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Linker for Activation of T Cells Integrates Positive and Negative Signaling in Mast Cells¹

Odile Malbec,∗† Marie Malissen,† Isabelle Isnardi,∗‡ Renaud Lesourne,∗‡ Anne-Marie Mura,‡ Wolf H. Fridman,∗ Bernard Malissen,‡ and Marc Daëron∗†

The transmembrane adapter linker for activation of T cells (LAT) is thought to couple immunoreceptors to intracellular signaling pathways. In mice, its intracytoplasmic domain contains nine tyrosines which, when phosphorylated upon receptor aggregation, recruit Src-homology 2-domain-containing cytosolic enzymes and adapters. The four distal tyrosines are critical for both TCR and FcεRI signaling. Unexpectedly, knock-in mice expressing LAT with a point mutation of the first or of the last three of these tyrosines exhibited an abnormal T cell development characterized by a massive expansion of TH2-like αβ or γδ T cells, respectively. This phenotype suggests that, besides positive signals, LAT might support negative signals that normally regulate terminal T cell differentiation and proliferation. We investigated here whether LAT might similarly regulate mast cell activation, by generating not only positive but also negative signals, following FcR engagement. To this end, we examined IgE- and/or IgG-induced secretory and intracellular responses of mast cells derived from knock-in mice expressing LAT with combinations of tyrosine mutations (Y136F, Y(175, 195, 235)F, or Y(136, 175, 195, 235)F). A systematic comparison of pairs of mutants enabled us to dissect the respective roles played by the five proximal and the four distal tyrosines. We found that LAT tyrosines differentially contribute to exocytosis and cytokine secretion and differentially regulate biological responses of mucosal- and serosal-type mast cells. We also found that, indeed, both positive and negative signals may emanate from distinct tyrosines in LAT, whose integration modulates mast cell secretory responses. The Journal of Immunology, 2004, 173: 5086–5094.

Intracellular signaling complexes that are generated when immunoreceptors are engaged at the cell surface by multivalent extracellular ligands build up around tyrosine-rich transmembrane adapters (1, 2). Among these, the linker for activation of T cells (LAT) (3, 4) has been shown to play critical roles for TCR signaling. TCR aggregation triggered reduced Ca²⁺ mobilization, MAPK activation, and NF-AT activation in LAT-deficient T cells (5, 6). Likewise, high affinity IgE receptor (FcεRI) signaling was markedly impaired in LAT-deficient mast cells (7). These observations suggested that LAT serves as a coupling molecule between immunoreceptors and intracellular signaling pathways leading to cellular responses.

LAT is a single-chain membrane molecule located in lipid microdomains (8). It has a short extracellular domain and a long intracytoplasmic domain which contains 10 tyrosine residues in humans and 9 in mice (3, 4). Following TCR engagement, LAT is tyrosyl-phosphorylated by ZAP70 (9) that is recruited to phosphorylated ITAMs and activated (9). Tyrosyl-phosphorylated LAT thus provides multiple docking sites for a variety of Src-homology 2 (SH2) domain-containing cytosolic enzymes and adapters. These include phospholipase C-γ (PLC-γ), protein tyrosine kinases of the Tec family, the p85 subunit of PI3K, the exchange factor Vav and the adapters Gads, Grap, and Grb2 (3, 4, 10). Previous works based on mutational analysis of LAT identified critical tyrosine residues involved in the recruitment of these molecules in T cells (10, 11). These were the four distal tyrosines (Y132, Y171, Y191, and Y226) (SH2) domain-containing leukocyte protein of 76 kDa and of PLC-γ in humans, and their homologues in mice Y136, Y175, Y195, and Y235). Specifically, Y132/Y136 was demonstrated as being the major binding site for PLC-γ and the three distal tyrosines (Y171/Y175, Y191/Y195, and Y226/Y235) binding sites for Gads, Grap, and Grb2 (10). The two sets of binding sites also contribute indirectly to the recruitment of other molecules such as SH2-containing leukocyte protein of 76 kDa via Gads and they cooperate to stabilize the binding of molecules recruited by each other. The mechanisms by which LAT controls FcεRI signaling are thought to be similar to the LAT-dependent mechanisms that control TCR signaling. FcεRI aggregation in bone marrow-derived mast cells (BMMCs) from LAT−/− mice triggered a reduced phosphorylation of SH2-containing leukocyte protein of 76 kDa and of PLC-γ, resulting in decreased Ca²⁺ mobilization and MAPK activation and, ultimately, in a decreased release of preformed mediators and secretion of cytokines (7). A mutational analysis of the four distal tyrosines of LAT was recently performed in LAT−/− BMMCs reconstituted in vitro with wild-type (wt) or mutant LAT (12). This study confirmed that these residues are phosphorylated upon FcεRI

⁴ Abbreviations used in this paper: LAT, linker of activation of T cells; SH2, Src-homology 2; PLC-γ, phospholipase C-γ; BMMC, bone marrow-derived mast cell; wt, wild type; PCMC, peritoneal cell-derived mast cell; GAM, goat anti-mouse Ig; GAR, goat anti-rabbit Ig; [Ca²⁺]i, intracellular free calcium concentration; SCF, stem cell factor; SHP, SH2 domain-containing phosphatase.

