function of IL-15 in vivo (26). In vitro, supraphysiologic levels of cytokine can engage intermediate and low affinity IL-15 receptors on lymphocytes lacking IL-15Rα. To determine whether IL-15Rα was expressed in OKT3-activated T cells, we evaluated IL-15-transduced lymphocytes as well as untransduced control cells grown in medium containing either IL-2 or IL-15 (Fig. 5A). Untransduced lymphocytes, stimulated with OKT3 and IL-2 on day 0 of culture, expressed IL-15Rα in both CD4− and CD8+ subsets (86 and 69% positive compared with isotype control, respectively). Untransduced lymphocytes stimulated with OKT3 and grown in medium containing 100 ng/ml IL-15 did not manifest IL-15Rα staining, nor did IL-15 vector-transduced lymphocytes. Thus, exogenous or endogenous IL-15 appeared to down-regulate the expression of IL-15Rα.

Phenotype of IL-15-transduced lymphocytes

T lymphocyte subsets are often defined by cell surface expression of costimulatory molecules, adhesion molecules, and receptors; these cell surface proteins, in turn, are subject to influences such as TCR stimulation and cytokine engagement. To further define the influence of IL-15 transduction on T lymphocytes, we evaluated long-term cultures of IL-15-transduced lymphocytes grown in the absence of exogenous cytokine as well as untransduced lymphocytes grown in medium containing either IL-2 or IL-15. IL-15-transduced lymphocytes demonstrated modest increases in staining for CD27, CD28, and CD62L compared with untransduced lymphocytes cultured in IL-2 or IL-15 (Fig. 5B). This was reflected in both the percentage of cells exhibiting positive staining compared with isotype controls and the mean fluorescence intensity. No differences were seen in the expression of CD45RA, CD45RO, or CCR7 (data not shown).

OKT3-stimulated PBLs transduced with IL-15 continue to proliferate in the absence of exogenous cytokine support and resist cytokine withdrawal-induced apoptosis

When OKT3-stimulated PBLs were transduced with the IL-15 gene, they remained viable for prolonged periods in culture, but ceased to increase in cell number. We sought to dissect the mechanisms for this persistence by first evaluating thymidine incorporation. On culture day 7, untransduced, control vector-transduced, and IL-15-transduced lymphocytes were washed free of IL-2, then cultured in the presence or the absence of IL-2 for 4 days. [3H]thymidine was added during the final 16 h of culture. Assessment of thymidine incorporation revealed that the IL-15-transduced cells continued to proliferate in the absence of exogenous cytokine, whereas the untransduced or control transduced cells did not (Fig. 6).

We next evaluated whether the IL-15-transduced cells were protected from apoptotic death upon IL-2 withdrawal. Fractions of cultures of untransduced, control vector-transduced, and IL-15-transduced PBLs were sequentially withdrawn from IL-2 for 3 consecutive days, starting on culture day 7. Four days later, cells

resulted in a 2- to 5-fold increase in cytokine production; between 1186 and 3957 pg/ml IL-15 was produced after stimulation (Fig. 4A). Stimulation-dependent activation of the retroviral long-terminal repeat is well described (22–24) and was previously reported in lymphocytes retrovirally transduced with IL-2 (25).

After 7 days in culture and 4 days after the final transduction, lymphocytes were thoroughly washed to remove all traces of soluble cytokines and were returned to culture in the absence of exogenous cytokine. Untransduced lymphocytes rapidly declined in both viability and cell count. By day 30 after cytokine withdrawal, no untransduced cells could be detected. In contrast, IL-15-transduced lymphocytes uniformly persisted in vitro for >60 days (Fig. 4B). After 60 days, two of five IL-15 vector-transduced cultures significantly declined in viability, whereas the remaining three cultures persisted beyond 80 days in the absence of added cytokine. In a similar experiment conducted over a longer time course, IL-15-transduced cells persisted in vitro for 181 days, whereas untransduced lymphocytes and lymphocytes transduced with a control gene were undetectable after culture for 30 days in the absence of exogenous cytokine (Fig. 4C).

During the course of these studies, lymphocytes from 17 patients were transduced with the IL-15 vector. Consistently, IL-15-transduced lymphocyte cultures demonstrated prolonged in vitro persistence after IL-2 withdrawal compared with control cultures. In 16 of 17 cultures, viable IL-15-transduced lymphocytes were detected from 40–181 days after cytokine withdrawal. However, one of the 17 IL-15-transduced cultures exhibited logarithmic, clonal expansion for >365 days (data not shown); this cell line is under active investigation.
Levels of Bcl-2 and Bcl-xL were down-regulated in untransduced and CD4 and CD8 lymphocytes after withdrawal from IL-2; Bcl-xL dem门槛 expression of these proteins was maintained in IL-15-transduced control-transduced cells 2 days after cytokine withdrawal. In contrast, as stimulators. Comparable quantities of IFN-γ culture experiments were performed using peptide-pulsed T2 cells_round of peptide stimulation (data not shown). Subsequently, co-tetramer, with 50–80% of CD8 cells staining positively after one lymphocytes exhibited identical patterns of staining with gp100 anomaly Ag gp100 were generated and subsequently transduced with the CO IL-15 vector. Untransduced and IL-15-transduced peptide-stimulated lymphocyte cultures with reactivity to the mel-crossing lymphocytes after cytokine withdrawal, we evaluated CD4 and CD8 lymphocytes for Bcl-2 and Bcl-xL expression (Fig. 8). The expression of these proteins was maintained in IL-15-transduced CD4 and CD8 lymphocytes after withdrawal from IL-2; Bcl-xL demonstrated no decrease in expression, whereas Bcl-2 displayed a modest decrease in expression (18 and 16% decreases in IL-15-transduced CD4 and CD8 cells vs 47–48 and 53–54% decreases in control CD4 and CD8 cultures).

