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Up-Regulation of Cysteinyl Leukotriene 1 Receptor by IL-13 Enables Human Lung Fibroblasts to Respond to Leukotriene C4 and Produce Eotaxin

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Cysteinyl leukotrienes (CysLTs) play an important role in eosinophilic airway inflammation. In addition to their direct chemotactic effects on eosinophils, indirect effects have been reported. Eotaxin is a potent eosinophil-specific chemotactic factor produced mainly by fibroblasts. We investigated whether CysLTs augment eosinophilic inflammation via eotaxin production by fibroblasts. Leukotriene (LT)C4 alone had no effect on eotaxin production by human fetal lung fibroblasts (HFL-1). However, LTC4 stimulated eotaxin production by IL-13-treated fibroblasts, thereby indirectly inducing eosinophil sequestration. Unstimulated fibroblasts did not respond to LTC4, but coincubation or preincubation of fibroblasts with IL-13 altered the response to LTC4. To examine the mechanism(s) involved, the expression of CysLT1R in HFL-1 was investigated by quantitative real-time PCR and flow cytometry. Only low levels of CysLT1R mRNA and no CysLT1R protein were expressed in unstimulated HFL-1. In contrast, stimulation with IL-13 at a concentration of 10 ng/ml for 24 h significantly up-regulated both CysLT1R mRNA and protein expression in HFL-1. The synergistic effect of LTC4 and IL-13 on eotaxin production was abolished by CysLT1R antagonists pranlukast and montelukast. These findings suggest that IL-13 up-regulates CysLT1R expression, which may contribute to the synergistic effect of LTC4 and IL-13 on eotaxin production by lung fibroblasts. In the Th2 cytokine-rich milieu, such as that in bronchial asthma, CysLT1R expression on fibroblasts might be up-regulated, thereby allowing CysLTs to act effectively and increase eosinophilic inflammation. The Journal of ImmunoL 2003, 170: 4290–4295.

In addition to direct effects, indirect effects of CysLTs on eosinophils have been reported. PBMCs isolated from patients with asthma produce IL-5 in response to stimulation with mite Ag. CysLTs may induce IL-5 production resulting in an eosinophil influx, which can be prevented by treatment with LTRA or anti-IL-5 Ab (11). Eotaxin is a potent eosinophil-specific chemotactic factor that plays a central role in eosinophilic airway inflammation in asthma (12). Recently, lung fibroblasts have been reported to produce eotaxin, and their activation can be modulated by inflammatory cytokines such as IL-4 and IL-13 (13–17).

Little is known about the response of fibroblasts to CysLTs and the expression of CysLT1R on fibroblasts. The in situ hybridization pattern of CysLT1R in normal human lung is characterized by the strict localization to smooth muscle cells (SMC) and some macrophages (18). Thivierge et al. (19) showed that Th2 cytokines such as IL-4, IL-5, and IL-13 have a regulatory role in CysLT1R expression in monocytes/macrophages (20) or HL-60 cells. We hypothesized that CysLTs augment eosinophil inflammation via eotaxin production by fibroblasts. To examine this hypothesis, we investigated whether LTC4, alone or in combination with IL-13, can stimulate eotaxin production by fibroblasts. Furthermore, we examined whether IL-13 up-regulates CysLT1R expression in fibroblasts.

Materials and Methods

Reagents

Human rIL-13 and anti-eotaxin neutralizing Ab were obtained from R&D Systems (Minneapolis, MN). LTC4, LTD4, and rabbit polyclonal anti-human CysLT1R Ab were purchased from Cayman Chemicals (Ann Arbor, MI). Rabbit IgG isolate and DAKO IntraStain kits were obtained from DAKO (Copenhagen, Denmark). The LTRAs pranlukast and montelukast were gifts from Ono Pharmaceutical (Osaka, Japan) and Merck (Whitehouse Station, NJ), respectively. Eotaxin ELISA kits were purchased from BioSource International (Camarillo, CA).
Cell culture
Human fetal lung fibroblasts (HFL-1; lung, diploid, human, passage 7–12 cells) were obtained from the American Type Culture Collection (Manassas, VA). The HFL-1 cells were seeded in 12-well tissue culture plates at a density of 5 × 10⁴ cells/ml and cultured at 37°C in a 5% CO₂-humidified incubator in Ham’s F12K medium (Sigma-Aldrich, St. Louis, MO), containing 10% heat-inactivated FBS, until confluence. The medium was then replaced with FBS-free F12K, and the cells were stimulated with IL-13 and LTC₄₅ or LTD₄ in the presence or absence of LTRA (pranlukast or montelukast).

Human bronchial SMC were obtained from BioWhittaker (Walkersville, MD) and cultured in SmGM-2 medium. Human alveolar macrophages (AM) were obtained from healthy volunteers by means of bronchoalveolar lavage (BAL).

