Bacterial Lipoprotein Induces Resistance to Gram-Negative Sepsis in TLR4-Deficient Mice via Enhanced Bacterial Clearance

Gavin C. O'Brien, Jiang Huai Wang and H. Paul Redmond


http://www.jimmunol.org/content/174/2/1020

---

**References**

This article cites 38 articles, 22 of which you can access for free at:

http://www.jimmunol.org/content/174/2/1020.full#ref-list-1

**Subscription**

Information about subscribing to *The Journal of Immunology* is online at:

http://jimmunol.org/subscription

**Permissions**

Submit copyright permission requests at:

http://www.aai.org/About/Publications/JI/copyright.html

**Email Alerts**

Receive free email-alerts when new articles cite this article. Sign up at:

http://jimmunol.org/alerts
Bacterial Lipoprotein Induces Resistance to Gram-Negative Sepsis in TLR4-Deficient Mice via Enhanced Bacterial Clearance

Gavin C. O’Brien, Jiang Huai Wang, and H. Paul Redmond

TLRs are highly conserved pathogen recognition receptors. As a result, TLR4-deficient C3H/HeJ mice are highly susceptible to Gram-negative sepsis. We have previously demonstrated that tolerance induced by bacterial lipoprotein (BLP) protects wild-type mice against polymicrobial sepsis-induced lethality. In this study, we assessed whether pretreatment of C3H/HeJ mice with BLP could induce resistance to a subsequent Gram-negative Salmonella typhimurium infection. Pretreatment with BLP resulted in a significant survival benefit in TLR4-deficient C3H/HeJ mice (p < 0.0002 vs control C3H/HeJ) after challenge with live S. typhimurium (0.25 × 10⁶ CFU/mouse). This survival benefit was associated with enhanced bacterial clearance from the circulation and in the visceral organs (p < 0.05 vs control C3H/HeJ). Furthermore, pretreatment with BLP resulted in significant increases in complement receptor type 3 (CR3) and FcγRI/II expression on polymorphonuclear neutrophils (PMNs) and macrophages (p < 0.05 vs control C3H/HeJ). There was impaired bacterial recognition and phagocytosis in TLR4-deficient mice compared with wild-type mice. However, a significant augmented uptake, ingestion, and intracellular killing of S. typhimurium by PMNs and peritoneal macrophages was evident in BLP-pretreated C3H/HeJ mice (p < 0.05 vs control C3H/HeJ). An up-regulation of inducible NO synthase and increased production of intracellular NO were observed in peritoneal macrophages from BLP-pretreated C3H/HeJ mice (p < 0.05 vs control C3H/HeJ). Depletion of PMNs did not diminish the beneficial effects of BLP with regard to both animal survival and bacterial clearance. These results indicate that BLP, a TLR2 ligand, protects highly susceptible TLR4-deficient mice from Gram-negative sepsis via enhanced bacterial clearance. The Journal of Immunology, 2005, 174: 1020–1026.

Toll-like receptors are a highly conserved family of type I transmembrane proteins characterized by an extracellular domain with leucine-rich repeats and a cytoplasmic Toll/IL-1R homology domain. Thus far, only 10 mammalian TLRs have been deciphered, yet these 10 receptors are thought to recognize and orchestrate the innate immune response to a vast morass of invading pathogens (1). It is believed that TLRs overcome this apparent discrepancy in receptor to pathogen ratio by functioning as pattern recognition receptors, specifically via the recognition of pathogen-associated molecular patterns (PAMPs) (2). PAMPs are conserved structures within pathogens that activate macrophages, monocytes, and neutrophils to produce cytokines and low molecular mass mediators, thus initiating the innate immune response.