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engagement and play a critical role for FcεRI signaling by recruiting the same set of molecules in mast cells as in T cells.

Recently, we (13) and others (14) used a knock-in strategy to evaluate the in vivo relevance of these data. Mice bearing a single Y136F mutation had an unexpected aberrant T cell development characterized by an exaggerated polyclonal differentiation of TCRαβ T cells into CD4 cells that secreted abnormally high levels of TH2 cytokines. As a consequence, serum IgG1 concentrations were 100-fold higher than in wt mice, serum IgE concentrations were in the range of milligrams per milliliter and peripheral tissues were massively infiltrated with eosinophils (13, 14). Likewise, mice bearing Y175F, Y195F, and Y235F mutations had an abnormal differentiation of TCRγδ T cells, also resulting in an exaggerated TH2 polarization and massive proliferation (15). Although they affect two distinct T cell lineages, respectively, the two types of LAT tyrosine mutations therefore seemed to inhibit a negative regulation that normally controls terminal T cell differentiation in both lineages of T cells. This suggests that LAT may contribute not only to positive signaling, but also to negative signaling.

The possibility that LAT might support inhibitory signals is not easily explained by the known functions of the four distal tyrosines whose mutation was responsible for the severe T cell abnormalities observed in knock-in mice. In an attempt to investigate this possibility, we undertook a functional analysis of FcR signaling in mast cells derived from the same knock-in mouse in which LAT had Y to F mutations of one (Y136), three (Y175, Y195 and Y235), or all four distal tyrosines and, as positive and negative controls, in mast cells from wt and LAT Y/− mice. Our rationale was that, not being known to depend on immunoreceptor signaling, mast cell development would not be affected by LAT mutations as T cell development was. We examined IgE- and/or IgG-induced responses in two types of mast cells: BMMCs, which are considered a model of mucosal-type mast cells, and peritoneal cell-derived mast cells (PCMCs), a novel type of cultured mast cells that we recently available model of serosal-type mast cells (O. Malbec et al., manuscript in preparation). We found that LAT differentially regulates the biological responses of mucosal- and serosal-type mast cells, and that LAT tyrosines differentially contribute to exocytosis, cytokine secretion, and intracellular signals. We also found that, in both cell types, LAT mutants could indeed generate not only positive but also negative effects that could be assigned to distinct tyrosine residues. It follows that, besides coupling FcRs to intracellular signaling pathways, LAT supports both positive and negative signals, whose integration by wt LAT results in a dominant positive effect, and that differentially regulate the biological responses triggered by FcRs in two types of mast cells.

Materials and Methods

Mice expressing mutant LAT

For the sake of clarity, we will adopt here a nomenclature showing which residues, among the four distal tyrosines of LAT, were (F) and were not (Y) mutated. Thus, LAT-YYYY will stand for wt LAT, LAT-FYYY for LAT with a single mutation (Y136F), LAT-YFFF for LAT with three mutations (Y175F, Y195F, and Y235F), and LAT-FFFF for LAT with four mutations (Y136F, Y175F, Y195F, and Y235F). Mice expressing LAT-FYYY have been described (13). To generate LAT-deleted neoR gene. Targeted embryonic stem cells were engineered in parallel (15). Mice were housed under specific pathogen-free conditions in accordance with institutional guidelines.

Abs and Ags

The mouse IgE anti-DNP mAb 2682-I was used as culture supernatant of a subclone of DNP-H1-e(26) hybridoma cells (18). BSA (Sigma-Aldrich, St. Louis, MO) was dinitrophenylated using dinitrobenzene sulfonic acid. Conjugates used for stimulation had a substitution number of 24 moles of DNP per mole of BSA. Mouse serum anti-GST was obtained by immunizing BALB/c mice i.p. with GST in Freund’s adjuvant. GST was purified from bacteria expressing a GST-containing pGEX vector. The rat anti-mouse FcγRIIBIIA mAb 2A4G2 (19) was purified by affinity chromatography on protein G-Sepharose (Pharmacia-ASAS, Guyancourt, France) from ascitic fluid of nude mice inoculated with 2.4G2 hybridoma cells i.p. Anti-LAT Abs were purchased from Upstate Cell Signaling Solutions (Euromex, Mundolsheim, France), FITC-conjugated Fab(1/2) of mouse anti-rat Ig and of goat anti-mouse Ig (GAM) from Jackson ImmunoResearch Laboratory (West Grove, PA), HRP-conjugated goat anti-rabbit Ig (GAR) and GAM Abs from Santa Cruz Biotechnology (Santa Cruz, CA), rabbit anti-phospho-ERK and anti-ERK Abs from New England Biolabs (Beverly, MA). The anti-phosphotyrosine mAb 4G10 was a kind gift of Dr. S. Latour (Institut National de la Santé et de la Recherche Médicale (INSERM) Unité 429, Hôpital Necker, Paris, France).

