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J Immunol 2005; 174:533-541;
doi: 10.4049/jimmunol.174.1.533
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Genetic Interactions in *Eae2* Control Collagen-Induced Arthritis and the CD4\(^+\)/CD8\(^+\) T Cell Ratio\(^1\)

Jenny Karlsson,* Martina Johannesson,* Therese Lindvall,* Patrik Wernhoff,† Rikard Holmdahl,* and Åsa Andersson\(^2\)*

The *Eae2* locus on mouse chromosome 15 controls the development of experimental autoimmune encephalomyelitis (EAE); however, in this study we show that it also controls collagen-induced arthritis (CIA). To find the smallest disease-controlling locus/loci within *Eae2*, we have studied development of CIA in 676 mice from a partially advanced intercross. *Eae2* congenic mice were bred with mice congenic for the *Eae3/Cia5* locus on chromosome 3, previously shown to interact with *Eae2*. To create a large number of genetic recombinations within the congenic fragments, the offspring were intercrossed, and the eight subsequent generations were analyzed for CIA. We found that *Eae2* consists of four *Cia* subloci (*Cia26, Cia30, Cia31,* and *Cia32*), of which two interacted with each other, conferring severe CIA. Genes within the other two loci independently interacted with genes in *Eae3/Cia5*. Investigation of the CD4/CD8 T cell ratio in mice from the partially advanced intercross shows that this trait is linked to one of the *Eae2* subloci through interactions with *Eae3/Cia5*. Furthermore, the expression of CD86 on stimulated macrophages is linked to *Eae2*. *The Journal of Immunology*, 2005, 174: 533–541.

Chronic inflammatory diseases such as rheumatoid arthritis (RA)\(^1\) and multiple sclerosis (MS) are characterized by a complex etiology including both genetic and environmental factors (1). Studies of autoimmune diseases in human cohorts and experimental animal models have shown that autoimmune diseases are polygenic, and it is believed that small contributions from many genes determine the disease outcome. Due to the genetic heterogeneity in the human population and the relatively low sample number in the investigations, the identification of genes significantly linked to susceptibility to autoimmune disease in humans has been limited to date (2, 3). For this reason, animal models resembling human disease are very important tools for studying complex genetic traits.

RA affects the wrists and small joints of the hands and feet, leading to deformity and disability. Susceptibility is associated with genes in the MHC (4, 5). Collagen-induced arthritis (CIA), a frequently studied model for RA, can be induced in genetically susceptible mouse strains. Immunization with collagen type II (CII) results in polyarthritis, and the disease process is dependent on both cell-mediated and humoral immune mechanisms (6–11). Studies of CIA development, followed by genetic linkage analyses on gene-segregating crosses between susceptible and resistant MHC-congenic mouse strains, have revealed a large number of non-MHC CIA susceptibility loci (http://informatics.jax.org). The B10.RIII mouse strain (H-2\(^d\)) is susceptible to chronic arthritis upon immunization with bovine CII. In an F\(_2\) intercross between B10.RIII and the CIA-resistant MHC congenic strain RIIIS/J (H-2\(^b\)), two loci on chromosome 3 (Cia3) and chromosome 13 (Cia10) were significantly linked to the development of CIA (12). Cia5 overlaps with the *Eae3* locus, which was previously demonstrated to interact with chromosome 15 (*Eae2*) in controlling susceptibility to experimental autoimmune encephalomyelitis (EAE) (13). EAE is an experimental model of MS, an inflammatory demyelinating disease in the CNS. B10.RIII mice are susceptible to chronic EAE induced by immunization with the myelin basic protein (MBP) peptide 89–101, whereas the RIIIS/J mouse strain is resistant to disease induction. In addition to *Eae3*, development of EAE with this immunization protocol has been linked to several gene regions (13–15). The *Eae2* locus is located on the centromeric part of chromosome 15. In the previous F\(_2\) intercross between B10.RIII and RIIIS/J, the increased incidence of EAE was linked to an interaction between the *Eae2* and the *Eae3* quantitative trait loci (QTL). It was observed that mice heterozygous in the *Eae2* region and homozygous for B10.RIII in *Eae3* were the most affected (13). To confirm these data, *Eae2* congenic mice were produced and studied in an EAE experiment (14). These data showed a nonsignificant tendency toward increased severity in the *Eae2* homozygous mice (14).