PAMPs tend to be invariant molecular structures that are essential for the survival of the pathogen and thus are far less likely to variability because mutations affecting these structures tend to result in death of the pathogen itself (2). The best-known and most exhaustively studied PAMP is endotoxin or LPS, a major structural component of Gram-negative bacteria. A wealth of biochemical and genetic evidence has identified TLR4 as the receptor that mediates cellular activation in response to LPS (3, 4). Conversely, it is known that TLR4-deficient C3H/HeJ mice are hyporesponsive to the biological effects of LPS (5). PAMPs for TLR2 include a variety of agonists derived from Gram-positive organisms such as peptidoglycan (6) and lipoteichoic acid (7–9). TLR2 plays a crucial role in the host defense against Gram-positive bacteria and their cell wall components. Therefore, it seems that TLR2 and TLR4 are activated primarily by different PAMPs to initiate the host response to Gram-positive and Gram-negative bacterial infection, respectively. This is further illustrated by the fact that TLR2 (but not TLR4) knockout mice are highly susceptible to Gram-positive bacteria, especially Staphylococcus aureus (10), whereas TLR4 (but not TLR2) knockout mice are highly susceptible to Gram-negative bacteria such as Salmonella typhimurium (5, 11).

The hyporesponsiveness of TLR4-deficient C3H/HeJ mice to LPS has been a stimulus to immunological research for >30 years (12). The apparently paradoxically high susceptibility of these C3H/HeJ mice to live Gram-negative S. typhimurium infection was somewhat clarified when the genetic defect was found to correlate with a mutation at a single locus, termed the defective Lps allele (Lpsd allele) (11). In 1997, Medzhitov et al. (13) described the first human homologue of Toll, now referred to as TLR4. Subsequently, the mutation in the Lpsd allele was gratifyingly found to correlate with the TLR4 mutation in these C3H/HeJ mice (14).

Several strategies have been explored over the years in attempts to modulate the host response to sepsis. In the early 1960s, it was...
observed that animals receiving a low-dose pre-exposure to LPS had a markedly reduced mortality when rechallenged with a "lethal" injection of endotoxin (15). This phenomenon is termed "endotoxin tolerance." Endotoxin tolerance is observed in vivo, with a decreased febrile response and an escape from lethality, as well as in vitro, with reduced production of inflammatory cytokines in response to secondary stimulation by LPS (16). Recently, there has been a dramatic resurgence in interest in the mechanisms responsible for the LPS-tolerant phenotype. It was hypothesized that LPS tolerance occurred via alteration of the intracellular signaling pathways of LPS, since LPS tolerance involves mobilization of NF-κB and a predominant accumulation of p50/p50 homodimers of NF-κB transcription factors in LPS-tolerant cells (17). It has subsequently been shown that endotoxin tolerance in mouse peritoneal macrophages correlates with down-regulation of surface TLR4 expression (16). This is an adaptive host response which may represent an essential regulatory mechanism during Gram-negative bacterial infection. The clinical applicability of LPS tolerance, however, is limited. We have previously demonstrated that LPS tolerance affords protection only against the lethal effects of a subsequent LPS challenge. LPS tolerance fails to confer protection against either a lethal bacterial (S. aureus and S. typhimurium) infection or polymicrobial sepsis (18).

Recently, in our laboratory, we have focused our attention on a lesser-studied bacterial agonist, bacterial lipoprotein (BLP). BLP is characterized by a unique NH₂-terminal lipo-amino acid, N-acyl-S-diacylglycerol cysteine and is the most abundant protein in the outer membrane of Gram-positive bacteria. BLP serves as a PAMP for Gram-positive and Gram-negative bacteria, initiating an innate immune response via TLR2. TLR2-deficient cells are sensitive to LPS stimulation but have no response to Gram-positive bacterial cell wall components (19). When these cells are transfected with TLR2, they respond to BLP stimulation (20). Furthermore, two recent studies have shown that activation of target cells by highly purified LPS is through TLR4 only and not through TLR2 (3, 21). These data indicate that, in contrast to TLR4, TLR2 is involved in this process.