Mast cells

BMMCs were generated from bone marrow cells and propagated as described previously (20) in 4% culture supernatant from X63 cells transfected with the cDNA encoding murine IL-3. PCMCs were obtained by culturing peritoneal cells with 4% culture supernatant from Chinese hamster ovary cells transfected with the cDNA encoding murine stem cell factor (SCF). IL-3- and SCF-secreting transfectants were kind gifts of Dr. P. D. S. Benz (NIBSC UI, Bethesda, MD, and INSERM Unité 139, Institut Paoli-Calmettes, Marseille, France). Homogeneous populations of FcεRI+ Kit+ BMMCs and PCMCs were obtained after 3 and 2 wk of culture, respectively. PCMCs exhibited the typical morphology and staining properties of peritoneal mast cells. Compared with BMMCs, they had the features of mature serosal-type mast cells (based on granule numbers and morphology, mediators, and murine MCP contents). Four- to 5-wk-old cultures were used for experiments. They were set up with bone marrow or peritoneal cells harvested from the same two mice. Comparable results were obtained in experiments performed with cells from two independent sets of cultures.

Immunofluorescence analysis

Cells were incubated at 0°C with undiluted 2682-I culture supernatant, 10 μg/ml 2.4G2, or medium alone, washed and stained with 50 μg/ml FITC-GAM Abs from Santa Cruz Biotechnology (Santa Cruz, CA), rabbit anti-phospho-ERK and anti-ERK Abs from New England Biolabs (Beverly, MA). The anti-phosphotyrosine mAb 4G10 was a kind gift of Dr. S. Latour (Institut National de la Santé et de la Recherche Médicale (INSERM) Unité 429, Hôpital Necker, Paris, France).

β-Hexosaminidase release

BMMCs and PCMCs were sensitized for 1 h at 37°C with the indicated dilutions of 2682-I hybridoma supernatant and washed. IgE-sensitized cells were challenged with the indicated concentrations of DNP-BSA for 15 min at 37°C. Nonsensitized PCMCs were challenged with IgE for 15 min at 37°C with preformed immune complexes made of the indicated dilutions of anti-GST and GST. Reactions were stopped by placing plates on ice and supernatants were collected. β-Hexosaminidase release was measured by incubating supernatants with p-nitrophenyl-N-acetyl-d-glucosaminide (a β-hexosaminidase substrate) (Sigma-Aldrich) for 2 h at 37°C. Reactions were stopped with glycine 0.2 M, pH 10.7, and absorbance was measured at 405 nm. The percentages of β-hexosaminidase released in supernatants were calculated using as 100% β-hexosaminidase contained in aliquots of cells lysed in 0.1% Triton X-100.

Secretion of TNF-α

BMMCs, previously sensitized by a 1-h incubation at 37°C with IgE, were incubated for 3 h at 37°C with DNP-BSA. Nonsensitized PCMCs were stimulated under the same conditions with GST-anti-GST preformed immune complexes. Serial dilutions of cell-free supernatants were harvested and assayed for TNF-α. TNF-α was titrated by a cytotoxic assay on L929 cells as described (21).

Quantitative analysis of cytokine transcripts by RNAse protection assay

BMMCs sensitized with IgE anti-DNP 2682-I were challenged for 3 h at 37°C with 0.1 μg/ml DNP-BSA or medium alone. Total RNA was extracted using RNA-Plus (Qi-biogene, Illkirch, France) and analyzed for multiplex transcript by RNAse protection assay using the MCK-1 RibOQuant
custom mouse template set (BD Pharmingen, Erembodegem, Belgium). 32P-labeled riboprobes were mixed with 10 μg of RNA, incubated at 56°C for 12–16 h, and treated with a mixture of RNase A and T1 and proteinase K. Protected 32P-labeled RNA fragments were fractionated on denaturing polyacrylamide gels and autoradiographed. The intensity of the bands was quantitated using a Fuji imaging system (St. Quentin-en-Yvelines, France).

Measurement of Ca2+ mobilization

Intracellular free calcium concentration ([Ca2+]i) was determined by preloading IgE-sensitized BMMCs with 5 mM Fluo-3 AM (Molecular Probes, Eugene, OR) in the presence of 0.2% Pluronic F-127 (Sigma-Aldrich) for 30 min at room temperature. Cells were washed three times in RPMI 1640, resuspended at 1 × 106 cells/ml in complete medium, and [Ca2+]i was monitored with a flow cytometer. After 3 min at 37°C, cells were stimulated with 0.1 μg/ml DNP-BSA, and [Ca2+]i was measured. The mean [Ca2+]i was evaluated with the software FCS assistant 1.2.9 beta (BD Biosciences).