The human diseases RA and MS as well as the corresponding animal models CIA and EAE are considered to be T cell-mediated diseases. Ag-specific Th1 cells, activated in the peripheral lymphoid organs, migrate into the target organ and become reactive by resident APCs (16–19). To make a proper Ag response, T cells need a costimulatory signal. The CD80 and CD86 costimulatory molecules expressed on different APCs interact with the CD28/CD152 molecules on T cells. The important role for this interaction in the induction of autoimmunity in CIA and EAE has been...
reported (20, 21). Although the main focus in studies of experimental models for autoimmune diseases has been on CD4+ T cells, it is suggested that the CD8+ T cells take part in the pathogenesis of EAE (22, 23). The specificities of the T cell repertoire in the periphery, including clones reactive toward autoantigens, are selected in the thymus. In addition, the relative numbers of CD4+ and CD8+ T lymphocytes in the lymphoid organs are genetically controlled and differ between mouse strains (15, 24). Comparison between the CIA/EAEx-resistant RIIIS/J and -susceptible B10.RIII strains with regard to the ratio between CD4+ and CD8+ T cells in the spleens of naïve animals reveals a higher CD4+/CD8+ T cell ratio in the RIIIS/J strain. This trait was mapped in a gene-segregating backcross between the two strains. The CD4+/CD8+ T cell ratio was linked to three genomic regions in this study, most interestingly to the Eae2 region on chromosome 15 (15).

In this study we report that the Eae2 congenic fragment harbors four sub-QTL (denoted Cia26, Cia30, Cia31, and Cia32) controlling CIA. The data are based on CIA experiments with mice derived from a partially advanced intercross (PAI) between Eae2 and Eae3/Cia5 bicongenic mice. The intercrossed bicongenic animals and single-congenic mice, altogether comprising 1051 animals, have been investigated for CIA development. We found that genes within the Eae2 region control CIA development in a complex interplay between genes inside the Eae2 fragment as well genes in the Eae3/Cia5 region on chromosome 3. Additionally, the CD4+/CD8+ T cell ratio in PBL was investigated and found to be controlled by interactions between genes within Eae2 on chromosome 15 and Eae3/Cia5 on chromosome 3. Moreover, based on a genetic linkage analysis on a backcross between the B10.RIII and RIIIS/J strains, we investigated the expression of costimulatory molecules on activated macrophages from Eae2 single-congenic mice, and we conclude that genes within the Eae2 locus, in addition, control CD86 expression.

Materials and Methods

Mice

B10.RIII mice were originally provided by I. Klein (Max-Planck-Institut für Biologie) and were kept in our breeding colony. RIIIS/J mice were purchased from The Jackson Laboratory. Eae2 and Eae3/Cia5 congenic mouse strains were bred in our colony at Lund University. RIIIS/J (donor) genes were bred onto the B10.RIII (recipient) background. The mice were backcrossed for 10 generations, and mice heterozygous in the Eae3 region on chromosome 3 and in the Eae2 region on chromosome 15, respectively, were used for breeding the next generation. Subsequently, the 106th generation was intercrossed to obtain mice homozygous for RIIIS/J genes in the respective congenic fragments. These mice have the genetic background from B10.RIII, except in the congenic fragments. Eae2/Eae3 bicongenic mice were intercrossed to produce F1 mice, heterozygous in both congenic loci. These mice were bred in a PAI by further intercrossing the mice for up to eight generations. In each generation the mice were genotyped and tested for CIA. In total, 1051 animals were analyzed for CIA. By association studies between genotype and disease outcome, the desired genetic regions for breeding the next generation were chosen. In this way, recombinations in each generation were collected.

(B10.RIII × RIIIS/J)F1 × B10.RIII animals were produced in our colony at the Section for Medical Inflammation Research, Lund University, and studied in an EAE experiment (15). Eighty animals were analyzed for staining at the Section for Medical Inflammation Research, Lund University, and studied in an EAE experiment (15). Eighty animals were analyzed for CIA and studied in an EAE experiment (15).

Disease induction and scoring

Bovine CII was prepared from calf nasal cartilage by pepsin digestion and was purified as previously described (25). CIA was induced in 10- to 16-wk-old mice by intradermal immunization with 100 μg of CII emulsified in IFA (Difco) in the base of the tail. The mice were boosted with 50 μg of bovine CII emulsified in IFA on day 35. The clinical arthritis severity was measured every second day by scoring the swelling and erythema in the joints of the paws. This scoring system is based on the number of inflamed joints, ranging from one to 15 for each affected paw, with a maximum score of 60/mouse. Each affected ankle/wrist was scored as 5, and each inflamed knuckle and toe was given 1 point.