Materials and Methods

Bacterial cell wall components and bacteria

BLP, a synthetic bacterial lipoprotein (PamCys-Ser-Lys-OH; Boehringer Mannheim Biochemicals), was endotoxin free as confirmed by the Limulus amebocyte assay (Charles River Endosafe). Staphylococcus typhimurium was obtained from the National University of Ireland culture collection. Bacteria were cultured at 37°C in trypticase soy broth (Merck), harvested at mid-logarithmic growth phase, washed twice, and resuspended in PBS (Invitrogen Life Technologies) for in vivo use. The concentration of re-suspended bacteria was determined and adjusted spectrophotometrically at 550 nm.

BLP pretreatment and S. typhimurium infection

Pyrogen-free male C3H/HeN and C3H/HeJ mice (8–10 wk old and 18–22 g) were purchased from Harlan. Mice were housed in barrier cages under standard aseptic conditions (22). Control mice received an equal volume (200 μl) of PBS 24 h before septic challenges. Gram-negative sepsis was induced via i.p. injection of 200 μl of PBS containing live S. typhimurium (0.25 × 10⁶ CFU/mouse). Survival was monitored for at least 14 days.

Enumeration of bacteria in blood and organs

Control and BLP-pretreated C3H/HeN and C3H/HeJ mice were cuffed at 24 and 48 h after i.p. injection of live S. typhimurium (0.25 × 10⁶ CFU/mouse). Samples of blood were taken and the dissected lungs, liver, and spleen were homogenized. Serial 10-fold dilutions of whole blood and organ homogenates in sterile water containing 5% Triton X-100 (Sigma-Aldrich) were plated on trypticase soy agar (Merck) and cultured for 24 h at 37°C for determination of bacterial CFU.

Neutrophil depletion

C3H/HeJ mice were injected i.p. with cyclophosphamide (Sigma-Aldrich) at a dose of 250 mg/kg on day 0 and 100 mg/kg on day 3 to produce neutropenia. On day 4, BLP pretreatment was conducted as described above. Twenty-four hours later, control and BLP-pretreated mice received an i.p. injection of 200 μl of PBS containing 0.25 × 10⁶ CFU of viable S. typhimurium, and animal survival was monitored for at least 14 days. The number of S. typhimurium in the blood and organs was also determined as described above.

Peritoneal macrophage isolation

Peritoneal cells were isolated from C3H/HeN and C3H/HeJ mice by peritoneal lavage using ice-cold PBS. Cells were incubated with RPMI 1640 medium (Invitrogen Life Technologies) containing 10% FCS (Invitrogen Life Technologies) in 24-well plates (Falcon) at 37°C in 5% CO₂ for 90 min; nonadherent cells were washed twice, and resuspended macrophages (2 × 10⁶ cells/well) were further cultured in fresh RPMI 1640 medium supplemented with 10% FCS.

FACScan analysis of CR3 and FcγRII/IIR expression

Heparinized blood and peritoneal lavage were collected from C3H/HeN, control, and BLP-pretreated C3H/HeN mice and dual-stained with anti-Ly-6G (a marker for murine polymorphonuclear neutrophils (PMNs; BD Pharmingen), anti-F4/80/Ag (a marker for murine macrophages; Serotec), and anti-Complement receptor type 3 (CR3; BD Pharmingen), and anti-FcγRII/IIR (BD Pharmingen) mAbs conjugated with PE or FITC. PE- or FITC-conjugated anti-mouse isotype-matched Abs (BD Pharmingen) were used as negative controls. Erythrocytes were lysed using lysis buffer (BD Biosciences). FACScan analysis was performed from at least 5000 events for detecting expression of CR3 and FcγRII/IIR on PMNs (Ly-6G-positive cells) and macrophages (F4/80-positive cells) using CellQuest software (BD Biosciences).