Analysis of LAT phosphorylation

Cells sensitized with IgE 2682-I and stimulated at 37°C for 1 min with 0.1 μg/ml DNP-BSA or with medium alone were lysed in buffer containing 50 mM Tris-HCl, pH 8.0, 0.5% Triton X-100, 1 mM Na3VO4, 10 μg/ml aprotinin, 10 μg/ml leupeptin, and 1 mM PMSF. Postnuclear lysates were immunoprecipitated with protein A-Sepharose beads coated with rabbit polyclonal anti-LAT Abs. Immunoadsorbents were washed in lysis buffer and boiled in sample buffer. Eluted material was fractionated by SDS-PAGE and transferred onto Immobilon-P (Millipore, Bedford, MA). Membranes were saturated with 5% BSA diluted in Western buffer containing 150 mM NaCl, 10 mM Tris, and 0.5% Tween 20 (Merck, Schuchardt, Germany), pH 7.4. Membranes were Western blotted with anti-phosphotyrosine Abs 4G10 or with anti-LAT Abs and HRP-conjugated GAM or GAR, respectively. Labeled Abs were detected using an ECL kit (Amersham, Freiburg, Germany).

Western blot analysis of ERK1/2 phosphorylation and of LAT expression

BMMCs sensitized with IgE 2682-I were stimulated at 37°C for indicated periods of time with DNP-BSA. Nonsensitized PCMCs were stimulated under the same conditions with preformed GST-anti-GST complexes. Cells were lysed by three cycles of incubation for 1 min in liquid nitrogen followed by 1 min at 37°C in lysis buffer containing 50 mM Tris, pH 8, 150 mM NaCl, 1% Nonidet P40, 1 mM Na3VO4, 5 mM NaF, 10 μg/ml aprotinin, 10 μg/ml leupeptin, 10 μg/ml pepstatin, and 1 mM PMSF. Lysates were centrifuged at 12,000 rpm for 15 min at 4°C. Proteins were quantitated using the Bio-Rad protein assay (Hercules, CA). Ten micrograms of proteins were electrophoresed and transferred onto Immobilon-P. Membranes were saturated with 5% skimmed milk (Régilait, Saint-Martin-Belle-Roche, France) diluted in Western buffer, and Western blotted with anti-ERK1/2 and with anti-phospho-ERK1/2 Abs followed by HRP-GAR. LAT expression was assessed by Western blotting with anti-LAT Abs on 10 μg of proteins from whole cell lysates of 1 × 106 cells that were lysed in the same buffer.

Results

LAT tyrosine residues differentially contribute to IgE-induced exocytosis and cytokine secretion in BMMCs

Five populations of BMMCs were generated simultaneously from the bone marrow of age-matched wt and mutant mice with the same genetic background. These were wt and LAT−/− mice, on...
the one hand, and, on the other hand, knock-in mice in which LAT had a Y to F mutation at one (Y136) (LAT-FYYY), three (Y175, Y195 and Y235) (LAT-YFFF), or four (Y136, Y175, Y195 and Y235) (LAT-FFFF) positions corresponding to the four distal tyrosines that were previously reported to account for most LAT activities. Resulting cells expressed comparable levels of high affinity IgE receptors (FceRI) (Fig. 1A) and Kit, the SCF receptor (not shown). BMMCs generated from wt mice or mice bearing LAT mutations contained comparable amounts of LAT whereas, as expected, no LAT was detected in BMMCs derived from LAT-deficient mice (Fig. 1B).

LAT was inducibly tyrosyl-phosphorylated upon stimulation of IgE-sensitized cells with Ag. The magnitude of LAT phosphorylation seemed higher in wt BMMCs than in mutant BMMCs. In the latter, LAT phosphorylation was not markedly affected by the type of mutation. Noticeably, LAT-FFFF was also inducibly phosphorylated, indicating that the five proximal tyrosines could be phosphorylated upon FcεRI aggregation (Fig. 1C).

The release of β-hexosaminidase was monitored in BMMCs sensitized with different concentrations of IgE anti-DNP and challenged with different concentrations of DNP-BSA (Fig. 1D). As shown previously (7), measurable amounts of β-hexosaminidase were still released by LAT−/− BMMCs. LAT−/− BMMCs, however, released much less β-hexosaminidase than BMMCs expressing wt LAT. LAT-FFFF BMMCs also released less β-hexosaminidase than wt BMMCs. This reduction was not seen upon challenge with an optimal concentration of Ag, but it was obvious for supraoptimal Ag concentrations. By contrast, LAT-FYYY and LAT-YFFF BMMCs responded as poorly as LAT−/− BMMCs. Surprisingly, LAT-FFFF BMMCs released more β-hexosaminidase than LAT-YFFF or LAT-FYYY BMMCs.