Cell preparation, in vitro stimulation, and FACS analyses

Peritoneal cell exudates were prepared by injecting 10 ml of DMEM (Invitrogen Life Technologies) supplemented with 5% FCS into the peritoneal cavity and were subsequently dispensed into a polypropylene tube. The cells were washed and resuspended in 1 ml of DMEM supplemented with 10% FCS, 1% penicillin-streptomycin, 0.1 M HEPES, and 2-ME (cell culture medium). The cells were seeded (105 cells/well) in a 48-well plate. The macrophages and T cells were allowed to adhere for 30–60 min and then washed with DMEM to remove nonadherent cells before culture overnight in cell culture medium. Subsequently, the cells were washed and stimulated with IFN-γ (100 U/ml) and LPS (10 ng/ml) for 48 h. The stimulated peritoneal cells were removed from the cell culture plate by gently scraping off the cells. Subsequently, the cells were incubated with normal rat serum and supernatant from the 2-4G2 hybridoma (anti-CD16/CD32, our hybridoma collection) to reduce non-αg-specific binding of IgG to FcRs. Abs against CD11b (Mac-1) conjugated with FITC (clone M1/70) and CD86 (B7.2) conjugated with PE (clone GL1; BD Pharmaningen) were used for staining the peritoneal cells. Negative controls were stained with the anti-CD11b-FITC only. Cells expressing high levels of CD11b were electronically gated for analysis of CD86 expression.

RBC were lysed with 0.84% ammonium chloride, and the cells were washed with staining buffer (PBS, 3% FCS, and 0.01% NaN3) and incubated with supernatant from the 2.4G2 hybridoma. The cells were subsequently stained with anti-CD4-PE (clone H129.19) and anti-CD8-CyChrome (clone 53-6-7; BD Pharmaningen) and analyzed on a FACSscan (BD Biosciences).

Genotyping

DNA was prepared from toe or tail biopsies according to a standard protocol (26). Alternatively, tissues were dissolved in 300 μl of 50 mM NaOH for 1–2 h at 95°C, and subsequently neutralized with 100 μl of 1 M Tris buffer (pH 8). Seventeen and 18 informative fluorescence-labeled microsatellite markers (Interactiva and MWG Biotech) were used for genotyping the Eae2 and Eae3/Cia5 congenic fragments, respectively. PCRs and genotyping analyses were performed as previously described (15).

Statistics and linkage analysis

The significance value for interactions between phenotype and two markers was calculated by two-way ANOVA. The Mann-Whitney U test was used to calculate the significance between two groups. Two-locus interaction analyses were also performed using R (copyright 2004, The R Foundation for Statistical Computing, version 2.0.0) and R/qtl (27). The analysis was performed using regression of phenotypes on multiple genotype probabilities (Haley-Knott regression). The two-QTL model compares a full model in the presence of covariates (y = μ + β1q1 + β2q2 + β3q2 × q2 + Ay + Zβq1 + Zβq2 + Zβq1 × q2 + e) to a null model (y = μ + Ay + e), e.g., likelihood of odds (LOD) joint values, where q1 and q2 represent QTL genotypes at two different locations, A is a matrix of covariates, and Z is a matrix of QTL interacting covariates. For the interaction, epistasis LOD scores, the full model is compared with an additive model (y = μ + β1q1 + β2q2 + Ay + Zβq1 + Zβq2 + e), e.g., LODint values. The experimental significance thresholds for the two-locus interactions were subsequently determined using permutation tests (n = 10000), where p < 0.05 was considered significant. One-locus linkage analysis and permutation tests were conducted using the MQM Map and QTXT software (http://mquip.rutgers.edu/ or swparkwell.gene.com/qntQTL.html). The marker map was generated using the Kosambi map function. Permutation tests were performed to establish the empirical significance thresholds on a genome-wide level (p < 0.05) for the phenotype. Interval mapping was conducted at 1-cM increments under the additive regression model to calculate the test statistics (likelihood ratio statistic (LRS)).

Results

The Eae2 region confers sensitivity or resistance to CIA depending on the inheritance of RIIIS/J alleles

In contrast to the Eae3/Cia5 locus, the Eae2 QTL has not previously been linked to CIA in any cross. Because genes within the Eae3 and the Eae2 QTL were postulated to interact in EAE (13), we hypothesized that this might also be true for CIA. First, the Eae2 single-congenic strain was immunized for CIA to investigate whether the Eae2 fragment alone controls CIA when background genes are neutralized, as in a congenic strain. The Eae2 locus was...
not linked to CIA development in a previous experiment (12), which might have been due to background genes. In an altered genetic environment in which only the Eae2 region differs from B10.RIII wild-type (wt), genes within the Eae2 region might be unraveled. The disease phenotype area under the curve (AUC) was used to assess disease severity. The AUC is the sum of scores for each individual mouse during a defined test period. The scoring started on day 18, mice were then boosted on day 35, and the experiment was ended on day 92. In addition to the whole test period, days 18–92 (AUC d18–92), we separated the early phase of the test period, days 18–35 (AUC d18–35) from the acute phase, days 53–92 (AUC d53–92). Other disease traits analyzed were the mean day of onset and the incidence of disease along with the mean maximum score. As shown in Fig. 1, we could demonstrate that the Eae2 locus indeed controlled arthritis, because an exacerbated disease was observed when mice had one allele from each parental strain, and protection from disease was conferred when mice had two RIIIS/J alleles. These data are in line with the previous F₂ intercross (13) and support our hypothesis that the same genes operate in EAE and CIA. RIIIS/J homozygous alleles in the Eae2 fragment protected against disease development in both males and females (Fig. 1), but the exacerbating effect from heterozygous alleles was found only in female mice (Fig. 1A). The incidence of CIA in female Eae2 congenic mice with a heterozygous fragment was 88% compared with 47% in B10.RIII females and 41% in congenic females with a RIIIS/J homozygous Eae2 fragment.