Analysis of bacterial uptake, phagocytosis, and intracellular killing

Staphylococcus typhimurium was heat-killed at 95°C for 20 min and labeled with 0.1% FITC (Sigma-Aldrich) in 0.1 M carbonate buffer (pH 9.5) at 37°C for 60 min. Heparinized blood and peritoneal lavage collected from...
CH3HeN, control, and BLP-pretreated mice were incubated with 1 × 10^6 CFU/ml heat-killed, FITC-labeled S. typhimurium at 37°C for 15 min. Bacterial uptake by PMNs and peritoneal macrophages was assessed by FACScan analysis. PMN and peritoneal macrophage populations were identified by their positive staining for anti-Ly-6G and anti-F4/80, respectively. Bacterial phagocytosis was further determined after the external fluorescence of the bound, but noningested bacteria was quenched with 0.025% crystal violet (Sigma-Aldrich). Intracellular bacterial killing was determined as previously described (18). Briefly, peritoneal macrophages were cultured in RPMI 1640 medium containing 10% FCS and incubated with live S. typhimurium (macrophage:bacteria = 1:20) at 37°C for 60 min in the presence of cytochalasin B (5 μg/ml; Sigma-Aldrich) to assess extracellular bacterial killing or absence of cytochalasin B to assess total bacterial killing. After macrophages were lysed, total and extracellular bacterial killing were determined by incubation of serial 10-fold dilutions of the lysates on tryptone soy agar (Merck) plates at 37°C for 24 h. Intracellular bacterial killing was calculated according to the total and extracellular bacterial killing.

Detection of inducible NO synthase (iNOS) expression and intracellular NO production

The expression of iNOS in peritoneal macrophages was detected by Western blot analysis. Briefly, peritoneal macrophages pooled from three control or three BLP-pretreated C3H/HeJ mice were lysed in ice with lysis buffer (1% Triton X-100, 20 mM Tris, 137 mM NaCl, 1 mM PMSF, 2 mM Na3VO4, 10 μg/ml leupeptin, and 2 μg/ml aprotinin). Protein concentrations were determined using a micro-BCA protein assay reagent kit (Pierce). The proteins were denatured at 95°C for 10 min in loading buffer (60 mM Tris, 2.5% SDS, 10% glycerol, 5% 2-ME, and 0.01% bromphenol blue). Aliquots containing equal amounts of total protein from each sample were separated in SDS-PAGE and transblotted onto nitrocellulose membranes (Schleicher & Schuell Microscience). After blocking for 2 h with TBS containing 0.1% Tween 20 and 6% nonfat milk, membranes were probed overnight at 4°C with anti-iNOS pAb (Chemicon). Blots were further incubated for 1 h with secondary HRP-conjugated anti-rabbit IgG (DakoCytomation) and developed using an ECL detection system (Santa Cruz Biotechnology) according to the manufacturer’s instructions. The intracellular NO formation in peritoneal macrophages was detected by using the fluorescent probe 4-amin-5-methylamino-2', 7'-difluorescein (DAF-FM) diacetate (Molecular Probes) as previously described (25, 26). Briefly, peritoneal cells isolated from control and BLP-pretreated C3H/HeJ mice by peritoneal lavage were incubated in RPMI 1640 medium without phenol red (Invitrogen Life Technologies). Cells were stained with anti-F4/80 Ag mAb conjugated with PE (Serotec) for identifying the macrophage population and further loaded with 5 μM DAF-FM diacetate for 30 min. FACScan analysis for detecting intracellular NO formation in peritoneal macrophages (F4/80-positive cells) was performed from at least 10,000 events on a flow cytometer (BD Biosciences) to detect the log of the mean channel fluorescence intensity with an acquisition of FL1 using CellQuest software (BD Biosciences).

Statistical analysis

All data are presented as the mean ± SD. Statistical analysis was performed using the log rank test for survival studies and the Mann-Whitney U test for all others. Differences were judged statistically significant when the p < 0.05.

Results

BLP induces resistance to Gram-negative sepsis in TLR4-deficient mice

We previously demonstrated that BLP is capable of inducing cross-tolerance to both LPS and live bacterial (S. aureus and S. typhimurium) challenges in mice (18). We hypothesized that BLP could protect TLR4-deficient mice against Gram-negative sepsis, to which these mice are highly susceptible. In our first experiment, TLR4-deficient C3H/HeJ mice were randomized to either receive 10 mg/kg BLP i.p. 24 h before bacterial challenge or to receive a control i.p injection of PBS. After 24 h, each mouse received 0.25 × 10^8 CFU of live S. typhimurium i.p. Survival was monitored for at least 14 days. Pretreatment with BLP conferred a statistically significant survival benefit against S. typhimurium infection and thus protects TLR4-deficient mice against Gram-negative sepsis.