The secretion of TNF-α was examined following stimulation of BMMCs with IgE and Ag. This was done by measuring the cytoxicity of serial dilutions of mast cell supernatants on L929 cells (Fig. 2A). This cytotoxic effect was previously shown to depend on TNF-α (21). The effect of LAT mutations on TNF-α secretion can be best appreciated by comparing the 60% cytotoxicity titers of supernatants deduced from dilution curves for each type of BMMC (Fig. 2B). A robust secretion of TNF-α was induced in IgE-sensitized wt BMMCs over a 3-log10 range of Ag concentrations. As previously reported, LAT−/− BMMCs still secreted TNF-α (7). However, they secreted lower amounts of TNF-α than wt BMMCs. The difference was observed at all Ag concentrations but it was more marked at high Ag concentrations. LAT-FFFF BMMCs secreted higher amounts of TNF-α than LAT−/− BMMCs, and LAT-FYYY and LAT-YFFF BMMCs secreted higher amounts of TNF-α than LAT-FFFF BMMCs. Surprisingly, the lowest response was observed in LAT-YFFF BMMCs.

Up-regulation of cytokine transcripts was examined using a RNase protection assay (Fig. 2C). The basal transcription of TGF-β1 observed in all five BMMCs was not up-regulated upon

![FIGURE 2. IgE-induced TNF-α secretion and up-regulation of cytokine transcripts in BMMCs from wt and LAT mutant mice. A, TNF-α secretion. BMMCs sensitized with mouse mAb IgE anti-DNP were challenged with the indicated concentrations of DNP-BSA for 3 h. TNF-α secreted in the supernatants was titrated by a bioassay on the TNF-α-sensitive L929 cells. Curves represent the percentage of cytotoxicity as a function of the dilution of BMMC supernatant. B, Supernatant dilutions that induced 60% cytotoxicity were plotted as a function of the concentration of DNP-BSA used for challenge. C, Up-regulation of cytokine transcripts. BMMCs sensitized with mouse mAb IgE anti-DNP were challenged with medium or DNP-BSA for 1 h. Cells were washed, lysed, and RNA was extracted. Cytokine transcripts were detected in 32P-labeled RNA by a RNase protection assay followed by gel fractionation and autoradiography. Two experiments performed with the same RNA extracts are shown. 1: LAT-YYYY; 2: LAT−/−; 3: LAT-FYYY; 4: LAT-YFFF; 5: LAT-FFFF.](http://www.jimmunol.org/doi/abs/10.4049/jimmunol.5089)
FcεRI aggregation. No significant basal transcription of other cytokine genes was observed in resting BMMCs. An IgE-induced up-regulation of IL-6, IL-13, and TNF-α transcripts was observed in all BMMCs but it was of a lower magnitude in BMMCs from mutant mice. Each LAT mutation affected similarly the up-regulation of cytokine transcripts. Compared with wt BMMCs, it was 20 times lower in LAT<sup>−/−</sup>, 9 times lower in LAT-FFFF and LAT-YFFF BMMCs, and 2.3 times lower in LAT-FYYY BMMCs for all three cytokines.

Taken together, the results described in this section indicate that although it is dispensable for exocytosis and cytokine secretion, LAT is required for these two responses to be optimal. This effect of LAT depends on its four distal tyrosines, but also on sequences containing the five proximal tyrosines. Among the four distal tyrosines, Y136 and the three distal tyrosines contributed differentially to exocytosis and cytokine secretion. Surprisingly, β-hexosaminidase release and TNF-α secretion were decreased when Y136 was present in the absence of the three distal tyrosines. β-hexosaminidase release was also decreased when the three distal tyrosines were present in the absence of Y136.

LAT tyrosine residues differentially contribute to IgE-induced intracellular signaling in BMMCs

We examined the intracellular concentration of Ca<sup>2+</sup> and the phosphorylation of the MAPKs ERK1/2, the effector molecules of two major intracellular signaling pathways known to be triggered by FcεRI in mast cells.

The fast and sustained increase in the intracellular Ca<sup>2+</sup> concentration observed upon challenge of IgE-sensitized cells with Ag in wt BMMCs was markedly reduced in LAT<sup>−/−</sup> BMMCs (Fig. 3A). As previously reported, some Ca<sup>2+</sup> mobilization was however still detectably induced in the absence of LAT. Ca<sup>2+</sup> mobilization observed in BMMCs expressing a LAT mutant was in all cases lower than Ca<sup>2+</sup> mobilization observed in wt BMMCs and higher than Ca<sup>2+</sup> mobilization observed in LAT<sup>−/−</sup> BMMCs. LAT-YYYF was more efficient than LAT-FYYY which was slightly more efficient than LAT-FFFF.

IgE-induced ERK1/2 phosphorylation was monitored at different times following challenge with Ag (Fig. 3B). An inducible phosphorylation of comparable magnitude was observed in all BMMCs at 3 min. ERK1/2 phosphorylation was slightly reduced at 10 min in LAT-YYYF BMMCs whereas it was not detectable any more in LAT<sup>−/−</sup> BMMCs. At 10 min, ERK1/2 phosphorylation was similarly decreased in LAT-FFFF and in LAT-FYYY BMMCs. It was even more reduced in LAT-FFFF BMMCs.