**Dissection of the Eae2 congenic fragment in a PAI reveals four loci controlling different phases of arthritis**

From the finding that the Eae2 congenic fragment indeed controls arthritis, the next step was to pinpoint the QTL for the disease-causing genes. The aim was to produce mice with multiple recombination in the fragment, followed by investigation of CIA to define the genetic linkage region(s). By crossing the single Eae2 and Eae3 congenic strains (Fig. 2A) and by further intercrossing the bicongenic strain, we could investigate the previously postulated genetic interaction between the Eae2 and Eae3 regions (13) and, in addition, obtain recombinations in the fragments. The heterozygous B10.RIII/RIIIS/J Eae2 × Eae3 F₁ offspring were subsequently bred in a PAI for eight generations, and the progeny was investigated for CIA (Fig. 2A). In this way, only genes within the congenic fragments segregated, because the genetic background from B10.RIII was identical in these mice. Altogether 1051 congenic mice were investigated for CIA, including wt control mice.

Different phases of CIA were analyzed in the Eae2/Eae3 single- and bicongenic mice. In this experiment it became clear that different genes operate in the early phase, before boost, compared with the acute phase of the disease. These traits were differentially inherited and linked to different subloci within the congenic fragments (Table 1). We found that the development of CIA was linked to four regions within the Eae2 congenic region. We have denoted the novel CIA QTL within Eae2 on chromosome 15 as Cia26, 30, 31, and 32. The Eae3/Cia5 on chromosome 3 was similarly split into three sub-QTL (Cia5, Cia21, and Cia22), controlling different phases of CIA (44).

**Cia30 confers protection from CIA**

The Eae2 single-congenic mice with a homozygous RIIIS/J fragment developed CIA with lower severity than B10.RIII wt mice (Fig. 1). By separating the congenic fragments in the PAI, we could show that suppression of disease development originated from homozygous RIIIS/J alleles in Cia30. The protective effect from Cia30 was only observed when Cia5 on chromosome 3 had homozygous B10.RIII alleles, because one or two RIIIS/J alleles in Cia5 conferred protection from disease (Fig. 2B). Fig. 2C shows the development of disease in mice with different genotypes in Cia30, but homozygous for B10.RIII alleles in Cia5. Thus, in the PAI we could show that the gene(s) responsible for the development of milder CIA is located in the uppermost part of the original Eae2 congenic fragment, Cia30.

**Intrachromosomal interaction within the Eae2 fragment controls severe development of CIA**

Many genes that have relatively small effects on the disease phenotype control the development of autoimmune disease. Epistatic interactions between gene products are therefore believed to be an important feature for the development of a full-blown disease. Moreover, genes within the same biological pathway tend to be clustered in the genome (28–30). Because we had collected mice with many recombinations in the Eae2 region, we wanted to investigate the possibility for gene interactions between genes within
FIGURE 2. The protective effect of RIIIS/J homozygous alleles in the Eae2 fragment originates from the most centromeric region, Cia30. A. The Eae2 and Eae3 congenic fragments derive from the resistant RIIIS/J mouse strain and span ~40 Mbp/chromosome on the genetic background of the susceptible B10.RIII strain. The (B10.RIII × RIIIS/J)F1 offspring were backcrossed to B10.RIII. Mice heterozygous in the respective congenic region were chosen for additional backcrossing for 10 generations. Subsequently, the congenic mice were crossed, and the bicongenic mice were intercrossed for eight generations in a PAI. This breeding strategy resulted in genetic recombinations within the congenic fragments. The possible allele combinations for each given genetic location are depicted in the figure. The locations of the microsatellite markers are given according to the Celera mouse genome database. B. Investigation of CIA in the PAI mice revealed that the protective effect of RIIIS/J homozygous alleles (aa) in Cia30 was observed only when chromosome 3 (Cia5) was homozygous for B10.RIII (bb) alleles (marked with circle), aa, RIIIS/J homozygous alleles; ab, heterozygous alleles; bb, B10.RIII homozygous alleles. C. Disease curve for mice homozygous for B10.RIII (bb) in Cia5 and with different genotypes in Cia30. Mice homozygous for RIIIS/J (aa) in Cia30 were protected from disease in contrast to mice with one (p < 0.0001, by Mann-Whitney U test) or two (p = 0.0136, by Mann-Whitney U test) B10.RIII alleles at this locus.

the locus. Surprisingly, we found that RIIIS/J homozygous alleles (aa) in the telomeric region of the Eae2 congenic fragment, denoted Cia26 (D15Mit182), interacted with homozygous B10.RIII alleles (bb) in Cia30 to promote severe disease development and a higher incidence of CIA (Fig. 3, A and B, and Table I). The effect was most prominent in female mice, which had 90% incidence of disease (data not shown).