BLP induces resistance to Gram-negative sepsis in TLR4-deficient mice via enhanced bacterial clearance

Having shown a survival benefit, we hypothesized that BLP may confer this benefit via enhanced bacterial clearance. To test this hypothesis, we challenged three groups of mice. The groups consisted of a control group of C3H/HeN mice with normal TLR4 and two groups of TLR4-deficient C3H/HeJ mice, one-half of which were pretreated with BLP (10 mg/kg). Following exposure to 0.25 × 10^6 CFU of live S. typhimurium, mice were culled at 24 and 48 h, respectively. At each time point, samples of blood and homogenates of lungs, liver, and spleen were cultured at 37°C for 24 h whereupon bacterial CFU were enumerated and compared. At the 24-h time point (Fig. 2a), we see the significant differences in bacterial dissemination in the bloodstream, lungs, and livers of control C3H/HeJ mice compared with the control C3H/HeN mice, highlighting the grossly dysfunctional Gram-negative bacterial clearance capabilities in animals with a mutated TLR4 (p < 0.05). We also saw how BLP pretreatment leads to a dramatic reinstatement of bacterial clearance function to TLR4-deficient mice. Even at this early time point of 24 h, pretreatment with BLP resulted in a dramatic attenuation in bacterial CFU counts in both the bloodstream and the lungs of the C3H/HeJ mice compared with their control counterparts (p < 0.05). BLP led to bacterial clearance rates from the bloodstream and lungs which approach those of the wild-type C3H/HeN mice.

At the 48-h time point (Fig. 2b), we note a gross difference in bacterial CFU counts between C3H/HeN mice and control C3H/HeJ mice in all samples (p < 0.05). Pretreatment with BLP resulted in a significantly enhanced bacterial clearance from the blood and all of the solid viscera of the C3H/HeJ mice compared with the control C3H/HeJ animals (p < 0.05). We conclude that BLP protects TLR4-deficient mice against Gram-negative sepsis via enhanced bacterial clearance.

Enhanced PMN and macrophage activation in BLP-pretreated TLR4-deficient mice

We determined CR3 and FcyIII/IIR expression on PMNs and peritoneal macrophages from control and BLP-pretreated TLR4-deficient mice. Pretreatment with BLP resulted in an increase in the
circulating PMN population and a recruitment of PMNs into the peritoneal cavity. There was an associated increase in surface expression of CR3 and FcγRIII/IIR expression on these cells \((p < 0.05; \text{Fig. 3})\). In addition, BLP pretreatment significantly up-regulated the expression of these two receptors on peritoneal macrophages \((p < 0.05)\) (Fig. 3).

### Increased bacterial recognition and bactericidal activity in BLP-pretreated TLR4-deficient mice

We next assessed the ability of PMNs and peritoneal macrophages to recognize, phagocytose, and kill bacteria. In this experiment, we compared cells from wild-type C3H/HeN mice with both control and BLP-pretreated C3H/HeJ mice. After incubation with heat-killed or live *Salmonella typhimurium*, PMNs and peritoneal macrophages from control mice had significantly less uptake, phagocytosis, and intracellular killing compared with cells from C3H/HeN mice with normal TLR4 receptors \((p < 0.05; \text{Fig. 4})\). Pretreatment of C3H/HeJ mice with BLP led to significant increases in PMN and macrophage uptake and phagocytosis of *S. typhimurium* \((p < 0.05; \text{Fig. 4a})\). Furthermore, intracellular killing of *S. typhimurium* by peritoneal macrophages was significantly enhanced in BLP-pretreated mice compared with control C3H/HeJ mice \((p < 0.05; \text{Fig. 4b})\).