Taken together, the data described in this section indicate that LAT is dispensable for both the Ca<sup>2+</sup> response and activation of the MAPKs ERK1/2. LAT nevertheless contributes to the Ca<sup>2+</sup> response and is required for a sustained phosphorylation of ERK1/2. Although both Y136 and the three distal tyrosines are involved in both responses, Y136 is more critical than the three distal tyrosines for the Ca<sup>2+</sup> response, whereas the three distal tyrosines are more critical than Y136 for ERK1/2 phosphorylation.

LAT tyrosine residues differentially contribute to the activation of mucosal-type and serosal-type mast cells

Five populations of PCMCs were expanded simultaneously from peritoneal cells collected from the same mice that were used to generate BMMCs, and propagated in parallel cultures in SCF-containing medium. PCMCs from wt mice or from mice bearing LAT mutants contained LAT, but not PCMCs from LAT-deficient mice (Fig. 4A). The lower amount of LAT seen in LAT-FFFF cells was comparable to the amount of LAT seen in LAT<sup>−/−</sup> cells.
not reproducibly observed. All PCMCs expressed comparable levels of FcεRI and of FcγRIIB/IIIA (Fig. 4B). FcεRI-dependent responses were examined in PCMCs sensitized with IgE and challenged with Ag. FcγRIIIA-dependent responses were examined in nonsensitized PCMCs challenged with preformed IgG immune complexes. Indeed, among other differences with BMMCs, peritoneal mast cells can be activated not only by IgE and Ag via FcεRI but also by IgG immune complexes via FcγRIIIA (22).

IgE-induced β-hexosaminidase release observed in wt PCMCs was more severely impaired in LAT−/− PCMCs than in LAT−/− BMMCs (Fig. 4C). It was partially restored in LAT-FFFF PCMCs. Noticeably, β-hexosaminidase release was comparable or lower in LAT-FYYY PCMCs than in LAT-FFFF PCMCs, but higher in LAT-YFFF PCMCs. The same hierarchy of responses was observed when examining IgG immune complex-induced β-hexosaminidase release by PCMCs (Fig. 5A). β-Hexosaminidase release observed in wt PCMCs was abrogated in LAT−/− PCMCs. It was of a similar or higher intensity in LAT-YFFF PCMCs and in wt PCMCs. It was markedly inhibited in LAT-FFFF and virtually abolished in LAT-FYYY PCMCs.

We found that PCMCs secreted much lower amounts of TNF-α than BMMCs in response to IgE and Ag or to IgG immune complexes. For this reason, no cytotoxicity titer could be calculated. However, TNF-α secretion varied like β-hexosaminidase release.

TNF-α secretion was of a comparable or of a higher intensity in LAT-YFFF PCMCs than in wt PCMCs and it was reduced to the same extent in LAT-FYYY and in LAT−/− PCMCs. Comparable results were observed in PCMCs stimulated with IgG immune complexes or with IgE and Ag (data not shown).

Similarly, IgG immune complex-induced ERK1/2 phosphorylation was of a higher intensity in LAT-YFFF than in wt PCMCs, whereas no ERK1/2 phosphorylation was detectably induced in LAT-FYYY and in LAT−/− PCMCs. LAT-FFFF permitted a faint ERK1/2 phosphorylation (Fig. 5B).

Taken together, data described in this section indicate that, by contrast with what was observed in BMMCs, LAT is mandatory for exocytosis, cytokine secretion, and MAPK activation in PCMCs and that LAT Y136 plays a prominent role for all responses examined in these cells. The three distal tyrosines, instead, seem to contribute negatively to these responses in PCMCs.

Discussion
We report here that, although LAT primarily generates positive signals as a consequence of FcεR engagement in mouse mast cells, it also contributes negative signals. Positive and negative signals depend on tyrosine residues that are phosphorylated following FcεR aggregation. Fig. 6 recapitulates the respective contributions of LAT tyrosine residues in mast cell activation unraveled by our work. This
MCs are a new in vitro model and possess all features characteristic of mast cells. BMMCs are immature mucosal-type mast cells. PCMCs are otherwise activated cells. We used here two types of cultured model for studying LAT-dependent immunoreceptor signaling in differentiation. It follows that mast cells provide a well-suited the TCR for T cell differentiation, FcRI knock-in mice. Another likely explanation is that, by contrast with were removed from the abnormal cytokine environment of the two mast cells. One possible reason is that, when cultured, mast cells –15), therefore does not affect LAT-YFFF mice, respectively (13). T cells toward a TH2 phenotype, observed in LAT-FYYY and in

FIGURE 5. IgG immune complex-induced β-hexosaminidase release, TNF-α secretion, and ERK1/2 phosphorylation in PCMCs from wt and LAT mutant mice. A. β-Hexosaminidase release. PCMCs were challenged with preformed immune complexes made of the indicated concentrations of GST and the indicated dilutions of a mouse anti-GST antiserum. β-Hexosaminidase released in the supernatants was measured by an enzymatic assay. Curves represent the percentage of β-hexosaminidase release as a function of the dilution of serum used to form immune complexes. B. ERK1/2 phosphorylation. PCMCs were challenged for 10 min with preformed immune complexes made of GST at the indicated concentrations and a 1/30 dilution of a mouse anti-GST antiserum. ERK1/2 and phospho-ERK1/2 were detected by Western blotting in whole cell lysates.