To further investigate the possibilities of intrachromosomal interactions, we used the R/qtl software to calculate and visualize the results. Fig. 3C demonstrates an interaction plot created from calculations in the R/qtl analysis program (27). The color diagram shows the significance values for the trait AUC (d18–92), where the colors indicate the significance of the interactions according to the scale. As shown in the figure, there are several intrachromosomal interactions between loci in Eae2. The Cia30 locus interacted with both Cia32 and Cia26. Additionally, an interaction between Cia31 and Cia26 was observed (Fig. 3C). Taken together, the Cia30 locus influenced the severity of CIA through interactions with genes on chromosome 3 and with genes located in the nearby region on chromosome 15.

Disease-promoting heterozygous alleles in Cia31 and Cia32

In female Eae2 heterozygous single-congenic mice, we observed an exacerbating effect on CIA development compared with that in wt mice (Fig. 1A). When investigating the heterozygous effect in the PAI, mice heterozygous in Cia31 (D15Mit21) and homozygous for B10.RIII alleles in Cia30 were backcrossed to B10.RIII (bb) (marked with circle). aa, RIIIS/J homozygous alleles; ab, heterozygous alleles; bb, B10.RIII homozygous alleles. C. Disease curve for mice homozygous for B10.RIII (bb) in Cia5 and with different genotypes in Cia30. Mice homozygous for RIIIS/J (aa) in Cia30 were protected from disease in contrast to mice with one (p < 0.0001, by Mann-Whitney U test) or two (p = 0.0136, by Mann-Whitney U test) B10.RIII alleles at this locus.

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Subphenotypes are important tools in the process of finding genes in Eae3/Cia5 controlled by genes in the Cia30 locus that interact with genes markers (D15Mit80 and D15Mit150) within the CD4/CD8 T cell ratio in spleen lymphocytes is linked to two linkages to the relative numbers of CD4

A

also in peripheral blood (Fig. 5

B

the previous study, we observed in the single-congenic mice that marker D3Mit103 within the region and the postulated interactions with

Cia31

and

Cia32

(loci. A summary of the novel QTL within the Eae2 region and the postulated interactions with Eae3/Cia5 controlling the development of CIA is shown in Fig. 4.

Proportions of CD4^+ and CD8^+ T cells in peripheral blood are controlled by genes in the Cia30 locus that interact with genes in Eae3/Cia5

Subphenotypes are important tools in the process of finding genes controlling complex traits. In a linkage study with 200 (B10.RII × RIIIS/J)F1 × B10.RII mice, we recently showed that the CD4/CD8 T cell ratio in spleen lymphocytes is linked to two markers (D15Mit80 and D15Mit150) within the Eae2 region. Linkages to the relative numbers of CD4^+ and CD8^+ T cells, respectively, was in the same study defined to the region of the marker D3Mit103 within the Eae3/Cia5 locus (15). In support of the previous study, we observed in the single-congenic mice that genes within the Eae2 indeed controlled the CD4/CD8 T cell ratio also in peripheral blood (Fig. 5A). RIIIS/J genes in Eae2 conferred an increased CD4/CD8 T cell ratio. Furthermore, by investigating this trait in 200 of the PAI mice of both sexes, we could pinpoint the CD4/CD8 ratio in peripheral T lymphocytes were reported (31). The loci have been denoted Trmq4 (T cell modifier locus) 1 and 2, and we denoted the locus on chromosome 15 Trmq4. The same markers (D15Mit80 and D15Mit150) as in the previous backcross experiment were linked to this trait (15). Furthermore, this locus was shown to interact with Cia5 (D3Mit187), such that homozygous RIIIS/J alleles in both Trmq4 (Cia30) and Cia5 (denoted Trmq5) restored the CD4/CD8 T cell ratio to the same level as in the RIIIS/J wt strain (Fig. 5B). The Trmq4 also interacted with a locus further telomeric in the Eae3/Cia5 fragment (D15Mit80*D3Mit103; p < 0.0001, by ANOVA). We denoted this locus Trmq6. In line with the data from the previously reported backcross between B10.RII and RIIIS/J (15), we found that the relative numbers of CD4^+ and CD8^+ T cells in PBL were strongly linked to D3Mit103 (Trmq6; D3Mit103; p < 0.0001, by ANOVA) in the PAI animals. The percentage of CD8^+ T cells in PBL was, in addition, strongly linked to Trmq4 (D15Mit150; p < 0.0001, by ANOVA). Fig. 5C briefly outlines a summary of the novel QTL and genetic interactions controlling T cell subpopulations.