Specific peptide recognition by IL-15-transduced lymphocytes is maintained after withdrawal from IL-2

To further evaluate the function of IL-15-transduced lymphocytes, peptide-stimulated lymphocyte cultures with reactivity to the melanoma Ag gp100 were generated and subsequently transduced with the CO IL-15 vector. Untransduced and IL-15-transduced lymphocytes exhibited identical patterns of staining with gp100 tetramer, with 50–80% of CD8 cells staining positively after one round of peptide stimulation (data not shown). Subsequently, coculture experiments were performed using peptide-pulsed T2 cells as stimulators. Comparable quantities of IFN-γ were released by control and IL-15-transduced lymphocytes upon exposure to T2 cells pulsed with serial dilutions of the gp100 peptide (Fig. 9A).

Peptide-specific reactivity was demonstrated by secretion of IFN-γ upon culture of control and IL-15-transduced lymphocytes with T2 cells pulsed with gp100, but not the HLA-A2-restricted influenza peptide (Flu). MART peptide reactivity was seen only in the culture transduced with the MART TCR (Fig. 9B). When the lymphocyte cultures were withdrawn from IL-2, control cultures (untransduced and MART TCR transduced) declined in viability and number, whereas IL-15-transduced lymphocytes remained viable; this was similar to the pattern previously demonstrated in OKT3-activated lymphocytes (data not shown). Five days after IL-2 withdrawal, the cells were again tested for peptide reactivity against T2-pulsed target cells; control cultures demonstrated diminished IFN-γ secretion upon encounter with gp100-pulsed T2, whereas IL-15-transduced cells maintained a high level of specific IFN-γ secretion (Fig. 9C).

Discussion

In this study we have described a novel approach in the retroviral transduction of activated human lymphocytes with a CO IL-15 gene. The codon optimization process improved IL-15 expression in transduced cells. Transduced human lymphocytes produced IL-15 in quantities with a measurable biological impact, persisting in vitro for up to 180 days in the absence of exogenous cytokine and resisting cytokine withdrawal-induced apoptosis. Transduction with IL-15 did not perturb lymphocyte Ag recognition or specificity. Furthermore, IL-15-transduced lymphocytes retained the ability to recognize Ag and secrete IFN-γ after withdrawal from exogenous cytokine support.

Codon optimization is a term that has been applied to a variety of approaches in which codons are systematically altered to enhance gene expression. Highly expressed genes evolve to use codons that are highly represented in the genome (28). In some instances, gene expression can be augmented by replacing rare codons with codons favored by highly expressed genes. This strategy has been applied to enhancing murine or human expression of parasitic and viral genes, which often contain codons infrequently used in mammalian genes (29–31). Conversely, altered codon usage has been applied to systems in which production of mammalian proteins by bacterial systems is desired (32, 33). In addition to codon bias considerations, RNA secondary structure formation and stability clearly impact protein expression and can be influenced by alternate codon usage (34). This approach continues to be refined, and its use is expanding. To our knowledge, codon optimization of cytokine...
expression by human cells has not been previously demonstrated. Inefficient expression of IL-15 has been well established. The 5' untranslated region, which contains multiple AUGs, has been implicated in the obstruction of mRNA translation (15, 16). Furthermore, an inhibitory effect of the secreted isoform of the IL-15 signal peptide and mature protein C terminus was demonstrated (16, 35). The expression of IL-15 was markedly increased when the native leader sequence was replaced with either the mouse IL-2 signal peptide or the bovine PPL sequence (6). We generated a construct containing the PPL sequence and further modified the coding sequence through alternate codon usage, thereby minimizing the usage of rare codons and minimizing the free energy (and thus potential folding) of the mRNA transcript.

Interestingly, cells transfected with either the wild-type or CO IL-15 constructs produced similar amounts of protein (Fig. 2). This may be explained by the fact that transfected cells receive hundreds of copies of the plasmid DNA, possibly saturating the protein production machinery. However, retrovirally transduced NIH3T3 cells receiving the CO gene demonstrated a ~2.5-fold increase in protein expression compared with cells receiving the
wild-type gene (Fig. 3A). The NIH-3T3 cells were transduced with retrovirus of comparable titer, and assessment of retroviral integration demonstrated similar efficiency of transduction (Fig. 3B).