First of all, a major difference between mast cells and T cells expressing a mutant version of LAT was pointed out by this study. By contrast with T cells from LAT-FYYY or LAT-FFFF mice, which exhibited constitutively activated phenotypes associated with an exaggerated spontaneous secretion of TH2-type cytokines (13–15), mast cells derived from the same mutant mice exhibited an apparently resting phenotype. Unstimulated BMMCs from mutant mice contained comparable amounts of cytokine transcripts as BMMCs from wt mice. Also, following stimulation by IgE and Ag, BMMCs from all mice bearing a LAT mutation contained lower amounts of cytokine transcripts than wt BMMCs. This is interesting as mast cells depend on TH2 cytokines for their differentiation (23) and, when activated, they secrete primarily TH2 cytokines (24). The enhanced differentiation of αβ T cells and of γδ T cells toward a TH2 phenotype, observed in LAT-FYYY and in LAT-FFFF mice, respectively (13–15), therefore does not affect mast cells. One possible reason is that, when cultured, mast cells were removed from the abnormal cytokine environment of the two knock-in mice. Another likely explanation is that, by contrast with the TCR for T cell differentiation, FcεRI play no role in mast cell differentiation. It follows that mast cells provide a well-suited model for studying LAT-dependent immunoreceptor signaling in nonotherwise activated cells. We used here two types of cultured mast cells. BMMCs are immature mucosal-type mast cells. PCMCs are a new in vitro model and possess all features characteristic of mature serosal-type mast cells (O. Malbec et al., manuscript in preparation).

All responses of LAT−/− mast cells were of a lower magnitude than those of wt mast cells (Fig. 6, column 1). Therefore, LAT positively regulates FcεRI signaling. A positive effect could be ascribed not only to the four distal tyrosines as previously reported (12), but also to the five proximal tyrosines. Biological responses of LAT-FFFF cells were indeed more intense than those of LAT−/− cells (Fig. 6, column 2). Unless residues other than tyrosines contribute to the functions of LAT, this is the first evidence that the five proximal tyrosines may contribute to immunoreceptor signaling. Supporting this conclusion, we observed an inducible tyrosyl phosphorylation of LAT-FFFF in BMMCs. Molecules that are possibly recruited by these phosphotyrosines are unknown. Noticeably, the positive effects of LAT sequences containing the five proximal tyrosines enhanced those of the four distal tyrosines.

Compared with LAT-YYYY cells, all the biological responses examined were decreased in LAT-FFFF cells (Fig. 6, column 3). The four distal tyrosines therefore have a positive effect. Among the four distal tyrosines, Y136 plays a critical role. All responses examined in BMMCs and in PCMCs were decreased in LAT-FYYY cells, compared with LAT-YYYY cells (Fig. 6, column 4). Likewise, the three distal tyrosines contributed positively: all responses examined in BMMCs, but apparently not in PCMCs, were decreased in LAT-FFFF cells compared with LAT-YYYY cells (Fig. 6, column 7). Both Y136 and the three distal tyrosines are therefore required for FcεRI to trigger optimal responses and intracellular signals in BMMCs, whereas Y136 is apparently sufficient in PCMCs.

Noticeably, the relative contributions of Y136 and of the three distal tyrosines were not identical for all mast cell responses. Y136
and the three distal tyrosines contributed equally to β-hexosaminidase release in BMMCs (Fig. 6, line 1). Y136 had a predominant role for the Ca\(^{2+}\) response in BMMCs (Fig. 6, line 4) and for all the responses examined in PCMCs (Fig. 6, lines 6–8). The three distal tyrosines had a predominant role for the secretion of TNF-α, the up-regulation of cytokine transcripts, and the activation of ERK1/2 in BMMCs (Fig. 6, lines 2, 3, and 5). Similar results were recently found in LAT-deficient BMMCs reconstituted by tyrosine mutants of LAT (12). As suggested by data from the same investigation and from others conducted in T cells, LAT-dependent positive signals are thought to result from a cooperation between signaling molecules recruited by Y136, such as PLC-γ, and molecules recruited by the three distal tyrosines including Gads and Grb2 (25). PLC-γ is critical for the Ca\(^{2+}\) response (26), Grb2 is critical for ERK activation (27), and both intracellular pathways are required for exocytosis and cytokine secretion. No such a cooperation seems to occur (and be necessary) in PCMCs.

