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Simvastatin Augments Lipopolysaccharide-Induced Proinflammatory Responses in Macrophages by Differential Regulation of the c-Fos and c-Jun Transcription Factors

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The 3-hydroxy-3-methylglutaryl-coenzyme A reductase inhibitors, or statins, are a widely used class of drugs for cholesterol reduction. The reduction in mortality and morbidity in statin-treated patients is incompletely explained by their effects on cholesterol, and an anti-inflammatory role for the drug has been proposed. We report in this work that, unexpectedly, simvastatin enhances LPS-induced IL-12p40 production by murine macrophages, and that it does so by activating the IL-12p40 promoter. Mutational analysis and dominant-negative expression studies indicate that both C/EBP and AP-1 transcription factors have a crucial role in promoter activation. This occurs via a c-Fos- and c-Jun-based mechanism; we demonstrate that ectopic expression of c-Jun activates the IL-12p40 promoter, whereas expression of c-Fos inhibits IL-12p40 promoter activity. Simvastatin prevents LPS-induced c-Fos expression, thereby relieving the inhibitory effect of c-Fos on the IL-12p40 promoter. Concomitantly, simvastatin induces the phosphorylation of c-Jun by the c-Jun N-terminal kinase, resulting in c-Jun-dependent activation of the TNF-α promoter, perhaps because the TNF-α promoter has C/EBP and AP-1 binding sites in a similar configuration to the IL-12p40 promoter. The fact that simvastatin potently augments LPS-induced IL-12p40 and TNF-α production has implications for the treatment of bacterial infections in statin-treated patients. The Journal of Immunology, 2004, 172: 7377–7384.
and TNF-α production in bone marrow-derived dendritic cells, and Monick et al. (14) reported that lovastatin increased TNF-α production by inhibition of Rho family GTPases in a murine macrophage cell line. Thus, statins have both anti-inflammatory and proinflammatory effects; however, the molecular mechanisms underlying these properties are poorly understood. In this study, we define the molecular mechanism underlying the proinflammatory response to statins. We demonstrate that simvastatin pretreatment enhances LPS-induced IL-12p40 production through activation of the IL-12p40 promoter, and that C/EBP and AP-1 binding sites are essential for this effect. Furthermore, we have shown that c-Fos repression and c-Jun N-terminal kinase activation underlie this mechanism. Simvastatin also augments TNF-α expression, which is also controlled by the C/EBP and AP-1 transcription factors.

Materials and Methods

Reagents

Simvastatin, c-Jun N-terminal kinase (JNK) inhibitor I, and negative control peptide were purchased from Calbiochem (La Jolla, CA). Simvastatin was converted to open acid form before use, as previously described (15). Briefly, 0.2 mg of simvastatin was suspended in 0.4 ml of ethanol, and 2.8 ml of 0.1 M NaOH was added. The solution was heated to 50°C for 2 h. To this heated solution, 3.6 ml of an aqueous solution containing 81 mM Na₂HPO₄ and 15 mM NaH₂PO₄ was added. The solution was heated to 40°C for 30 min, and the pH was adjusted to 7.3 with concentrated HCl.

The carrier was used as vehicle control in the same dilution as simvastatin. R848 was a gift from R. Ulevitch (The Scripps Research Institute, La Jolla, CA) (21). The expression vector (Promega, Madison, WI) (20). pGL2-mouse TNF-β, C/EBP, and AP1, and the IP-10 probe were described previously (24). Sequences for IL-12p40 and NF-κB were described previously (24). Simvastatin, farnesylpyrophosphate, geranylgeranylpyrophosphate, and Mevalonate was converted to open acid form before use, as previously described (15).

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Luciferase assay

RAW264.7 cells were seeded at 1.5 × 10⁵ cells/well in 18 cm plate on the day before transfection. pGL3-IL-12p40 promoter constructs and pBluescript SKII(+) were transiently transfected by FuGene6 (Roche Diagnostics, Indianapolis, IN), following the manufacturer's instruction. Cells were subsequently treated with vehicle or 8 μM simvastatin for 20 h and stimulated with or without 10 ng/ml LPS for 5 h. Luciferase assays were done by dual-luciferase assay kit, according to the manufacturer's instruction (Promega). The pooled stable transfectants of RAW264.7 cells, expressing either the ELAM or IL-12p40 promoter-driven firefly luciferase, were seeded at 5 × 10⁵ cells/well into 96-well plates on the day before stimulation. Cells were subsequently treated with vehicle or 8 μM simvastatin for the indicated time periods and stimulated with or without 1 ng/ml LPS for 6 h. Luciferase assays were performed using the luciferase 1000 assay system (Promega). The protein concentration of the cell lysate was determined by the bicinchoninic acid method (Pierce, Rockford, IL) in parallel. The luciferase values were normalized to protein concentration. All assays were done in triplicate, and each experiment was repeated at least three times. The luciferase activity was calculated for each individual assay. Fold activation was calculated by dividing the luciferase values for the test conditions by the relative luciferase value for the control condition. In all cases, values are the mean ± SD of triplicate wells and are representative of at least three separate experiments.