### Up-regulation of iNOS expression and increased intracellular NO production in BLP-pretreated TLR4-deficient mice

To further identify possible mechanisms involved in the enhanced bactericidal activity observed in BLP-pretreated TLR4-deficient mice, we assessed the expression of iNOS and the production of intracellular NO in peritoneal macrophages from control and BLP-pretreated C3H/HeJ mice. Western blot analysis revealed that iNOS expression was significantly up-regulated in peritoneal macrophages after pretreatment with BLP in TLR4-deficient mice, whereas this protein was undetectable in peritoneal macrophages from control C3H/HeJ mice (Fig. 5a). Furthermore, assessment of intracellular NO formation using the fluorescent probe DAF-FM diacetate-based FACS analysis showed that the increased production of intracellular NO by the peritoneal macrophages from BLP-pretreated C3H/HeJ mice was statistically significant when compared with NO levels produced by the peritoneal macrophages from control C3H/HeJ mice \((p < 0.05; \text{Fig. 5b and c})\).

### BLP-allowed protection against Gram-negative sepsis is partly independent of neutrophil function

To determine whether PMNs play a crucial role in BLP-allowed protection against Gram-negative bacterial infection in TLR4-deficient C3H/HeJ mice, we repeated both survival study and bacterial clearance experiments, but first depleted the C3H/HeJ mice of their neutrophil population using i.p. cyclophosphamide injection (mean reduction of 96% PMNs, data not shown). Although neutrophil depletion resulted in a faster mortality rate in both groups,
pretreatment of C3H/HeJ mice with BLP resulted in a significant survival benefit over the control mice ($p < 0.0043$; Fig. 6). Furthermore, in neutropenic C3H/HeJ mice, BLP pretreatment resulted in a statistically significant increase in bacterial clearance from the blood, lungs, and spleens compared with the control neutropenic C3H/HeJ mice at the 24- and 48-h time points ($p < 0.05$; Fig. 7).

**Discussion**

Despite significant advances in our understanding of the molecular and genetic basis of sepsis and its associated immunologic response, sepsis remains a problem worldwide. Sepsis is the primary cause of death in the noncoronary intensive care unit (27) and septic shock is estimated to be responsible for $\geq 100,000$ deaths annually in the United States (28). In most institutions, Gram-negative bacteria account for $\sim 50\%$ of all bacteremias, with mortality estimated at $10\%$ for normal individuals and $30\%$ in immunocompromised patients (29).

The cloning of TLR4 and its identification as the primary LPS receptor in mammals prompted the discovery that specific mutations in the TLR4 gene (Asp299Gly and Thr399Ile) are associated with a diminished airway response to inhaled LPS in normal human volunteers (30). Subsequently, it has been shown that within an intensive care unit-based septic shock cohort, patients with a TLR4 polymorphism had more severe disease and an increased frequency of Gram-negative infections. These findings indicate that mutations in the TLR4 may predispose patients to Gram-negative septic shock (31). A similar study in a human septic-shock population suggested an association between a novel TLR2 (Arg753Gln) polymorphism and Gram-positive staphylococcal infection (32).

