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Nasal Vaccination with Myelin Oligodendrocyte Glycoprotein Reduces Stroke Size by Inducing IL-10-Producing CD4$^+$ T Cells$^1$

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Inflammation plays an important role in ischemic stroke and in humans IL-10 may have a beneficial effect in stroke. We mucosally administered myelin oligodendrocyte glycoprotein (MOG) 35–55 peptide to C57BL/6 mice before middle cerebral artery occlusion (MCAO) to induce an anti-inflammatory T cell response directed at CNS myelin. Nasal and oral administration of MOG$_{35-55}$ peptide decreased ischemic infarct size at 24 and 72 h after MCAO surgery. Nasal MOG$_{35-55}$ peptide was most efficacious and reduced infarct size by 70% at 24 h and by 50% at 72 h ($p < 0.0001$ vs control) and also improved behavior score. Immunohistochemistry demonstrated increased IL-10 and reduced IFN-$\gamma$ in the area surrounding the ischemic infarct following nasal treatment. Nasal MOG did not reduce infarct size in IL-10-deficient mice. Adoptive transfer of CD4$^+$ T cells from nasally tolerized mice before MCAO surgery decreased stroke size ($p < 0.001$ vs control), whereas, CD4$^+$ T cells from nasally tolerized IL-10-deficient mice had no effect. Our results demonstrate that IL-10-secreting CD4$^+$ T cells induced by nasal MOG reduce injury following stroke. In addition, we observed a dramatic reduction of CD11b$^+$ cells in nasal MOG-treated animals. CD11b$^+$ cells may contribute to secondary infarct expansion by enhancing NO synthesis that may be reduced by elevated IL-10 levels. Modulation of cerebral inflammation by nasal vaccination with myelin Ags that increase IL-10 in the brain may improve outcome after stroke and enhance mechanisms of recovery. *The Journal of Immunology*, 2004, 172: 6549–6555.

Ischemic stroke results from transient or permanent reduction in cerebral blood flow. It is one of the main causes of morbidity and mortality worldwide. The mortality from stroke is ~30%, 80–90% of stroke survivors exhibit motor weakness, and 40–50% experience sensory disturbances (1). In the center of the perfusion deficit, cerebral blood flow is typically 80% below normal levels (2). Ischemic tissue dies over minutes to many hours (2).

Inflammation is also initiated by ischemia at the blood-microvascular endothelial cell interface and contributes significantly to CNS damage. Polymorphonuclear leukocytes rapidly enter injured brain tissue (3) and white blood cells traverse the blood-brain barrier (BBB) 12–24 h after onset and may provide a source of oxygen-free radicals. Eventually, the infarcted zone is infiltrated with lymphocytes, polymorphonuclear cells, and macrophages (4). Neutrophils, important cellular components of the innate immune response, produce a number of potentially harmful substances including toxic oxygen metabolites, destructive enzymes, and pro-inflammatory cytokines with neurotoxic properties (5). Thus, the severity of postischemic injury can be affected by manipulation of the inflammatory response. Agents that limit white blood cell adhesion to endothelial receptors, such as Abs to P-selectin (6) and ICAM-1 (7) reduce ischemic injury. However, in a multicenter acute clinical trial, anti-human ICAM-1 Ab increased mortality (3). Another immunological therapeutic approach is modulation of cytokine production.