ELISA

The concentration of IL-12p40 and TNF-α was measured with DuoSet ELISA system (R&D Systems, Minneapolis, MN). The production of IFN-γ-inducible protein-10 (IP-10) was determined with IP-10 ELISA system (Cedarlane Laboratories, Hornby, Ontario, Canada).

Real-time PCR analysis

After stimulation, total RNA was isolated with an RNeasy minikit (Qiagen, Valencia, CA). Approximately 2 μg of total RNA was treated with DNase I (Fisher Scientific, Pittsburgh, PA) and reverse transcribed using Moloney murine leukemia virus reverse transcriptase and oligo(dT) primers (Promega). Quantitative real-time PCR was performed on an ABI 7700 (Applied Biosystems, Foster City, CA) using TaqMan Universal PCR Master Mix (Applied Biosystems). All data were normalized to elongation factor-1α (EF-1α) expression in the same cDNA set. Each experiment was performed independently at least three times, and the results of one representative experiment are shown. Sequences of TaqMan probes and forward and reverse primers for TNF-α and IFN-α were described previously (24). Sequences for IL-12p40 and IP-10 are as follows: IL-12p40 probe, CTGACGAGAACACATTGCACATTGG; forward primer, GCTCCAGATGCTTACACACCT; reverse primer, TCTTCTTATGTTCCACCTTCTT. IP-10 probe, TACTGTCGCGTCTACGTATGGA; forward primer, GACGGTC GTGCCAAGCT; reverse primer, GCTTCCATGGCCCTTAT. "EMSA"

The nuclear extracts were prepared, as described previously (25). The protein concentration was determined by bicinchoninic acid method (Pierce).
Probes were made by annealing single-stranded oligonucleotides and labeled by filling in with [α-32P]dCTP using Klenow fragment (Promega). Five micrograms of nuclear extracts were mixed with 5 × 10^6 cpm probe in 20 mM HEPES (pH 7.9), 50 mM KC1, 1 mM EDTA, 0.2 mM EGTA, 5% glycerol, 1 mM DTT, and 1× protease inhibitor mixture (Roche Diagnostics) at room temperature for 30 min. Bound and free DNAs were then resolved by electrophoresis through a 5% polyacrylamide gel (0.5× Tris-glycine-EDTA buffer) at 25 mA for 90 min. For supershift analysis, Abs were incubated with the nuclear extracts for 1 h on ice, followed by an additional incubation for 30 min with labeled probes. Binding activities were measured by PhosphorImager (Molecular Dynamics, Sunnyvale, CA).

**Immunoblot analysis**

Forty micrograms of nuclear extracts or 100 μg of whole cell lysates were denatured in SDS, electrophoresed on a 12% SDS-polyacrylamide gel, and transferred to polyvinylidene difluoride membrane (Immobilon-P; Millipore, Billerica, MA). Ponceau S staining was performed to ensure equivalent gel loading. Membranes were then incubated with the indicated rabbit polyclonal Abs, followed by HRP-conjugated goat anti-rabbit IgG (Zymed Laboratories, South San Francisco, CA), which were detected using West-Pico chemiluminescence reagent (Pierce).

**In vitro kinase assay**

RAW264.7 cells (2 × 10^5 cells in a 35-mm dish) were washed twice with ice-cold PBS and lysed in lysis buffer (1× PBS, 0.2% IGEPAL CA630, 0.1% SDS, 0.25% sodium deoxycholate, 2 mM Na3VO4, 1× complete protease inhibitor). Lysates were passed through a 26-gauge needle five times, clarified by centrifugation, and assayed for protein content (Pierce). Five hundred micrograms of whole cell lysates were suspended in JNK assay buffer containing 10–15 μM GST-c-Jun1–89 beads (Cell Signaling Technology) on a rotator for 1 h. The beads were washed and transferred to JNK assay buffer containing 10 μCi of [γ-32P]ATP, and JNK activity was determined (26).