Measurement of eotaxin in the culture supernatant
The concentrations of eotaxin were measured in the cell supernatant fluids from HFL-1 cells using an ELISA kit (BioSource International) according to the manufacturer’s instructions. The minimum concentration detected by this method was 2 pg/ml.

Eosinophil purification
Eosinophils were isolated from 60 to 80 ml of peripheral venous blood obtained from healthy donors by means of a magnetic cell separation system (MACS, Miltenyi Biotec, Bergisch-Gladbach, Germany) as described by Hansel et al. (21). Venous heparinized blood was diluted 1/1 in PBS before being loaded on 67% Percoll (density, 1.077 g/ml). After centrifugation at 800 × g for 30 min, mononuclear cells were removed, and the erythrocytes in the cell pellet were hypotonically lysed for 60 s. The polymorphonuclear leukocytes thus obtained were incubated for 1 h at 4°C with anti-CD16 beads, and the suspension was placed in a MACS column for final separation. The purity of eosinophils was generally >98%, and the viability was >95%, as judged by trypan blue exclusion. Eosinophils were suspended in RPMI 1640 containing 10% FBS.

RT-PCR analysis
Total RNA was extracted with the use of TRIzol reagent (Life Technologies, Frederick, MD) from HFL-1 cells cultured in 6-well tissue culture plates. The total RNA was reverse transcribed with a ThermoScript RT-PCR System (Life Technologies) according to the manufacturer’s protocol. PCR were performed with a thermal cycler (Takara Bio, Ohtsu, Japan) under the following conditions: 1 cycle at 94°C for 5 min, followed by 30–36 cycles consisting of 1 min denaturation at 94°C, 1 min annealing at 60°C, and 2 min extension at 72°C. The last cycle was extended to 5 min at 72°C. Products were analyzed by 2% (w/v) agarose gel electrophoresis. The sequences of the 5’ sense primers and the 3’ antisense primers synthesized based on published sequence data (22) and used in this study are as follows: CysLT1R, sense, 5’-GACAGCCATGACCCCTTTCC-3’; and antisense, 5’-ATGCACCCAGAAGACCTT-3’ (product size, 514 bp); and 5’-actin, sense, 5’-AAGAGGAGCTCCATCCACC-3’, and antisense, 5’-TACATGGCTGGGTTGTTA-3’ (product size, 234 bp).

Real-time PCR analysis
Reverse transcription was performed using 1 μg of total RNA and oligo(dT) primers in a 20-μl reaction according to the manufacturer’s protocol (PE Applied Biosystems, Foster City, CA). CysLT1R primers and β-actin primers were designed using Primer Express software (PE Applied Biosystems). The following are sequences for CysLT1R and β-actin: forward primer, 5’-GACACACATCGTGGTACACC-3’; reverse primer, 5’-ATACCTCACCACAAACCTGGC-3’; forward primer, 5’-AAGAGGAGCTCCATCCACC-3’; and reverse primer, 5’-TACATGGCTGGGTTGTTA-3’. Real-time PCR was performed on the ABI Prism 7700 sequence detection system (PE Applied Biosystems) by using SYBR green (Roche Diagnostics, Somerville, NJ) as a dsDNA-specific binding dye. The PCR were cycled 40 times after initial denaturation (95°C, 2 min) with the following parameters: denaturation, 95°C, 15s; and annealing and extension, 60°C, 1 min. The threshold cycle (CT) was recorded for each sample to reflect the mRNA expression levels. A validation experiment provided the linear dependence of the CT value for both CysLT1R and β-actin concentration and consistency of ACT (CysLT1R average CT − β-actin average CT) in a given sample at different RNA concentration. Therefore, ΔCT was used to reflect the relative CysLT1R expression levels. To determine the effects of different stimuli on CysLT1R gene expression as compared with unstimulated cells, ΔΔCT was calculated (ΔΔCT = ΔCT stimulus − ΔCT nonstimulated cells). CysLT1R mRNA was indexed to the β-actin using the following formula: 2^−ΔΔCT × 100%. The value of 2^−ΔΔCT was calculated to demonstrate the fold changes of CysLT1R gene expression in stimulated cells as compared with unstimulated cells.

Flow cytometry
The expression of CysLT1R on HFL-1 was analyzed by flow cytometry as described by Thivierge et al. (20). Cells were washed with PBS and fixed in 2% paraformaldehyde for 15 min at room temperature, followed by permeabilization with DAKO IntraStain reagent B for 15 min at room temperature. Cells were resuspended with PBS and labeled with anti-CysLT1R Ab (1:1000) or isotype control Ab for 60 min at 4°C. After washing with cold PBS, the cells were incubated with FITC-conjugated goat anti-rabbit IgG F(ab')₂ (Rockland, Gilbertsville, PA) for 60 min at 4°C. Finally, the cells were washed again and resuspended in PBS. Single-color immunofluorescence analysis of 5000 cells was performed on a FACScan flow cytometer (BD Biosciences, San Jose, CA).