Following ischemia, most cells in the brain, including endothelial cells, microglia, astrocytes, and neurons, can produce IL-1$\beta$, IL-6, and TNF-$\alpha$ that can directly induce cell death as well as contribute to vessel wall injury, hemorrhage, edema, and tissue necrosis (8). There are immunoregulatory cytokines that can modulate immune processes and inhibit expression of inflammatory Th1-type responses as well as affect inflammation in general. Among them are IL-10 and TGF-$\beta$ (9). IL-10 is preferentially produced by Tr1-type regulatory T cells (10) and TGF-$\beta$ is preferentially produced by Th3-type regulatory cells which suppress both Th1 and Th2 cells (9, 11). It has been reported that elderly patients with a history of stroke had significantly lower median IL-10 levels compared with elderly patients without stroke (12, 13). Moreover, TGF-$\beta$ is expressed in brain following ischemia (14) and may play a role as an anti-inflammatory cytokine with neuroprotective properties. Mucosal (nasal or oral) administration of Ag preferentially induces regulatory T cells that secrete IL-10 and TGF-$\beta$ at the anatomic site where the mucosally administrated Ag is located, a phenomena termed bystander suppression. Becker et al. (15) previously demonstrated reduced infarct size in the rat after oral tolerance with myelin basic protein and suggested that TGF-$\beta$ may play a role because there was increased TGF-$\beta$ expression in the brain of orally tolerized animals. To investigate the role of T cells during the course of stroke, we studied mucosal tolerance in the mouse model of stroke. We mucosally administrated myelin oligodendrocyte glycoprotein (MOG)$^3$ 35–55 peptide in a mouse stroke model and demonstrate reduction of infarct size that is mediated by IL-10-dependent CD4$^+$ T cells (15). This

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3 Abbreviations used in this paper: MOG, myelin oligodendrocyte glycoprotein; MCAO, middle cerebral artery occlusion; BBB, blood-brain barrier.
approach has applicability both as prophylactic therapy to reduce the severity of stroke in patients with transient ischemia and potentially as therapy during acute stroke.

Materials and Methods

Animals

Female C57BL/6 and C57BL/10−/− were purchased from The Jackson laboratory (Bar Harbor, ME). These mice were 8–10 wk of age and were housed in Harvard Medical School Animal Care Facilities according to the institutional guidelines. All protocols for animal experiments were approved by local committee review and were conducted according to the National Institutes of Health Guide for Care and Use of Laboratory Animals.

Oral and nasal MOG administration

Oral administration. Mice were fed with 250 µg of MOG35–55 (obtained from Dr. J. Elliot, Yale university, New Haven, CT) each day for 5 days.

Nasal administration. Mice were treated nasally with 25 µg MOG35–55 three times every other day. As a control protein, mice received OVA325–345 peptide (SynPep, Dublin, CA). For in vitro studies, mice were immunized in the footpad with 100 µg MOG35–55 mixed 1:1 with CFA 2 days following the last treatment. Mice were subject to middle cerebral artery occlusion (MCAO) surgery 2 days following the last treatment.

Cell culture of lymphocytes

For proliferation and cytokine assays, spleen cells from mice were pooled and cultured in 96-well plates at 5 × 10^4 and 10^5 cell/mL, respectively, in serum-free medium and X-VIVO 20 (BioWhittaker, Walkersville, MD). To measure cytokines, culture supernatants were collected at 24 h for IL-2 and IL-4; 40 h for IL-6, IL-10, and IFN-γ; and at 72 h for TGF-β. For proliferation, cells were pulsed with thymidine at 72 h and radioactivity was determined 16 h later (16).

ELISA for cytokines

Quantitative ELISA for IL-2, IL-4, IL-10, and IFN-γ were performed using paired mAb specific for corresponding cytokines per the manufacturer’s recommendations (BD PharMingen, San Diego, CA). TGF-β was determined as previously described (17).

Model of focal cerebral ischemia

Animals were anesthetized with 2% isoflurane in 70% N2 and balanced O2 by a face mask. Cerebral infarcts were produced by 2 h of MCAO followed by reperfusion as described previously (18). To do so, we introduced a silicone-coated 8–0 monofilament into the left internal carotid artery and advanced 9 mm from the carotid bifurcation so as to occlude the middle cerebral artery. After 2 h, the animals were briefly reanesthetized and the filament was withdrawn. In randomly selected animals, relative changes of focal cerebral blood flow were monitored by laser Doppler flowmetry (P92B; Perimed, Stockholm, Sweden) as previously described (18, 19), and the left femoral artery was cannulated for arterial blood pressure and blood gas determination. Arterial blood samples were analyzed for pH, arterial oxygen pressure, and partial pressure of carbon dioxide by using a blood gas pH analyzer (Corning 178; Ciba-Corning Diagnostics, Medford, MA). Rectal temperature was maintained at 36.5–37.5°C with a temperature control unit (FHC, Brunswick, ME) during the monitoring period. In all experiments, animals underwent surgery, were scored, and the infarct volume was measured in blinded fashion.

