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Cutting Edge: \( \text{CD}^+ \text{CD}25^+ \) Regulatory T Cells Impaired for Intestinal Homing Can Prevent Colitis

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Transfer of \( \text{CD}^+ \text{CD}45RB^{\text{high}} \) \( T \) cells into \( \text{RAG}^{-/-} \) mice causes colitis, which can be prevented by \( \text{CD}^+ \text{CD}25^+ \) regulatory \( T \) cells (Treg). Colitis induction by \( \text{CD}^+ \text{CD}45RB^{\text{high}} \) \( T \) cells requires \( \beta_7 \) integrin-dependent intestinal localization, but the importance of \( \beta_7 \) integrins for Treg function is unknown. In this study, we show that \( \beta_7^{-/-} \) Treg were effective in preventing colitis. Treg expanded in vivo to the same extent as \( \text{CD}^+ \text{CD}45RB^{\text{high}} \) \( T \) cells after transfer and they did not inhibit \( \text{CD}^+ \text{CD}45RB^{\text{high}} \) \( T \) cell expansion in lymphoid tissues, although they prevented the accumulation of Th1 effector cells in the intestine. \( \beta_7^{-/-} \) Treg were significantly reduced in the large intestine, however, compared with wild-type Treg, and regulatory activity could not be recovered from the intestine of recipients of \( \beta_7^{-/-} \) Treg. These data demonstrate that Treg can prevent colitis by inhibiting the accumulation of tissue-seeking effector cells and that Treg accumulation in the intestine is dispensable for colitis suppression. The Journal of Immunology, 2005, 174: 7487-7491.

The transfer of naive or \( \text{CD}^+ \text{CD}45RB^{\text{high}} \) \( T \) cells into SCID or \( \text{RAG}^{-/-} \) mice leads to colitis mediated by Th1 cytokines (1, 2), and this can be prevented by cotransfer of \( \text{CD}^+ \text{CD}25^+ \) regulatory \( T \) lymphocytes (Treg) (3). The mechanisms by which \( \text{CD}^+ \text{CD}25^+ \) Treg prevent intestinal inflammation in vivo include IL-10 and TGF-\( \beta \) secretion (4-8).

Efficient localization and retention of lymphocytes in the intestine are dependent on \( \beta_7 \)-containing integrins (9). \( \alpha \),\( \beta_7 \) integrin binds to the mucosal addressin cell adhesion molecule 1, which facilitates extravasation of lymphocytes through high endothelial venules in the mucosa (10, 11). The \( \alpha \),\( \beta_7 \) integrin may facilitate entry and/or retention of lymphocytes into the epithelial layer via interactions with E-cadherin-expressing epithelial cells (12-14). The importance of \( \beta_7 \) integrins is evidenced by the paucity of intestinal lymphocytes in mice lacking \( \beta_7 \) integrins (\( \beta_7^{-/-} \) mice) (15). There also is evidence from experimental models that interaction of \( \alpha \),\( \beta_7 \) integrin with mucosal addressin cell adhesion molecule 1 is important for colitis pathogenesis (16-18). It is not known, however, whether \( \text{CD}^+ \text{CD}25^+ \) Treg likewise require \( \beta_7 \) integrin expression and intestinal homing to prevent disease. In this study, we have analyzed these issues by comparing transferred \( \beta_7^{+/+} \) and \( \beta_7^{-/-} \) Treg for their ability to accumulate in different sites and their effects on cotransferred \( \text{CD}^+ \text{CD}45RB^{\text{high}} \) \( T \) cells and colitis induction.

Materials and Methods

Mice

C57BL/6J, C57BL/6 CD45.1 congenic, and C57BL/6 \( \beta_7^{-/-} \) mice (15) were obtained from The Jackson Laboratory. C57BL/6 CD45.1 congenic RAG2-/- mice were obtained from the Taconic Laboratory. Studies were approved by the Institutional Animal Care and Use Committee.