**Results**

**Simvastatin augments LPS-induced IL-12p40 production by macrophages**

To study the effect of simvastatin on IL-12 production by macrophages, we pretreated murine resident peritoneal macrophages with simvastatin for 20 h and stimulated them with LPS for 8 h. As shown in Fig. 1A, simvastatin primed the cells for enhanced LPS-induced IL-12p40 production, and this occurred in a dose-dependent manner. Simvastatin also augmented IL-12p40 production in response to other Toll-like receptor agonists, such as R848 and Pam3-CSK4 (Fig. 1A). The effect was not general, because LPS-induced IP-10 secretion was decreased in simvastatin-treated cells (Fig. 1B). Because mechanistic studies are not possible in primary macrophages, we decided to focus on the murine macrophage cell line RAW264.7. The primary cell data were recapitulated in the cell line, thereby validating the model (Fig. 1, C and D). We next examined whether simvastatin regulates IL-12p40 production at the mRNA level. RAW264.7 macrophages were pretreated with simvastatin for 20 h and stimulated with LPS, and the levels of IL-12p40 and IP-10 mRNA were measured by quantitative PCR. As shown in Fig. 1E, during the early time points (2–6 h), simvastatin pretreatment enhanced LPS-induced IL-12p40 mRNA levels by 5- to 10-fold. Interestingly, at later time points (12–24 h), the increased IL-12p40 mRNA levels persisted, hinting at additional forms of regulation. By contrast, simvastatin pretreatment inhibited LPS-induced IP-10 mRNA levels (Fig. 1F). Taken together, these data indicate that simvastatin pretreatment enhances LPS-induced IL-12p40 production, whereas it inhibits LPS-induced IP-10 production.

**Simvastatin activates IL-12p40 promoter-luciferase constructs**

To investigate whether simvastatin enhances IL-12p40 production at the transcriptional level, we compared the effect of simvastatin on RAW264.7 cells that stably express the IL-12p40 promoter linked to the firefly luciferase gene with cells that stably express the ELAM promoter-firefly luciferase gene. As shown in Fig. 2B, simvastatin activated the IL-12p40 promoter, and substantially augmented the LPS response. By contrast, simvastatin did not activate the ELAM promoter and did not augment LPS stimulation of the ELAM promoter. This clearly demonstrates that simvastatin regulates the IL-12p40 promoter. The promoter of the gene encoding IL-12p40 has been studied in great detail. The murine IL-12p40 promoter has an Ets, an NF-κB, a C/EBP, and an AP-1 binding site at −218 to −213, −131 to −122, −96 to −88, and −79 to −74 relative to the transcription start site, respectively (Fig. 2A). These transcription factors have been shown to functionally cooperate with each other for efficient transcription of IL-12p40 in response to LPS (27–30). To delineate the statin-responsive elements, we made a series of luciferase reporter constructs in which we serially deleted the
known cis elements of the IL-12p40 promoter (Fig. 2C). The −352 construct has all known cis elements for IL-12p40 induction. The −194 construct lacks the Ets-binding element; the −127 construct lacks the NF-κB-binding element; the −81 construct lacks the C/EBP binding site. RAW264.7 cells were transiently transfected with these constructs and then treated with simvastatin. After 20 h of incubation, the cells were stimulated for 5 h with 10 ng/ml LPS, and the luciferase activities were measured. The −352, the −194, and the −127 luciferase constructs were activated by 5- to 7-fold in response to simvastatin treatment (Fig. 2C). Both the −81 and the empty pGL3 constructs were activated −2-fold by simvastatin, suggesting a small, promoter-independent effect.

Progressive deletion of the IL-12p40 promoter demonstrated the requirement for Ets, NF-κB, and C/EBP in LPS responsiveness in simvastatin-treated cells (Fig. 2C). By contrast, simvastatin alone activated the −352, −194, and −127 constructs equally, suggesting that the statin-responsive elements are contained within the −127 construct (Fig. 2C). To more precisely delineate the statin-responsive elements, we introduced a 2-bp substitution in either the NF-κB, C/EBP, or AP-1 binding sites in the −352 construct, and examined their response to simvastatin and/or LPS. As previously described, all mutants lost the response to stimulation with LPS alone (Fig. 2D) (29). By contrast, the NF-κB mutant showed a normal response to simvastatin treatment, whereas both C/EBP and AP-1 mutants were unresponsive (Fig. 2D). Taken together, these data demonstrate that NF-κB does not play a role in the simvastatin effects on the IL-12p40 promoter.