Neurological deficits

Animals were tested for neurological deficits and scored as described elsewhere (20): 0, no observable neurological deficit (normal); 1, failure to extend right forepaw (mild); 2, circling to the contralateral side (moderate); and 3, loss of walking or righting reflex (severe). Animals were rated by a rater blinded to the treatment protocol either 24 or 72 h after reperfusion by an observer naive to the treatment group.

Determination of infarct size

After sacrifice, cerebral infarct sizes were determined on 2.35-triphenyltetrazolium chloride-stained 2-mm brain sections (24 h) or H&E-stained 20-mm cryostat sections (72 h) by means of an image analysis system (M4; Imaging Research, St. Catherine’s, Ontario, Canada) and calculated according to the formula: percentage of (contralateral hemisphere volume − noninfarct volume in ipsilateral hemisphere)/contralateral hemisphere volume.

Histology and immunohistology

Histology was performed on animals sacrificed 24 or 72 h after ischemia. Brain sections (frozen tissue sections) from mice before and after MCAO surgery were fixed in 4% paraformaldehyde overnight followed by 4.5% sucrose for 4 h, then 20% sucrose for overnight at 4°C. Brains were frozen in the presence of OCT and stored until used at −70°C. The staining included immunological markers for T cells (CD3, CD4, and CD8), macrophages and neutrophils (CD11b), plus pro-inflammatory (IFN-γ) and anti-inflammatory (IL-4, IL-10, and TGF-β) cytokines as previously described. Sections were evaluated in a blinded manner, and controls included use of isotype-matched mAbs and a demonstration that preabsorption of anti-cytokine Abs with respective or irrelevant cytokins (5 µg/ml, 16 h, 4°C) either blocked or left unchanged the results of Ab staining, respectively (21).

Adoptive transfer of CD4+ T cells

Mice were nasally treated with 25 µg of MOG on days 1, 3, and 5. On day 7, they were immunized with MOG in CFA. Ten days after immunization, both lymph nodes and spleens were removed and stimulated in vitro with 40 µg of MOG35–55 in 24-well plates (1 ml in each well containing 5 × 10^6 cells) in T cell medium buffer (22) for 2 days. On the third day, cells were split into two wells and incubated for 1 more day with IL-2. On day 4, the cells were harvested and CD4+ T cells were purified by negative selection using a mouse CD4+ T cell column (catalogue no. MCD43; R&D Systems, Minneapolis, MN). Briefly, 10^6 CD4+ T cells were injected in 0.2 ml DMEM i.v. Mice were subjected to MCAO surgery 2 days after adoptive transfer of cells. For controls, cells from animals immunized with CFA were treated in an identical fashion.

Data analysis

All continuous and ordinal data are expressed as mean ± SEM. Statistical analysis was performed by unpaired Student’s t test or by one-way ANOVA followed with Duncan’s test (infarct size) or by the Mann-Whitney U test (behavior score). Values of p < 0.05 were considered to be statistically significant.

Results

Cytokine profile following nasal or oral administration of MOG35–55 Peptide

Before studies in the stroke model, we investigated the cytokine profile induced by mucosal administration in C57BL/6 mice fed 250 µg of MOG35–55 peptide five times on consecutive days or nasally administrated 25 µg peptides three times every other day. Two days after the last treatment, mice were immunized with 100 µg MOG35–55 peptide in CFA and splenocytes were taken 10 days later for in vitro assays. As shown in Fig. 1, both nasally treated and fed mice showed a significant reduction in production of the proinflammatory cytokine IFN-γ compared with the control (p ≤ 0.0008 and p ≤ 0.007 respectively). Only nasally treated animals showed a significant elevation in the anti-inflammatory cytokine IL-10 (p ≤ 0.0001 vs control) and a decrease in the total cell proliferation (p < 0.02). No active TGF-β was observed in either nasally or orally treated animals. Having shown the tolerogenic potential of mucosally administrated MOG in C57BL/6 mice, we then proceeded to test mucosal MOG in the stroke model.