Adoptive transfer of \( T \) cells

Splenocytes were enriched for \( \text{CD}^+ \) cells by positive selection on MACS columns using anti-CD4 microbeads (Miltenyi Biotec). Cells then were stained with FITC anti-CD4 (16A), PE anti-CD25 (PC61), and PE-Cy5 anti-TCR (H57-597) and sorted into \( \text{CD}^+ \text{CD}45RB^{\text{low}} \) (CD51.1 donors) or \( \text{CD}^+ \text{CD}45RB^{\text{high}} \) \( \beta_7^{+/+} \) or \( \beta_7^{-/-} \) CD51.2 donors) populations. CD45.1 RAG2-/- recipients were injected i.v. with 5 X 10^6 \( \text{CD}^+ \text{CD}45RB^{\text{low}} \) lymphocytes from either \( \beta_7^{+/+} \) or \( \beta_7^{-/-} \) mice.

Assessment of disease

Recipient mice were weighed weekly and observed for signs of illness. Intestinal tissue samples were fixed, embedded, stained, and scored as described previously (19). All samples were coded and scored blinded to the experimental conditions.

Flow cytometric analysis of lymphocytes

Mucosal lymphocytes were isolated according to a previously published method (20). Isolated splenocytes, peripheral lymph node cells, mesenteric lymph node (MLN) cells, intraepithelial lymphocyte, and lamina propia lymphocyte (LPL) were stained with PE-Cy5 or allophycocyanin anti-TCR (H57-597), FITC anti-CD45.1 (A20), PE anti-CD25 (PC61), and PE-Cy5 or allophycocyanin anti-CD4 (GK1.5) (BD Pharmingen).

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5 Abbreviations used in this paper: Treg, regulatory T cell; MLN, mesenteric lymph node; LLPL, large intestine lamina propia lymphocyte.
**In vitro stimulation of lymphocytes**

Large intestine IEL (LIEL) or large intestine LPL (LLPL) at 10^9/ml were stimulated with 10 μg/ml anti-CD3ε clone 2C11 (BD Pharamingen). In cocultures, 1 × 10^5 CFSE-labeled CD4^+ CD45RB^high T cells were cultured at a 1:1 ratio with total CD4^+ IEL/LLPL isolated from RAG^-/- transfer recipients. LIEL and LLPL were combined and further purified by positive selection with anti-CD4 microbeads, and the cells were stimulated with 5 × 10^9 T cell-depleted spleen cells as APC and 1 μg/ml anti-CD3ε for 48 h.

For ELISA or coculture experiments, supernatants or cells were harvested for 2 h before the addition of GolgiStop (BD Pharmingen) and cultured for an additional 6 h. Cells were permeabilized and stained with allophycocyanin-labeled anti-IL-4 clone BVD4-1D11 and PE-labeled anti-IFNγ clone XMG1.2 (BD Pharmingen).

**Results**

**β^-/- Treg prevent colitis**

Transfer of splenic CD4^+ CD45RB^high T cells into RAG^-/- mice led to a severe wasting syndrome by 6-12 wk (Fig. 1a), but at 12 wk after reconstitution with CD4^+ CD45RB^high T cells and β^-/- Treg, all recipients appeared healthy and continued to gain weight (Fig. 1a). To assess the role of intestinal localization in Treg function, we used β^-/- CD4^+ CD25^+ Treg. Because it has been reported that αβ-expressing CD4^+ CD25^+ Treg are more potent than their αβ^-/- negative counterparts from wild-type mice (21), we assessed the potency of CD4^+ CD25^+ Treg from β^-/- mice to ascertain they are not impaired. CD4^+ CD25^+ Treg from β^-/- mice were as capable as wild-type cells in inhibiting the proliferation of CD4^+ CD45RB^high T cells in vitro (data not shown). We reasoned that β^-/- CD4^+ CD25^+ Treg might not efficiently repopulate the intestine of recipients. Interestingly, however, β^-/- Treg were also capable of inhibiting all clinical signs of colitis, including weight loss (Fig. 1a).

Consistent with the lack of weight loss, recipients of either β^-/+/+ or β^-/- Treg displayed no histological evidence of intestinal inflammation (Fig. 1b). Tissue sections from CD4^+ CD45RB^high T cell recipients had an average histological score of 6.1 ± 0.3 (Fig. 1a) and were characterized by inflammatory infiltrate, epithelial hyperplasia, crypt cell damage, and goblet cell depletion. By contrast, when 1 × 10^5 Treg from either β^-/+ or β^-/- mice were cotransferred, the average histological scores were zero (Fig. 1a). Transfer of only 5 × 10^4 β^-/- Treg also completely prevented colitis pathogenesis by 10 times as many CD4^+ CD45RB^high T cells (data not shown). This is a higher ratio of pathogenic cells to Treg than has been used in most other experiments, suggesting that the β^-/- Treg are highly effective at preventing disease. It remains possible, however, that wild-type Treg are effective at even lower ratios and that β^-/- Treg are slightly impaired compared to wild-type cells.