Expression of CD86 on stimulated macrophages is linked to Eae2

It has been recently shown that subphenotypes, such as relative numbers of splenic lymphocyte populations, are genetically linked to disease loci (15, 24). To compare the macrophage compartment between B10.RII and RIIIS/J wt mice, we investigated macrophage expression of cell surface molecules ex vivo or upon stimulation in vitro. We did not observe a difference in the expression of MHC class I, MHC class II, or CD80 on peritoneal macrophages from B10.RII and RIIIS/J (data not shown). Similarly, the expression of CD86 ex vivo or after activation with IFN-γ and LPS for 24 h revealed the same expression levels in the two strains (data not shown). However, after 48 h of activation in vitro, macrophages from B10.RII mice continued to have high expression of CD86, whereas the expression on RIIIS/J macrophages was at levels comparable to nonstimulated cells (Fig. 6A). The difference in CD86 levels after activation was investigated in two strains (data not shown). However, after 48 h of activation in vitro, macrophages from B10.RII mice continued to have high expression of CD86, whereas the expression on RIIIS/J macrophages was at levels comparable to nonstimulated cells (Fig. 6A).

Table I. CIA loci on proximal mouse chromosome 15

<table>
<thead>
<tr>
<th>Phenotype</th>
<th>QTL</th>
<th>Phenotype Value</th>
<th>Linkage Markers (Mb)</th>
<th>p</th>
</tr>
</thead>
<tbody>
<tr>
<td>AUC (days 18–92)</td>
<td><strong>Cia30</strong></td>
<td>82 ± 15</td>
<td>D15Mit10 (7.5 Mb)</td>
<td>0.0002</td>
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<tr>
<td></td>
<td><strong>Cia5</strong></td>
<td>196 ± 20</td>
<td>D3Mit187 (84.3 Mb)</td>
<td>&lt;0.0001</td>
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<tr>
<td></td>
<td></td>
<td>130 ± 14</td>
<td>D15Mit10*D3Mit187</td>
<td>0.02</td>
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<tr>
<td>AUC (days 53–92)</td>
<td><strong>Cia26</strong></td>
<td>166 ± 27</td>
<td>D15Mit182 (42.1 Mb)</td>
<td>0.3</td>
</tr>
<tr>
<td></td>
<td><strong>Cia30</strong></td>
<td>64 ± 13</td>
<td>D15Mit11 (6.5 Mb)</td>
<td>0.4</td>
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<td></td>
<td></td>
<td>64 ± 7</td>
<td>D15Mit11*D15Mit182</td>
<td>0.0003</td>
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<tr>
<td>Incidence CIA</td>
<td><strong>Cia26</strong></td>
<td>79%</td>
<td>D15Mit182 (42.1 Mb)</td>
<td>0.04</td>
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<tr>
<td></td>
<td><strong>Cia30</strong></td>
<td>94%</td>
<td>D15Mit11 (6.5 Mb)</td>
<td>0.3</td>
</tr>
<tr>
<td></td>
<td></td>
<td>44%</td>
<td>D15Mit11*D15Mit182</td>
<td>0.04</td>
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<tr>
<td>AUC (days 18–35)</td>
<td><strong>Cia31</strong></td>
<td>4 ± 1</td>
<td>D15Mit21 (26.4 Mb)</td>
<td>0.0004</td>
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<tr>
<td></td>
<td><strong>Cia5</strong></td>
<td>21 ± 5</td>
<td>D3Mit187 (84.3 Mb)</td>
<td>&lt;0.0001</td>
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<td></td>
<td></td>
<td>5 ± 2</td>
<td>D15Mit21*D3Mit187</td>
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<tr>
<td>Day of onset</td>
<td><strong>Cia31</strong></td>
<td>77 ± 3</td>
<td>D15Mit21 (26.4 Mb)</td>
<td>0.0007</td>
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<tr>
<td></td>
<td><strong>Cia5</strong></td>
<td>60 ± 3</td>
<td>D3Mit40 (84.6 Mb)</td>
<td>&lt;0.0001</td>
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<td></td>
<td></td>
<td>72 ± 3</td>
<td>D15Mit21*D3Mit40</td>
<td>0.13</td>
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<tr>
<td>AUC (days 18–92)</td>
<td><strong>Cia32</strong></td>
<td>52 ± 13</td>
<td>D15Mit111 (29.6 Mb)</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td></td>
<td><strong>Cia32</strong></td>
<td>184 ± 25</td>
<td>D3Mit103 (107.7 Mb)</td>
<td>&lt;0.0001</td>
</tr>
<tr>
<td></td>
<td></td>
<td>51 ± 13</td>
<td>D15Mit111*D3Mit103</td>
<td>&lt;0.0001</td>
</tr>
</tbody>
</table>