Mucosal tolerance to MOG35–55 peptide reduces tissue damage after stroke

Occlusion of the middle cerebral artery produces a localized infarct within the cerebral cortex and caudate putamen, which closely resembles the “at risk territory” in most embolic stroke patients (23). To investigate whether mucosal administration of MOG35–55 peptide affected tissue damage after ischemia insult, MOG35–55 peptide was administered by five repetitive gavages or
RESULTS

Cytokine profile of splenocytes from animals treated orally or nasally with MOG35-55 peptide. C57BL/6 mice were tolerized with MOG35-55 peptide through oral administration (250 μg every day for 5 days) or nasal administration (25 μg every other day for 5 days). Nasally PBS-treated mice served as a control group. Ten days after immunization, splenocytes were cultured at 0.5 × 10^6 (for proliferation assay) or 1 × 10^6 cells/well with 100 μg/ml MOG35-55 Peptide in 0.2 ml of medium. For proliferation, cells were pulsed with thymidine at 72 h and radioactivity was determined 16 h later (16). Cytokines were measured by ELISA at 24 h (IL-2 and IL-4), 40 h (IL-6, IL-10, and IFN-γ) was determined 16 h later (16). Cytokines were measured by ELISA at 24 h and 72 h (TGF-β) and results are expressed as SEM. *, p ≤ 0.01, nasal vs oral/control for IL-10 and IFN-γ; **, p = 0.02 oral vs control for IFN-γ.

Immunohistochemistry of poststroke brain following mucosal treatment

Surrounding the area of the lethally damaged core of the ischemic infarct lies the penumbra, an area of constrained blood flow with partially preserved energy metabolism (2). Given time and without treatment, the penumbra progresses to infarction because of ongoing excitotoxicity, posts ischemic inflammation, and apoptosis (24).

FIGURE 1. Cytokine profile of splenocytes from animals treated orally or nasally with MOG35-55 peptide. C57BL/6 mice were tolerized with MOG35-55 peptide through oral administration (250 μg every day for 5 days) or nasal administration (25 μg every other day for 5 days). Nasally PBS-treated mice served as a control group. Ten days after immunization, splenocytes were cultured at 0.5 × 10^6 (for proliferation assay) or 1 × 10^6 cells/well with 100 μg/ml MOG35-55 Peptide in 0.2 ml of medium. For proliferation, cells were pulsed with thymidine at 72 h and radioactivity was determined 16 h later (16). Cytokines were measured by ELISA at 24 h (IL-2 and IL-4), 40 h (IL-6, IL-10, and IFN-γ), and 72 h (TGF-β) and results are expressed as SEM. *, p ≤ 0.01, nasal vs oral/control for IL-10 and IFN-γ; **, p = 0.02 oral vs control for IFN-γ.

FIGURE 2. Cerebral ischemic infarct size following mucosal tolerance with MOG. A, Protocol of mucosal MOG peptide treatment before MCAO. B, At 24 h, brains were sectioned coronally (2 mm) and incubated in 2% 2,3,5-triphenyltetrazolium chloride (Sigma-Aldrich, St. Louis, MO) at room temperature for 30 min. The pale regions denote the infarcted tissue. Arrows indicate where tissue for immunohistochemistry was taken. The sections presented are from the entire brain of representative individual mice.