**β^-/- Treg are underrepresented in the intestine**

To compare the in vivo expansion of CD4^+ CD45RB^high T cells and Treg, tissues were analyzed 10–12 wk after transfer for the ratio of CD4^+ CD45RB^high (CD45.1^+) T cells: CD4^+ CD25^+ (CD45.2^+) Treg. When the input ratio of CD4^+ CD45RB^high β^-/+/+ CD4^+ CD25^+ Treg was 5:1, recipients approximately maintained this 5:1 ratio in almost all organs analyzed, including LPL from the large intestine (Fig. 2). This demonstrates that Treg expanded in vivo in immune-deficient mice to a similar extent as CD4^+ CD45RB^high T cells. RAG^-/- mice cotransferred with CD4^+ CD45RB^high T cells and β^-/- CD4^+ CD25^+ Treg also maintained ratios of CD4^+ CD45RB^high T cells:Treg similar to the input 5:1 ratio in the spleen and lymph nodes, but not in the intestine (Fig. 2).

**FIGURE 1. β^-/- CD4^+ CD25^+ Treg prevent colitis.** a. CD4^+ CD45RB^high T cells from C57BL/6 CD45.1 congenic mice were injected into C57BL/6 CD45.1 congenic RAG^-/- mice in the presence or absence of 1 × 10^9 CD4^+ CD25^- Treg derived from CD45.2^+ C57BL/6 β^-/+/+ or β^-/- mice. Weight gain/loss was scored weekly. Values are shown as a percentage of original weight. Error bars indicate SEM and the number of mice in each group is indicated in parentheses. Next to the weight loss curves, the average histology scores (H.S.) are indicated. Data represent one of three independent experiments with similar results. b. Mice were transplanted with the indicated cell populations and isolated tissues from the proximal large intestine were sectioned, stained, and scored. Note goblet cell depletion, epithelial disorganization, and massive lymphocytic infiltration in the tissue section from a recipient of CD4^+ CD45RB^high T cells without Treg. Original magnification, ×100.

**FIGURE 2. β^-/- Treg fail to accumulate in the large intestine.** RAG^-/- recipients of a 5:1 ratio of CD4^+ CD45RB^high:CD4^+ CD25^- Treg were analyzed by gating on TCRβ^-/- CD45.2^- cells (progeny of donor CD4^+ CD45RB^high cells) or TCRβ^-/- CD45.2^- (from donor CD4^+ CD25^- cells). The composite ratios of CD4^+ CD45RB^high:CD4^+ CD25^- T cells after 12 wk are indicated on the y-axis. Gray bars indicate recipients of β^-/+/+ and solid bars indicate recipients of β^-/- CD4^+ CD25^- Treg. Error bars represent SEM and the asterisks indicate a statistically significant difference comparing β^-/- to β^-/+/+ Treg (Student’s t test). Results shown are from five to nine mice per group.
In LLPL of recipients of $\beta_7^{+/+}/CD4^{+}CD25^{+}$ Treg, the ratio of CD4$^+CD45RB^{high}$ T cells:Treg was $>20:1$. This difference compared with cotransfer of $\beta_7^{-/-}/CD4^{+}CD25^{+}$ Treg was statistically significant ($p < 0.05$). The ratios of CD4$^+CD45RB^{high}\ T\ cells$: $\beta_7^{-/-}$ Treg were even higher in LIEL.