* QTL on chromosome 15 in bold. Cia5 and Cia22 are located within the Eae3/Cia5 locus on chromosome 3.
* Phenotype value is the mean ± SE of the phenotype values for the respective genotypes.
* The Mb positions are according to the Celera mouse genome database.
* Mean AUC days 18–92 in the different subphenotypes.
* The genotype for the indicated QTL. aa = homozygous RIIIS/J alleles; ab = heterozygous alleles; bb = homozygous B10.RII alleles.
* Denotes an interaction between the indicated genetic markers calculated with two-way ANOVA statistical method.
* Female specific.
D17Mit20 (by Mann-Whitney U test, \( p = 0.0001 \); LRS, 14.3; LRS significance level, 12.6). The Eae2 region, peaking on D15Mit21 (in this study denoted Cia31) on chromosome 15 (by Mann-Whitney test, \( p = 0.0017 \); LRS, 10.9; LRS significance level, 12.1), was suggestively linked to this trait. To confirm the linkage to chromosome 15, we investigated the expression of CD86 on stimulated macrophages from Eae2 congenic and B10.RIII wt mice. The results showed a difference in CD86 expression level between mice with one or two RIIIS/J alleles in the Eae2 region compared with mice homozygous for B10.RIII genes (Fig. 6B) and confirmed that there is one or more genes in this region that control this trait. However, in the Eae2 congenic mice with one or two RIIIS/J alleles, the expression of the CD86 molecule was higher compared with the B10.RIII mice (Fig. 6B). This is in contrast to what was
observed in the parental RIHS/J strain (Fig. 6A) and in the backcross animals, where macrophages from the heterozygous mice had lower expression of the CD86 molecule (data not shown).

**Discussion**

From this and other studies one can conclude that the centromeric part of mouse chromosome 15 harbors several genes that control the development of inflammatory disease in different experimental models, such as CIA (Cia26 and Cia30–32) (in the present study), EAE (Eae2) (13, 15), proteoglycan-induced arthritis (Pgia8) (32), progression of autoimmune arthritis in MRL mice (Paam1) (33), *Borrelia burgdorferi*-associated arthritis (Bbaa14) (34), and Theiler's murine encephalomyelitis virus-induced demyelination (Tmevd8) (35). Moreover, the homologous region on human chromosome 5 has been associated with MS susceptibility (36). To date, >20 Cia loci have been reported, including the four novel loci defined in the present study. Similarly, >25 Eae loci have been described. Approximately half the QTL for EAE overlap with Cia loci or with loci linked to disease in other experimental models of arthritis (http://informatics.jax.org). It has been discussed that this would reflect that the same pathways are operating in different autoimmune diseases. Another point of view is that genes potentially involved in the development of autoimmunity are clustered in the genome, and this would explain linkages to the same genetic regions in different diseases (1, 37).

The two original QTL discussed in this paper, Eae2 and Eae3, were defined in a genetic linkage analysis on data from an EAE experiment with mice from an F2 intercross between the B10.RIII and RIHS/J mouse strains (13). Subsequently, Eae3 was also linked to the development of CIA in a cross between the two strains, and was denoted Cia5 (12). CIA development was not linked to the Eae2 region in the F2 intercross, but as demonstrated in the present study, CIA development is controlled by genes within the Eae2 locus in the context of an Eae2 congenic mouse strain. In addition, we have demonstrated that the original Eae2 fragment harbors genes that counteract each other. The fact that Eae2 was not linked to CIA development in the Jirholt et al. (12) cross could be explained by a low number of animals that did not allow the necessary recombinations needed to obtain the genetic context for these genes to operate in a disease-promoting way.
The linkage of the CD4 spleen to chromosomes 2, 6, and 15 (13) (15). There was no recently reported genetic linkage of the CD4 from studies of a backcross between B10.RIII and RIIIS/J, we investigated the expression of the costimulatory molecule CD86 on stimulated macrophages. A complete genomic screen, followed by linkage analysis showed linkage to chromosome 15 and genetics of CIA.

In the CIA experiments performed in this study, we observed an inhibitory effect on disease development from homozygous RIIIS/J genes in the uppermost QTL of the novel Cia locus on chromosome 15, Cia30. However, the incidence and severity could be restored to the wt levels in the presence of one or two B10.RIII alleles. Moreover, the promotion of disease in mice with one or two B10.RIII alleles in Cia30 was only observed when genes in Cia5 were B10.RIII homozygous. The presence of one or two RIIIS/J alleles in Cia5 promoted protection that was stronger than the disease-inducing genes in Cia30. This could be compared with the finding in the EAE experiment on the original F2 intercross, in which none of the mice with homozygous RIIIS/J genes in Eae2 developed disease (13). Interestingly, the protection from CIA in mice with homozygous RIIIS/J alleles in Cia30 and Cia5 was accompanied by a high CD4+/CD8- T cell ratio in peripheral blood. From studies of a backcross between B10.RIII and RIIIS/J, we recently reported genetic linkage of the CD4+/CD8- T cell ratio in spleen to chromosomes 2, 6, and 15 (Eae2) (15). There was no linkage of the CD4+/CD8- T cell ratio to the Eae3/Cia5 region; this could be explained by the fact that in the backcross there were no RIIIS/J homozygous alleles. In the backcross, however, the Eae3/Cia5 locus was linked to the relative numbers of CD4+ and CD8+ T cells, which now has been confirmed in the PAI animals as being confined to the same genetic marker (D3Mit103).