FIGURE 3. Ischemic infarct volume size and behavior score at 24 or 72 h after MCAO. A, Infarct volume (corrected for the presence of edema) at 24 h in oral (n = 12), nasal (n = 9), control (n = 12)-treated mice and at 72 h in oral (n = 9), nasal (n = 7), and control (n = 9) mice. Results are included from seven individual experiments at 24 h and eight individual experiments at 72 h. B, Behavior scores were determined at 24 or 72 h following MCAO surgery. *, p = 0.0001, nasal vs control for infarct volume/behavior; **, p < 0.02, oral vs control for infarct volume.
However, even in the most severe ischemic insult tissue destruction continues for hours to days following the insult (3, 4). Thus, it is evident that a prime goal of neuroprotection is to suppress ischemic inflammation within the penumbra. As shown in Fig. 4A, at 24 h IFN-γ, a proinflammatory cytokine, was not expressed within the penumbra in mice receiving nasal or oral therapy, compared with the control group. Both nasal and oral treatment groups showed the presence in the penumbra of the anti-inflammatory cytokine IL-10 in parallel with the presence of T cells, whereas only the nasal-treated group also showed TGF-β expression at 24 h. Neither IL-10 or TGF-β expression was seen in the brains of control mice. These data suggest a role for T cells and IL-10 in the reduction of ischemic infarct size poststroke following mucosal therapy. We also quantified CD4 and CD11b cells and cells expressing IFN-γ, IL-10, and TGF-β at 72 h. As shown in Fig. 4B, there was a reduction in the number of CD11b cells in the nasally treated group. No significant difference in CD4+ cells or in the expression of TGF-β (data not shown) was observed. Cells expressing intracellular IL-10 were increased in both the nasally and orally treated groups (p = 0.02 and p = 0.03, respectively, vs control). In addition, there was a decrease in the number of cells expressing IFN-γ in both the nasally and orally treated groups (p = 0.007 and p = 0.015, respectively, vs control).

The neuroprotective effect of nasal MOG is absent in IL-10−/− mice
To investigate the role of IL-10 in reduction of stroke size following nasal MOG, IL-10−/− mice were subjected to MCAO. As shown in Fig. 5, there was no significant reduction in ischemic infarct volume or improvement in behavior score following nasal tolerization with MOG35–55 peptide as compared with vehicle treatment in IL-10−/− mice. A role for IL-10 was also observed in IL-10−/− animals treated with a control peptide (OVA), as these mice showed a higher infarct volume 24 h after the surgery as compared with control mice.

FIGURE 4. Immunohistology of brain at 24 and 72 h after stroke. A, IL-10 and/or TGF-β expression within penumbra of MOG-treated mice at 24 h after stroke. The area of the penumbra where histochecistry was taken is shown by arrows in Fig. 2B. Serial sections of brains from control-, oral-, or nasal-treated mice 24 h after MCAO were labeled using (a–c) isotype control mAbs or with mAbs directed against (d–f) CD11b (g–i), CD4 (j–l), IFN-γ (m–o), or TGF-β (p–r). Results are representative of the blinded analysis of three samples/group (immunoperoxidase, hematoxylin counterstain; original magnifications, ×120). B, Brain sections at −1.9 Bregma from MOG35–55-treated mice and control (OVA) mice were stained with hematoxylin and with Abs against CD4, CD11b, IFN-γ, and IL-10 at 72 h after stroke. Quantification of cytokine expression in the brain was performed by counting the total number of cells staining in the area of the penumbra. Results are the SEM of three different brain sections from each group. *: Nasal vs control: for CD11b, p < 0.001; for IFN-γ, p = 0.02; for IL-10, p = 0.007. **: Oral vs control: for IFN-γ, p = 0.015; for IL-10 p = 0.03.
Mucosal Ag induces tolerance by a number of mechanisms, including anergy, deletion, and active cellular regulation (25). Our results thus far suggested that nasal MOG was effective in the stroke model by inducing IL-10-secreting CD4 T cells. To investigate the role of CD4 T cells in reduction of stroke size following nasal MOG, adoptive transfer experiments were performed. C57BL/6 mice were tolerized with MOG35–55 peptide through nasal administration as described in Fig. 2A. To obtain sufficient cells for adoptive transfer, 2 days after the last nasal treatment, mice were immunized with 100 μg of MOG35–55 peptide in CFA in the hind footpad and draining lymph node, and spleen cells were taken 10 days later and stimulated in vitro with 40 μg/ml MOG35–55 peptide. After 4 days on in vitro culture, CD4 T cells were purified by negative selection and adoptively transferred to untreated mice that were then subjected to MCAO surgery 48 h after CD4 T cell transfer. Adoptive transfer of CD4 T cells from nasal control-treated mice that were then immunized with CFA served as a control group. As shown in Fig. 6A, cerebral ischemic infarct size was reduced by 56% (from 70 ± 3% to 31 ± 3%, p < 0.0003) in animals that received CD4 T cells from MOG nasally treated mice, as compared with animals that received CD4 T cells from control mice immunized with CFA. To establish that IL-10 was also crucial in these adoptive transfer experiments, CD4 T cells from MOG35–55–tolerized mice to naive mice before MCAO surgery. As shown in Fig. 6B, no reduction of infarct size was observed when CD4 T cells from nasal MOG-treated wild-type (n = 6) or IL-10–/– (n = 10) vs control animals (n = 10). C, Behavior score 24 h following MCAO surgery. *p < 0.003, wild-type nasal MOG CD4 T cells vs wild-type nasal MOG CD4 T cells.