Noticeably, Y136 and the three distal tyrosines could apparently also exert a negative role. This mainly occurred when either Y136 (Fig. 6, column 8) or the three distal tyrosines (Fig. 6, column 5) were mutated. Except for the Ca\(^{2+}\) response, the positive role of Y136 was either decreased or replaced by a negative role when the three distal tyrosines were mutated in BMMCs (Fig. 6, compare columns 4 and 5). Likewise, the positive role of the three distal tyrosines was decreased or lost when Y136 was mutated in BMMCs (Fig. 6, compare columns 7 and 8). Although it was not mentioned in the work by Saitoh et al. (12), one observes a similar negative effect of Y136 on the recruitment of Grb2 by LAT in BMMCs. No negative effect was however observed by these authors on other mast cell responses. One reason might be that they used BMMCs in which LAT mutants were overexpressed following infection, whereas we used unmodified BMMCs from knock-in mice that constitutively expressed normal amounts of LAT. By contrast with BMMCs, we found that the three distal tyrosines were inhibitory on β-hexosaminidase release and ERK activation in PCMCs whether Y136 was mutated or not (Fig. 6, compare columns 7 and 8). Negative signals might be explained by the recruitment of molecules involved in negative signaling, such as SH2 domain-containing phosphatases (SHP), by mutant LAT. Although it does not exclude this possibility, we failed to detectably coprecipitate SHP-1, SHP-2, SHIP1, or SHIP2 with LAT-FYYY or LAT-YFFF (not shown). This is reminiscent of the negative effects of another LAT-like molecule, named LAX, recently described in lymphoid cells (28). LAX was tyrosyl-phosphorylated upon BCR or TCR engagement, and it recruited Gads, Grb2, and the p85 subunit of PI3K but, apparently, no phosphatase that could have accounted for its inhibitory properties.

The possibility that the negative effects observed in cells that expressed LAT mutants, whatever their mechanism, reveal that negative signals may emanate from wt LAT but be masked by overall dominant positive signals remains to be demonstrated. It is however supported by the following set of data. Positive and negative signals generated by different segments of the LAT molecule are apparently additive. Sequences containing the five proximal tyrosines could abrogate the negative effects of Y136 in the absence of the three distal tyrosines or the negative effects of the three distal tyrosines in the absence of Y136, e.g., on β-hexosaminidase release in BMMCs (Fig. 6, compare columns 5 and 6 and columns 8 and 9). Most importantly, LAT can apparently integrate positive and negative signals even when in a wt configuration. Thus, the positive effect of the four distal tyrosines on ERK1/2 activation in PCMCs appeared as resulting from an integration of the positive effect of Y136 and of the negative effect of the three distal tyrosines (Fig. 6, line 9). Surprisingly, negative cooperation of distinct LAT segments was also observed. This is most strikingly exemplified by β-hexosaminidase release in BMMCs: together, the four distal tyrosines had a positive effect (Fig. 6, line 1, column 3), but of a lower magnitude than the intense positive effects of either Y136 alone (Fig. 6, line 1, column 4) or of the three distal tyrosines alone (Fig. 6, line 1, column 7). These observations would be best explained if LAT could promote the assembly of a signaling complex composed of a mixture of intracellular molecules with antagonistic properties. Supporting this possibility, Gab2, that had been proposed to negatively regulate FceRII signaling in mast cells (29), was recently reported to negatively regulate TCR signaling when recruited to LAT via Grb2/Gads (30).

Finally, an unexpected finding of our work was that, as recurrently pointed out throughout this discussion, LAT differently
regulates FcR-dependent BMMC and PCMC activation. We found the following three main differences. First, LAT is dispensable in BMMCs (even though it is required for optimal responses), but indispensable in PCMCs. Comparable results were observed whether PCMCs were stimulated via FcεRI or FcγRIIA which share the same FcγRIγ and FcγR transduction subunits in mouse mast cells. Second, Y136 is necessary and apparently sufficient for LAT to support FcR signaling in PCMCs whereas BMMCs require all four distal tyrosines. Third, the three distal tyrosines could have a negative role in the two types of mast cells, whereas Y136 could have a negative role in BMMCs, but not in PCMCs. At least two nonexclusive mechanisms can explain differences between the two cell types: 1) a fyn-dependent, LAT-independent pathway was described in BMMCs (31) and 2) LAT-related molecules, named NTAL (32) or LAB (33), have been identified in mast, NK, and B cells. As LAT, these transmembrane adapters recruit SH2 domain-containing signaling molecules when tyrosyl-phosphorylated, and they positively regulate immunoreceptor signaling. That LAT was mandatory in PCMCs suggests either that the fyn-dependent pathway is not functioning in these cells or/and that LAT-like adapters that can partially replace LAT in BMMCs are absent in PCMCs.

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References