Thus, we conclude that the codon optimization improved gene expression at the level of mRNA stability and/or efficiency of translation.

Having established a means to engineer human lymphocytes to efficiently express IL-15, we were able to make several observations. We detected IL-15Rα on the surface of activated CD4+ and CD8+ T cells, contrary to a previous report in which activated human CD8+ T cells, expressed this receptor (36). The IL-15Rα Ab used in our studies did not bind to lymphocytes cultured in exogenous IL-15 or transduced with the IL-15 gene, implying an interaction between IL-15 and the high affinity α receptor (Fig. 5A). In these OKT3-activated T cells, we speculate that the Ab-binding site on the receptor is altered or blocked upon binding IL-15. Alternatively, IL-15Rα may be internalized after capturing IL-15. Based on the observed expression of IL-15Rα on activated T cells, it is possible that IL-15 gene expression in individual cells led to their persistence via an autocrine mechanism or that paracrine effects from a fraction of lymphocytes expressing IL-15 were sufficient to sustain the entire lymphocyte culture.

Common γ-chain cytokines promote the survival of activated T lymphocytes through common signaling pathways that induce the expression of antipapoptotic Bcl-2 family proteins (37–39). We have demonstrated that IL-15-transduced lymphocytes, withdrawn from exogenous cytokine support, exhibited continued proliferation as well as resistance to apoptosis; this coincided with the maintenance of Bcl-2 and Bcl-xL (Figs. 6–8). In vitro, this resulted in prolonged survival in IL-15-transduced lymphocytes (Fig. 4). Furthermore, peptide-stimulated, IL-15-transduced lymphocytes retained Ag recognition and specificity even after withdrawal from exogenous IL-2 (Fig. 9). In most respects, IL-15-transduced lymphocytes behaved similarly to control cell populations maintained in the presence of IL-2. This is not entirely unexpected given that IL-2Rs and IL-15Rs share β and γ subunits as well as signaling through common Jak/Stat pathways (26). Our studies corroborate previous reports in which IL-2 and IL-15 shared the ability to stimulate activated T cells in vitro (1, 2).

In several preclinical models, the addition of IL-15 to cell transfer therapy regimens significantly improved antitumor activity (13, 40, 41). Briefly, these studies have demonstrated that culturing antitumor lymphocytes ex vivo in the presence of IL-15, delivering lymphocytes bearing an IL-15 transgene, or administering adjuvant IL-15 during the course of adoptive cell transfer resulted in superior antitumor activity. These models also demonstrated superior treatment effects of IL-15 compared with IL-2 when both cytokines were used in a similar fashion. The fact that IL-2 and IL-15 seem to have similar functions in vitro, but distinct and often opposing actions in vivo, is at least partially explained by murine models demonstrating that IL-15 is captured by APCs and stromal cells expressing high levels of the high affinity IL-15Rα and subsequently presented to lymphocytes and NK cells (42–45). It seems likely that the complex interactions between IL-15-presenting cells and responding lymphocytes powerfully influence the differentiation of effector T cells. This is strongly supported by the finding that IL-15Rα-deficient memory CD8+ T cells could be sustained by other cells expressing IL-15Rα (46). Furthermore, IL-15 antitumor activity in vivo may also be enhanced by the effects of IL-15 on cells other than the transferred lymphocyte, such as APCs or NK cells. To determine whether results reported in murine models, demonstrating superiority of IL-15 in adoptive cell transfer cancer models, would be recapitulated in humans would require clinical trials in which patients receive treatment with IL-15.

The use of IL-15-expressing T cells in adoptive transfer studies involving human patients has potential drawbacks. IL-15-transgenic mice develop autoimmunity and can ultimately succumb to lymphocytic leukemia (47). In humans, abnormal expression of IL-15 has been associated with rheumatoid arthritis, inflammatory bowel disease, adult T cell leukemia, and tropical spastic paraparesis (48, 49). In our experience with IL-15-transduced human lymphocytes, one of 17 cultures evolved into a clonal population that exhibited growth for >1 year in the absence of exogenous cytokine (data not shown); it remains uncertain whether this is a consequence of IL-15 overexpression or vector integration at a site critical to cell cycle regulation. With these considerations, the clinical application of gene-engineered T cells constitutively expressing IL-15 would probably require a reliable way to terminate the response should adverse effects of the treatment arise. HSV thymidine kinase-engineered lymphocytes have proven effectiveness in the clinic (50–52). This suggests that the use of IL-15 as a survival factor in adoptive cell transfer therapy is an effective approach, provided that proof-of-principle is demonstrated in preclinical studies.
effective in controlling graft-vs-host disease in human bone marrow transplant recipients (50, 51). Thus, the development of an IL-15 retroviral vector carrying HSV thymidine kinase might permit the safe administration of IL-15-transduced, tumor-reactive lymphocytes to patients with metastatic cancer.

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Disclosures
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References