Adoptive transfer of CD4 T cells from MOG-tolerized mice to untreated mice reduces ischemic infarct size

In order to further investigate the role of CD4 T cells in the reduction of ischemic stroke, we employed adoptive transfer experiments. C57BL/6 mice were tolerized with MOG35–55 peptide through nasal administration as described in Fig. 2A. Adoptive transfer of CD4 T cells from nasal MOG-tolerized mice to naive mice before MCAO surgery. As shown in Fig. 6B, no reduction of infarct size was observed when CD4 T cells from nasal MOG-treated wild-type (n = 6) or IL-10–/– (n = 10) vs control animals (n = 10). C, Behavior score 24 h following MCAO surgery. *p < 0.003, wild-type nasal MOG CD4 T cells vs wild-type nasal MOG CD4 T cells.

Discussion

Accumulated data demonstrate that inflammation plays an important role in the pathophysiology of ischemic stroke. Between the lethally damaged ischemic core and the normal brain lies the peri-infarct zone or penumbra, an area of constrained blood flow with partially preserved energy metabolism (2). Given time and without treatment, the penumbra progresses to infarction because of ongoing excitotoxicity, posts ischemic inflammation, and apoptosis. However, even in the most severe ischemic insult, the process of tissue destruction may not be completed for hours or days (3, 5, 15). This provides an opportunity for strategies of neuroprotection to salvage the ischemic inflammation within the penumbra.
The presence of recruited leukocytes at the site of inflammation is dependent upon the coordinated expression of adhesion molecules (ligands and receptors) on inflammatory cells and the activated capillary endothelium, respectively. Naive T cells that are not activated generally do not cross the BBB. Nonetheless, T cells readily cross the BBB if a T cell response against a CNS autoantigen is initiated in lymphoid organs. T cells are restimulated upon encounter with the target immunogen presented by local APCs. Thus, several types of CNS cells that may act as APCs are activated following stroke including microglia- and macrophage-like cells that express MHC molecules and produce proinflammatory cytokines such as TNF-α and IL-1β, which may enhance the appearance of adhesion molecules (26). Following MCAO surgery, we observed macrophage-type cells (CD11b) by immunostaining of brain sections at 24 and 72 h (Fig. 4). These cells could act to enhance the destructive effect of infiltrating CD4+ T cells. Our results show that CD11b cells were decreased in nasal MOG-treated animals.

Oral or nasal Ag preferentially generates a Th2 (IL-4/IL-10)- or a Th3 (TGFB)-type response and these cells have suppressive properties for Th1 and other immune cells. Because regulatory T cells are triggered in an Ag-specific fashion but suppress via cytokine release in an Ag-nonspecific fashion, they mediate "by-stander suppression" when they encounter the fed autoantigen at the target organ. Thus, mucosal tolerance can be used to treat inflammatory processes that are not autoimmune in nature via the secretion of cytokines such as TGFB, IL-4, and IL-10 after Ag-specific triggering (25).

To address the role of tolerance and T regulatory cells in stroke, we tolerized C57BL/6 mice with the myelin autoantigen MOG35-55. MOG is a target in the autoimmune model experimental autoimmune encephalomyelitis and is further expressed when there is CNS damage (27). We found that animals tolerized against MOG had enhanced expression of the anti-inflammatory cytokine IL-10 and reduced expression of the proinflammatory cytokine IFNg in the pneumonia. It has recently been shown that IFNg parallels inducible NO synthase activity during the course of stroke (3). The role of IL-10 as mediator of the ischemic infarct was established by investigation of IL-10+/− mice.