We next used dominant-negative inhibitors to address the involvement of C/EBP and AP-1 transcription factors in the simvastatin response. We used a strategy in which we introduced artificial homologues that form heterodimers with C/EBP and AP-1, thereby preventing them from activating target genes; C/EBP was inhibited with A-C/EBP (16), and AP-1 was inhibited by A-Fos (17). Either A-C/EBP or A-Fos substantially inhibited the simvastatin-mediated response. Moreover, A-C/EBP suppressed the basal activity of the IL-12p40 promoter by 70% (Fig. 2E). These data suggest that the C/EBP and AP-1 transcription factors have a role in the augmentation of the IL-12p40 promoter activity by simvastatin.

We explored the temporal effect of simvastatin on IL-12p40 promoter activity in stably transfected RAW264.7 cells. At early time points (up to 4 h), simvastatin had no effect on the IL-12p40 promoter activity (Fig. 2F). After 8 h, there was a gradual increase in IL-12p40 promoter activity (Fig. 2F). Importantly, there was no temporal difference in AP-1 binding to the IL-12p40 promoter (Fig. 2G). Taken together, these data clearly demonstrate that there is no initial spike in AP-1 activity.

A-Fos are dominant-negative forms of C/EBP and AP-1 transcription factors, respectively. The luciferase titers are shown as relative luciferase units (R.L.U.) compared with mock-transfected cells with vehicle treatment. F. The effect of simvastatin on IL-12p40 promoter activity. The pooled stable transfected of RAW264.7 cells expressing the IL-12p40 promoter-driven luciferase was treated with vehicle or 8 μM simvastatin for the indicated time periods. Luminescence was measured and normalized to protein concentration. G. AP-1-binding activity was conducted by EMSA in RAW264.7 cells. The cells were incubated with vehicle or 8 μM simvastatin for the indicated periods. Nuclear extracts (5 μg/lane) were mixed with radiolabeled probe −88–56 in the presence of 0.5 μg of poly(dI-dC). The probe −88–56 encompasses the AP-1 binding site on the IL-12p40 promoter. AP-1-binding activity was measured by PhosphorImager.
Simvastatin reduces c-Fos binding to the IL-12p40 promoter

We examined the effect of simvastatin on the binding activities of the C/EBP and AP-1 transcription factors to the IL-12p40 promoter. Radiolabeled double-stranded oligonucleotide probes spanning the sequence from position −103 to −76, and −88 to −56, were prepared and used to detect the C/EBP and AP-1 complexes, respectively. LPS stimulation enhanced the binding activities of both C/EBP and AP-1, as previously reported (data not shown) (29, 30). Simvastatin pretreatment did not show any effects on either constitutive or LPS-induced C/EBP-binding activities (data not shown). C/EBP refers to a family of transcription factors; and the formal possibility existed that a different C/EBP family member might be induced to bind to the IL-12p40 promoter when the cells were treated with simvastatin. Supershift assays were performed using polyclonal Abs against C/EBPα, -β, -δ, or -ε, and nuclear extracts from LPS-stimulated RAW264.7 cells with or without simvastatin pretreatment. The majority of the C/EBP complex consisted of C/EBPβ and C/EBPδ, and this was not changed by the presence of simvastatin (Fig. 3A). Simvastatin pretreatment slightly inhibited the augmentation of AP-1-binding activities by LPS (data not shown). Supershift assays demonstrated that the AP-1 complex formed in response to LPS contained c-Jun, JunB, c-Fos, FosB, and Fra-2 (Fig. 3B). Importantly, simvastatin pretreatment decreased c-Fos dramatically from the AP-1 complex (Fig. 3B). This decrease in c-Fos is due to a simvastatin-mediated inhibition of LPS-induced c-Fos synthesis (Fig. 3C).

Ectopic expression of c-Fos negatively regulates the IL-12p40 promoter, whereas expression of c-Jun positively regulates the promoter

The simvastatin-mediated inhibition of c-Fos binding appeared to be paradoxical. On the one hand, simvastatin clearly triggered IL-12p40 production through the AP-1 binding site (Fig. 2, D and E). In contrast, simvastatin drastically reduced LPS-mediated c-Fos induction and binding to the promoter (Fig. 3, B and C). This suggested that the reduction of c-Fos contributed to the activation of the IL-12p40 promoter. Ectopic expression of c-Fos reduced p40–127 promoter activity by 70% in the simvastatin-treated cells, while c-Jun expression enhanced it (Fig. 3D). Thus, these findings suggest a resolution to the paradox: c-Fos negatively regulates the IL-12p40 promoter, whereas c-Jun augments it. Ectopic expression of JunB, FosB, and Fra-2 had no effect on IL-12p40 promoter activity (Fig. 3D).