### Treg prevent the accumulation of T lymphocytes in the intestine

In addition to the ratios, the absolute numbers of donor CD4$^+CD45RB^{high}$ lymphocytes and Treg in RAG$^{-/-}$ recipients were analyzed. In the absence of Treg, CD4$^+CD45RB^{high}$ T cells accumulated in all recipient sites analyzed (Table I). As might be expected, this accumulation was especially evident in the large intestinal lamina propria, where the bacterial antigenic load is greatest and inflammation was pronounced. When either $\beta_7^{+/+}$ or $\beta_7^{-/-}/CD4^{+}CD25^{+}$ Treg were cotransferred, there was no difference in the accumulation of CD4$^+CD45RB^{high}$ T cells in lymph nodes and an ~2-fold decrease in the spleen. The presence of Treg had the greatest impact, however, on CD4$^+CD45RB^{high}\ T\ cell\ accumulation\ in\ the\ large\ intestine$. In the lamina propria, CD4$^+CD45RB^{high}$ T cell numbers were 7.9- to 4.3-fold reduced in recipients of $\beta_7^{+/+}$ and $\beta_7^{-/-}/CD4^{+}CD25^{+}$ Treg, respectively, compared with recipients without Treg (Table I). A similar reduction in donor-derived CD4$^+CD45RB^{high}\ T\ cells\ in\ the\ epithelium$ was observed in the epithelium.

### Intestinal IFN-$\gamma$ production is suppressed by Treg

A hallmark of the transfer model of colitis is secretion of IFN-$\gamma$ and TNF-$\alpha$ by donor lymphocytes in the lamina propria (22). Consistent with this, LLPL, derived from recipient mice injected with CD4$^+CD45RB^{high}$ T cells secreted 36.4 ± 8.5 (SEM) ng/ml IFN-$\gamma$ upon in vitro restimulation with anti-CD3e mAb. LLPL cytokine production was decreased to 1.7 ± 0.3 ng/ml when the recipients had received a 5:1 ratio of CD4$^+CD45RB^{high}$ T cells:$\beta_7^{+/+}$ Treg, $\beta_7^{-/-}$ Treg had a similar effect, with 1.4 ± 0.3 ng/ml IFN-$\gamma$ released. This was confirmed by intracellular cytokine staining of anti-CD3-stimulated LIEL or LLPL from the recipients. Cotransfer of either $\beta_7^{+/+}$ or $\beta_7^{-/-}/CD4^{+}CD25^{+}$ Treg greatly reduced the percentage of progeny of the donor CD4$^+CD45RB^{high}\ T\ cells\ that\ produced\ IFN-$\gamma$ (Fig. 3), while no intracellular IL-4 was detected. Cotransfer of $\beta_7^{+/+}$ or $\beta_7^{-/-}/CD4^{+}CD25^{+}$ Treg also led to a decreased percentage of LIEL that could be stimulated to produce TNF, but there was no decrease in the percentage of TNF producing LLPL or cells from MLN (data not shown). Since the absolute number of donor T cells in the large intestine was markedly decreased (Table I), however, the absolute number of TNF-producing cells in the intestine was correspondingly decreased.

### $\beta_7$ integrins are required for Treg activity in the intestine

The very low numbers of Treg in the intestine of protected RAG$^{-/-}$ recipient mice were not sufficient for purification and in vitro assay, although it remained possible that the few cells in the intestine were extremely potent. To address this issue, $\beta_7^{+/+}$ or $\beta_7^{-/-}$ Treg were cotransferred with CD4$^+CD45RB^{high}$ T cells into RAG$^{-/-}$ mice, and the combined total of CD4$^+LIEL$ and LLPL from the recipients were assessed for their ability to inhibit the proliferation of naive CD4$^+CD45RB^{high}\ cells\ from\ the\ spleen$ of wild-type mice (Fig. 4a). As total CD4$^+$ cells from the intestine of recipient mice were added to the naive spleen cells, the majority could not have been Treg. We reasoned, however, that if Treg acted locally to prevent inflammation, then in a 1/1 dilution in culture they still might be able to prevent proliferation in vitro. Intestinal T cells from mice cotransferred with $\beta_7^{+/+}$ Treg and CD4$^+CD45RB^{high}$ T cells prevented the in vitro proliferation by CFSE-labeled, naive spleen cells (Fig. 4b). In dramatic contrast, intestinal T cells obtained from mice that had been cotransferred with $\beta_7^{-/-}$ Treg could not inhibit naive T cell proliferation in vitro. These data indicate that although $\beta_7^{+/+}$ Treg in the intestine contributed to ex vivo regulatory function, this activity was not a prerequisite for protection from colitis in vivo.