IL-10 is an immunoregulatory cytokine that can modulate immune processes, inhibiting the expression of inflammatory Th1-type responses as well as affecting APC function. IL-10 has been shown to reduce inflammation in a variety of animal models including experimental autoimmune encephalomyelitis (22) and atherosclerosis (28). Within the brain, IL-10, may deactivate macrophage-like cells and astrocytes and thus limit their involvement in a secondary inflammatory process. Furthermore, IL-10 limits the role of glutamate cytotoxicity by inactivation of NF-kβ (29), a transcription factor that modulates inflammation and key regulatory proteins in cerebral ischemia (30). Moreover, IL-10 targets the interface between the CNS and periphery (BBB) by preventing adhesion and extravasation of leukocytes.

Becker et al. (15) previously demonstrated reduced infarct size in the rat after oral tolerance with myelin basic protein. Furthermore, recent experiments by Becker et al., (31) showed that adoptive transfer of splenocytes from nasal myelin basic protein-tolerized rats to naive rats reduces infarct size. This work suggested that TGFB may play a role since there was increased TGFB expression in the brain of orally tolerized animals. TGFB may be increased following stroke independent of mucosal tolerization, is expressed in neuronal cells, and may play an important role in neuronal recovery (14). We found increased expression of TGFB in nasally treated animals compared with controls at 24 h and increases in both treated and control animals at 72 h. Nevertheless, in our studies IL-10 appears to be the crucial cytokine in decreasing stroke size.

Our results cannot be directly compared with Becker et al. (15, 31) who used a different Ag (myelin basic protein), a rat model, and did not perform adoptive transfer experiments of CD4+ T cells to establish the immunological mechanism responsible for the effect described. Becker et al.’s (15, 31) results are consistent, however, in suggesting that mucosal tolerance can be used to treat stroke by inducing an anti-inflammatory milieu in the CNS.

In our study, C57BL/6 mice were investigated, as it is a common background for transgenic mice. Nasal in addition to oral administration was chosen because the level of IL-10 has been shown to be increased following nasal tolerization (32, 33) and elderly patients with a history of stroke were reported to have significantly lower median IL-10 production levels at baseline compared with elderly patients without stroke (12, 13). We found that nasal administration of MOG35-55 peptide was more effective then oral MOG and consistent with this, we found more IL-10 and less CD11b cells in nasally treated animals. It is known that nasal administration of proteins preferentially induce IL-10 responses as compared with oral administration (11).

Adaptive transfer of T cells is a classic method to demonstrate an active immunological role for both disease-inducing and disease-ameliorating T cells in animal disease models. Of note is that Schwartz and colleagues (34–36) in a series of studies have demonstrated a neuroprotective role for myelin-reactive T cells in models of optic nerve injury and spinal cord trauma. Our study is the first to demonstrate that CD4+ T cells expressing a regulatory T cell phenotype (IL-10) have a beneficial effect in stroke. Moreover, IL-10+/− CD4+ T cells lack this activity, further establishing an IL-10-dependent cascade.

In addition to the IL-10 effects we observed, we also found a marked reduction of the CD11b+ cell number in mice nasally administered MOG compared with the wild type. CD11b is a marker for cells of the microglia/macrophage lineage. These cells increase the level of NO synthase (leading to NO synthesis) and thus may play a role in the evolving lesion and functionally contribute to secondary infarct expansion after ischemic injury (37). The reduction of CD11b+ cells, which may have been secondary to IL-10 production, may also have contributed to the beneficial effects of nasal MOG we observed.

Enhancing recovery from stroke and limiting ischemic damage are major goals to decrease stroke mobility and mortality. Given that it has been suggested in humans that IL-10 may have a beneficial effect in stroke (13), our findings provide a novel immunological approach that is nontoxic and widely applicable for delivery of IL-10 to the site of ischemia to reduce stroke injury. Experiments are in progress to determine the effectiveness of this approach as therapy at the time of stroke. Nevertheless, based on our results, this approach may have applicability as prophylactic therapy to reduce the severity of stroke in high-risk patients with transient ischemic attacks.

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