We have previously shown that BLP, a TLR2 agonist, is capable of inducing cross-tolerance to *S. typhimurium*, Gram-negative bacteria which signals via TLR4. In this article, we have gone on to show that BLP pretreatment can protect TLR4-deficient mice against live *S. typhimurium* challenge, to which these mice are extremely susceptible. The main mechanism underlying this process seems to be associated with enhanced bacterial clearance. Pretreatment of TLR4-deficient mice with BLP leads to enhanced clearance from the bloodstream and greatly reduced end-organ bacterial dissemination of *S. typhimurium* compared with control animals.
Since C3H/HeJ mice have mutated TLR4 receptors, it is clear that the enhanced bacterial clearance cannot be due to an up-regulation or modification of TLR4 but rather due to enhanced nonspecific antibacterial function. The uptake, phagocytosis, and killing of invading bacteria are mediated via activation of CR3 and FcγRII/IIIR (33, 34). Overexpression of these two receptors is associated with enhanced bacterial clearance (35). To investigate possible mechanisms contributing to the enhanced host defense against Gram-negative sepsis observed in BLP-pretreated mice, we assessed CR3 and FcγRII/IIIR expression on PMNs and peritoneal macrophages. Pretreatment with BLP resulted in an increased PMN population in the circulation and peritoneal cavity, with overexpression of CR3 and FcγRII/IIIR on PMNs as well as on peritoneal macrophages. To determine whether this altered phenotype of PMNs and peritoneal macrophages is associated with BLP- afforded protection against Gram-negative sepsis, we further assessed the ability of these cells to recognize, ingest, and kill S. typhimurium. PMNs and peritoneal macrophages from BLP-pretreated TLR4-deficient mice, in keeping with the activation of phagocytic receptors on these cells, exhibited increased bacterial recognition, phagocytosis, and intracellular killing. The enhanced bacterial recognition and microbicidal activity observed in BLP-pretreated TLR4-deficient mice may contribute toward their improved survival over control TLR4-deficient mice following lethal Gram-negative sepsis.

Next, we attempted to identify possible molecular mechanisms that contribute to the enhanced peritoneal macrophage-mediated intracellular killing of S. typhimurium observed in BLP-pretreated TLR4-deficient C3H/HeJ mice. Macrophages, the resident professional phagocytes, are an early sensor of bacterial infection via up-regulation of iNOS expression and NO production. NO produced by macrophages is responsible for the initial microbicidal activity against intracellular pathogens including S. typhimurium (36, 37). In iNOS knockout mice, failure to generate NO leads to susceptibility to S. typhimurium infection with an increased mortality (37, 38). Furthermore, peritoneal macrophages derived from iNOS knockout mice display impaired intracellular killing of S. typhimurium (36). In the present study, we showed that pretreatment with BLP in TLR4-deficient C3H/HeJ mice resulted in up-regulation of iNOS expression and increased production of intracellular NO. This may partially explain the enhanced intracellular killing of S. typhimurium by peritoneal macrophages in vitro and the accelerated bacterial clearance in vivo observed in BLP-pretreated C3H/HeJ mice.

PMNs are the first line of defense against bacterial infection. To further assess the mechanisms underlying the enhanced survival and bacterial clearance seen in BLP-pretreated TLR4-deficient mice, we repeated both the survival study and the bacterial clearance studies, first having depleted the mice of their neutrophils using i.p. cyclophosphamide. To our surprise, BLP-pretreated, TLR4-deficient, neutropenic mice again displayed a statistically significant survival benefit over their control counterparts. Despite neutropenia, BLP-pretreated mice showed enhanced bacterial clearance from the bloodstream and visceral organs following live S. typhimurium challenge. These findings, in combination with our in vitro findings, suggest that macrophage-mediated phagocytosis and microbicidal activity must play a paramount role in BLP-pretreated TLR4-deficient mice.

The emerging wealth of evidence pointing toward a genetic predisposition in individual patients to develop sepsis will have important ramifications in the clinical setting. Already studies have shown a clinical correlation between TLR4 and TLR2 polymorphisms in humans and the development of Gram-negative and Gram-positive sepsis, respectively (31, 32). Individuals with these naturally occurring polymorphisms represent an extremely high-risk population in terms of the development of postoperative sepsis, for whom, to date, no specific prophylactic measures exist.
With future increased availability of genetic profiling, it will become possible to preoperatively identify patients with TLR polymorphisms. This will allow the application of tailored prophylactic measures to prevent postoperative sepsis and morbidity. Our findings may represent one such measure. Pretreatment with BLP mediates a novel adaptive host response associated with unique protective effects during septic shock. Unlike LPS tolerance, BLP can induce cross-tolerance to LPS, live bacterial, and polymicrobial sepsis-induced lethality. In this study, we have demonstrated the unique ability of BLP to induce resistance to *S. typhimurium* infection in TLR4-deficient mice and thus protect these mice against Gram-negative sepsis.

References