By splitting the Eae2 locus in the context of the PAI, an intrachromosomal interaction between Cia30 and Cia26 was revealed. As mentioned above, homozygous RIIIS/J alleles in the centromeric Cia30 locus suppressed disease development and overcame the disease-promoting genes in Cia26. However, in mice with B10.RIII homozygous genes in Cia30 and RIIIS/J homozygous genes in Cia26, the incidence and severity of CIA were very high. Statistical analyses suggested an interaction between the two loci to promote disease. A similar effect was reported in studies of the genetic contribution to susceptibility to autoimmune ovarian dysgenesis (38). By using recombinant congenic lines, it could be concluded that different alleles on the same chromosome influenced disease development in opposite directions. The clustering of genes within an original QTL, that in different ways contribute to disease development, has been shown in models for systemic lupus erythematosus and type I diabetes (28–30). This demonstrates that complex diseases are controlled by multiple genetic interactions that can only be dissected out by breeding recombinant congenic lines or advanced intercrosses with congenic lines.

The genetic linkage results from an EAE experiment on an F2 intercross between B10.RIII and RIIIS/J indicated an interactive effect of homozygous B10.RIII alleles on Eae3 and heterozygous alleles on Eae2 (13). Studies of Eae2 congenic mice could, however, not confirm that genes within this locus control EAE development (14). In the present study we observed that females with a heterozygous congenic Eae2 fragment had a higher incidence and severity of CIA compared with heterozygous males as well as with female B10.RIII wt mice. In the PAI, we reproduced the disease severity phenotype in females and located the genetic linkage to the Eae2 sub-QTL Cia32. Furthermore, we demonstrated that B10.RIII homozygous alleles on chromosome 3 interacted with heterozygous alleles in Cia32. Thus, we have confirmed the confidence interval for the disease-exacerbating gene within the Eae2 region on chromosome 15 from ~40 Mbp down to 7 Mbp.

The Cia31 locus in the heterozygote state promoted significantly greater severity in the early phase of disease along with early onset of disease. Statistical calculations showed that this trait was also the result of a true interaction between Cia31 and a locus on chromosome 3 (D15Mit21*D3Mit187), thus confirming the Sundvall data (13), in addition to supporting the hypothesis that common genes control different inflammatory disorders. Interestingly, the ank locus, encoding for a trans-membrane protein that controls pyrophosphate levels in cells, is located in the Cia31 region. Mutations in the ank gene result in unregulated calcification, leading to inflammation and arthritis (39).

During the initiation of EAE, CD4+ T cells infiltrate the CNS accompanied by macrophages. It was recently demonstrated that the invading macrophages express MHC class II and B7 molecules, a prerequisite to function as APCs (40). The role for the CD86 molecule in the development of EAE, however, has revealed somewhat contradictory results (41–43). In mice from a backcross between B10.RIII and RIIIS/J, we investigated the expression of the costimulatory molecule CD86 on stimulated macrophages. A complete genomic screen, followed by linkage analysis showed linkage to chromosome 17 in male mice and to the Cia31 region on chromosome 15 in all mice. In line with the RIIIS/J wt mice, the backcross mice heterozygous in the Cia31 region down-regulated CD86 on 48-h stimulated macrophages, indicating that the genes were dominantly inherited. In Eae2 congenic mice, it was confirmed that genes in this locus control the expression of CD86. The inheritance was, however, opposite to the backcross and wt data. It is likely that the different inheritance is due to genetic interactions.
within Eae2, in line with the complex genetic interactions controlling CIA that were observed in this study. It can be speculated that the Cia30-Cia26 genetic interactions or the effect of Cia31 also control this trait, but this has to be further investigated in mice with smaller congenic fragments.

In this study we have elucidated a complex genetic interplay between disease loci on two different chromosomes as well as between loci closely located within one chromosome. It can be assumed that additional loci on other chromosomes will also contribute to the disease trait through interactions with the disease loci on chromosome 15. However, by combining genetic regions on chromosome 15 and 3 to obtain the strongest phenotype, it will be possible to further limit the QTL and eventually come closer to the responsible genes.

Acknowledgments

We thank M. Neptin and H. Malm for technical assistance, C. Palestros and I. Bohlin for help with animal care, and A. Bäcklund for comments and linguistic corrections.

References