Simvastatin induces c-Jun phosphorylation

c-Jun is activated when c-Jun N-terminal kinase (JNK) phosphorylates Ser63 and Ser73. An in vitro kinase assay showed that simvastatin pretreatment provoked JNK activation at 12 h in RAW264.7 cells (Fig. 4A). Moreover, simvastatin pretreatment significantly induced Ser63 phosphorylation of the endogenous c-Jun protein (Fig. 4B, lane 6) and substantially augmented LPS-induced phosphorylation at the same site (Fig. 4B, lanes 7-10). Thus, JNK activation appeared responsible for the simvastatin-mediated IL-12p40 promoter activation. This was tested directly using a cell-permeable peptide inhibitor of JNK (31). Indeed, the JNK inhibitor diminished the simvastatin-mediated JNK activation in a dose-dependent manner (Fig. 4C), and this was accompanied by the partial inhibition of transcriptional activation of the IL-12p40 promoter (Fig. 4D). Taken together, simvastatin diminished the negative regulator, c-Fos, by inhibiting its induction, and augmented the positive regulator, c-Jun, through phosphorylation.

FIGURE 3. The effect of simvastatin on C/EBP and AP-1 DNA-binding activity, c-Fos expression, and the regulation of IL-12p40 promoter activity stimulated by ectopic expression of members of the AP-1 complex. A, RAW264.7 cells were pretreated with vehicle or 4 μM simvastatin for 16 h and stimulated with 10 ng/ml LPS for 4 h. Nuclear extracts (5 μg/lane) were prepared and incubated with 400 ng of the following polyclonal Abs on ice for 1 h in the presence of 1.5 μg of poly(dI-dC): anti-C/EBPα (lanes 1 and 6), anti-C/EBPβ (lanes 2 and 7), anti-C/EBPδ (lanes 3 and 8), anti-C/EBPε (lanes 4 and 9), and anti-C/EBPβ plus anti-C/EBPδ (lanes 5 and 10). Following this incubation, labeled probe −103–76 was added at room temperature for 30 min before electrophoresis. The probe −103–76 encompasses the C/EBP binding site on the IL-12p40 promoter. B, RAW264.7 cells were pretreated with vehicle or 4 μM simvastatin for 16 h and stimulated with 10 ng/ml LPS for 4 h. Nuclear extracts (5 μg/lane) were incubated with 400 ng of the following polyclonal Abs on ice for 1 h in the presence of 0.5 μg of poly(dI-dC): normal IgG (lanes 1 and 9), anti-c-Jun (lanes 2 and 10), anti-JunB (lanes 3 and 11), anti-JunD (lanes 4 and 12), anti-c-Fos (lanes 5 and 13), anti-FosB (lanes 6 and 14), anti-Fra-1 (lanes 7 and 15), and anti-Fra-2 (lanes 8 and 16). After this incubation, labeled probe −88–56 was added at room temperature for 30 min before electrophoresis. The probe −88–56 encompasses the AP-1 binding site on the IL-12p40 promoter. C, Effect of simvastatin on c-Fos expression. RAW264.7 cells were pretreated with vehicle or 4 μM simvastatin for 16 h and stimulated with 10 ng/ml LPS for the indicated time periods. Nuclear extracts (40 μg/lane) were subjected to SDS-PAGE. Expression of c-Fos was determined by immunoblot analysis. D, The effect of ectopic expression of members of the AP-1 complex on IL-12p40 promoter activity. RAW264.7 cells were transfected with either 10 ng of empty vector (Mock), or 10 ng of plasmid expressing c-Jun, JunB, c-Fos, FosB, or Fra2, together with 180 ng of the IL-12p40–127 promoter construct and 20 ng of pHRG-TK. Cells were incubated with vehicle or 8 μM simvastatin for 24 h, and luciferase activity was measured. The luciferase titers are shown as R.L.U. compared with mock-transfected cells with vehicle treatment.