![FIGURE 3. Intestinal IFN-$\gamma$ production is suppressed by $\beta_7^{-/-}$ Treg. LLPL and LIEL were isolated from RAG$^{-/-}$ recipients of CD45.1$^+$ CD4$^+CD45RB^{high}$ T cells alone or cotransfected with either $\beta_7^{+/+}$ or $\beta_7^{-/-}/CD4^{+}CD25^{+}$ Treg 10–12 wk after reconstitution. The cells were stimulated in vitro with anti-CD3e and analyzed for intracellular cytokine. Dot plots are gated on TCRB$^+$ CD45.1$^+$ cells and the numbers represent the percentage of positive cells in the indicated quadrants.](http://www.jimmunol.org/)
FIGURE 4. Regulatory activity cannot be detected in the intestine of recipients of β7−/− Treg. Total CD4+ T cells obtained from combined LIEL and LLPL were isolated from Rag−/− recipents of CD4+CD45RBhigh T cells in combination with either β7+/+ or β7−/− CD4+CD25+ Treg. These cells were cultured in vitro with CFSE-labeled CD4+CD45RBhigh T cells, anti-CD3ε, and Thy-1-depleted, irradiated splenocytes as diagrammed. Cultures were harvested after 48 h and proliferation was analyzed by flow cytometry.

Discussion
In this study, we provide evidence that β7−/− CD4+CD25+ Treg are impaired in their ability to populate the intestine during cotransfer with CD4+CD45RBhigh T cells into Rag−/− mice and therefore are greatly out numbered in the large intestine by progeny of the CD4+CD45RBhigh donor T cells. Despite this, they are capable of preventing intestinal inflammation and the associated Th1 polarization of pathogenic T cells in the intestine as Treg from wild-type mice. Therefore, while T cell pathogenesis in this colitis model is dependent on β7+ T cells, in spite of this, they are capable of preventing intestinal inflammation.

Alteration of the initial differentiation of naive T cells in the periphery may be a fundamental mechanism as to how Treg prevent inflammation in the transfer model of colitis (23). Consistent with this possibility, we showed that both β7−/− and β7+/+ CD4+CD25+ Treg migrated to the spleen and lymph nodes and expanded as efficiently there as CD4+CD45RBhigh T cells. In experimental autoimmune encephalomyelitis (24) and diabetes in mice (25), it also was shown that transferred, protective Treg accumulated primarily in the peripheral lymph nodes and failed to infiltrate the target organ.

Although the β7− independence of Treg action is proven, we cannot formally exclude the possibility that the very few β7+/+ Treg in the intestine contributed to the prevention of colitis. It is evident, however, that even with a 10-fold excess of CD4+CD45RBhigh T cells transferred, and impaired mucosal localization of Treg due to β7− deficiency, that Treg function was not markedly diminished. Furthermore, because no regulatory activity could be detected in the intestine of recipients that received β7−/− Treg, it is highly unlikely that the very few β7−/− Treg in the intestine were exceptionally potent suppressors.

The mixture of CD4+ T cells in the large intestine after cotransfer of wild-type Treg and CD4+CD45RBhigh T cells could prevent the proliferation of naive T cells in vitro. Our data therefore are consistent with the hypothesis that wild-type Treg can act in the intestine as well as in lymph nodes. It remains to be determined whether the local Treg stimulated some of the progeny of the CD4+CD45RBhigh T cells in the intestine to convert to Treg (26), although this did not happen when β7−/− Treg were transferred. It was reported recently that CD4+CD25+ Treg can treat established colitis, and it was suggested in that case the Treg were regulating pathogenesis in the intestine (27). It is conceivable that the major site of action of Treg will depend on the experimental model, with the extent of preexisting inflammation as one critical variable.

In a previous colitis study, it was found that cotransfer of equal numbers of Treg and CD4+CD45RBhigh T cells reduced the systemic expansion of the CD4+CD45RBhigh T cells more efficiently, compared with our data generated by a cotransfer of much higher ratios of CD4+CD45RBhigh T cells:Treg (7). This suggests that Treg are versatile suppressors, and the mode of action and extent of immune suppression caused by Treg transfer could be highly dependent upon the number of cells transferred. Our data indicate that the effects of either the β7+/+ or β7−/− Treg were concentrated in the intestine, where the total number of infiltrating CD4+CD45RBhigh T cells was reduced and the ability of the few cells in the intestine to produce Th1 cytokines diminished. These results indicate that in our experiments Treg acted by inhibiting the generation of tissue-seeking Th1 effector cells, which is sufficient to prevent disease in the transfer model of mouse colitis.

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Disclosures
The authors have no financial conflict of interest.

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