The intermediates of the cholesterol biosynthetic pathway restore the normal response to LPS in simvastatin-pretreated cells

To show that the simvastatin effect was due to its inhibition of the HMG-CoA reductase, we demonstrated that mevalonate, farnesylpyrophosphate, and geranylgeranylpyrophosphate, intermediates in the cholesterol biosynthetic pathway, reversed the effect.
Simvastatin also augments TNF-α production by transcriptional activation

The promoter analysis indicates that both C/EBP and AP-1 binding sites are critical for the simvastatin-induced augmentation of IL-12p40 promoter. Other promoters containing both C/EBP and AP-1 binding sites might also be expected to be activated by simvastatin treatment. To confirm this hypothesis, we examined another inflammatory cytokine whose promoter has a similar structure to the IL-12p40 promoter. The unique feature of IL-12p40 promoter is that the C/EBP and AP-1 binding sites are located close to each other. The human TNF-α promoter has C/EBP and AP-1 binding sites similarly apposed (32–34), and, based on sequence homology, so does the murine TNF-α promoter. As expected, simvastatin induced TNF-α promoter-luciferase construct by 7-fold (Fig. 6A), and this was mirrored by increased TNF-α mRNA and secreted protein (Fig. 6, B and C).

Discussion

We have shown that simvastatin augments IL-12p40 production by macrophages in response to LPS stimulation. Our promoter analysis indicates, for the first time, that C/EBP and AP-1 binding sites are close to each other. The human TNF-α promoter has that the C/EBP and AP-1 binding sites are located close to each other. The human TNF-α promoter has C/EBP and AP-1 binding sites similarly apposed (32–34), and, based on sequence homology, so does the murine TNF-α promoter. As expected, simvastatin induced TNF-α promoter-luciferase construct by 7-fold (Fig. 6A), and this was mirrored by increased TNF-α mRNA and secreted protein (Fig. 6, B and C).
are essential for simvastatin-mediated IL-12p40 enhancement. Previous studies reported that statins increase the production of proinflammatory cytokines by myeloid cells (13, 35), but the mechanism by which this occurs has not been addressed. Our data strongly suggest that statin-mediated c-Fos repression is responsible for increased proinflammatory cytokine production. Simvastatin inhibited the association of c-Fos with the AP-1 binding site of the IL-12p40 promoter, suggesting that c-Fos negatively regulates proinflammatory cytokine production. This was confirmed by the observation that ectopically expressed c-Fos inhibited IL-12p40 promoter activation. c-Fos is known to form stable heterodimers with Jun proteins, thereby enhancing their DNA-binding activities (36). Although c-Fos cannot homodimerize and bind DNA, c-Jun can. Although a number of studies have demonstrated that heterodimers of c-Fos and c-Jun can activate the transcription of proinflammatory genes, recent evidence demonstrates that this heterodimer also has the capacity to inhibit transcription (37). Furthermore, we show in this study that c-Jun and c-Fos have opposing effects on IL-12p40 promoter activity, suggesting that c-Fos/ c-Jun heterodimers are inhibitory, whereas c-Jun/c-Jun homodimers are stimulatory. The JNK is known to phosphorylate c-Jun, resulting in the activation of this transcription factor. Consistent with this, we found that simvastatin stimulated JNK-dependent phosphorylation of c-Jun.

Although the AP-1 binding site was critical for simvastatin-mediated activation of the IL-12p40 promoter, so was the C/EBP binding site, and this was confirmed by the demonstration that dominant-negative C/EBP completely abolished the simvastatin effect. It is not clear how these sites functionally interact to mediate the simvastatin effect; one possible explanation is that AP-1 requires the assistance of C/EBP to enhance IL-12p40 transcription.

Two recent reports present compelling evidence that statins have potent anti-inflammatory effects in an EAE model. Atorvastatin induced STAT6 phosphorylation and secretion of Th2 cytokines such as IL-4, IL-5, IL-10, and TGFB-1. In contrast, STAT4 phosphorylation was inhibited, resulting in suppression of Th1 cytokine secretion (IL-2, IL-12, IFN-γ, and TNF-α) (9, 10). Although these observations appear to be at variance with our results, a number of important differences exist between the two experimental systems. Principally, Ag presentation is crucial for both the initiation and the progression of EAE (38), whereas we were examining LPS-induced cytokine production by macrophages.

There is also evidence for a proinflammatory effect of statins in human disease. Mevalonate kinase (MK) is the next enzyme after HMG-CoA reductase in the cholesterol biosynthetic pathway. Mutation of the MK gene results in two inactivation states. MK inhibition of c-Fos repression is responsible for increased proinflammatory responses when treated individuals experience bacterial infection. Although these observations appear to be at variance with our results, recent evidence demonstrates that this heterodimer has the capacity to inhibit transcription (37).

References

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