Eosinophil transmigration assay
Chemotaxis assays were performed with the use of a 96-well microchemotaxis chamber (NeuroProbe, Bethesda, MD). Medium alone (negative control) or cell supernatant fluid from HFL-1 cells (37 μl) was added to individual wells of the bottom plate. A polycyvinylpyrrolidone-free polycarbonate framed filter (5-μm pore size) was placed onto the filled bottom plate. The silicone gasket and the upper piece of the chamber were applied, and 50 μl of the eosinophil suspension (2 × 10⁴ cells/ml in F12K) was placed into the upper chambers on top of the membrane (at least n = 3 wells/condition). After 90 min of incubation in humidified air with 5% CO₂ at 37°C, the filter was fixed in methanol and stained with Diff-Quik. Nonmigrated cells on the under side of filter were wiped away with a cotton swab and rinsed with PBS. Eosinophil chemotaxis activity was expressed as the total number of migrated eosinophils counted in 10 high-power fields under a light microscope at ×400 magnification.

Statistical analysis
Data are expressed as means ± SEM. Statistical significance was determined by one-way ANOVA or paired t test. A value of p < 0.05 was considered to indicate statistical significance.

Results
Production of eotaxin by HFL-1
HFL-1 produced a small amount of eotaxin in the culture supernatant spontaneously without stimulation for 48 h. LTC₄₅ alone at concentration from 10⁻¹² to 10⁻⁶ M did not increase the production of eotaxin as compared with control (Fig. 1A). In contrast, IL-13 increased eotaxin production in a concentration-dependent manner (Fig. 1B). The effect of 10 ng/ml IL-13 was significantly augmented by LTC₄₅ in a concentration-dependent manner (Fig. 2). This synergistic effect of LTC₄₅ (10⁻⁷ M) and IL-13 (10 ng/ml) on eosinophil production was suppressed by planlukast and montelukast.

![FIGURE 1. Eotaxin production by HFL-1 in response to treatment with LTC₄₅ (A) or IL-13 (B). HFL-1 cells were cultured with various concentrations of LTC₄₅ or IL-13 for 48 h, and eotaxin concentrations in the culture supernatant were measured. Data are means ± SEM (n = 4). * p < 0.0001 vs untreated cells.](http://www.jimmunol.org/Downloadedfrom)
To determine whether the observed synergy between LTC₄ and IL-13 depends on the priming of HFL-1 by IL-13, we examined the effect of LTC₄ on eotaxin production by HFL-1 prestimulated with 10 ng/ml IL-13 for 24 h. After preincubation of HFL-1 with 10 ng/ml IL-13 for 24 h, the cells were washed twice and further cultured for 24 h in the presence or absence of various concentrations of LTC₄. Data are means ± SEM (n = 4). *p < 0.0001 vs medium alone; †p < 0.001, and ††p < 0.0001 vs IL-13 (10 ng/ml) without LTC₄.

Production of eotaxin by HFL-1 previously stimulated with IL-13

To determine whether the observed synergy between LTC₄ and IL-13 depends on the priming of HFL-1 by IL-13, we examined the effect of LTC₄ on eotaxin production by HFL-1 previously stimulated with IL-13. After preincubation of HFL-1 with 10 ng/ml IL-13 for 24 h, the cells were washed twice and further cultured for 24 h in the presence or absence of various concentrations of LTC₄. LTC₄ prestimulated with 10 ng/ml IL-13 could respond to LTC₄, and eotaxin production was increased in an LTC₄ concentration-dependent manner (Fig. 4). These results suggest that IL-13 can alter the response of HFL-1 to LTC₄.

Expression of CysLT1R mRNA on HFL-1

To investigate the mechanism of increased responsiveness of HFL-1 to LTC₄, the expression of CysLT1R mRNA on HFL-1 was determined by semiquantitative RT-PCR. Unstimulated HFL-1 expressed only low levels of CysLT1R mRNA as compared with the expression levels in human AM or human bronchial SMC (Fig. 5). Both AM and SMC have been reported to express CysLT1R constitutively (23). Real-time PCR revealed that stimulation with IL-13 at a concentration of 10 ng/ml for 24 h significantly up-regulated CysLT1R mRNA expression in HFL-1 (Fig. 6A). This effect of IL-13 was time dependent by 48 h (Fig. 6B).

Flow cytometric analysis of CysLT1R expression

To test whether transcription of CysLT1R mRNA is associated with augmented cellular expression of CysLT1R protein, flow cytometry analysis was performed with the use of a specific anti-CysLT1R Ab. Changes in HFL-1 mRNA levels were paralleled by increased cellular expression of CysLT1R. Only IL-13 at a concentration of 10 ng/ml significantly up-regulated CysLT1R protein expression (Fig. 7). In contrast, no expression of CysLT1R protein was detected on unstimulated cells or cells stimulated with 1 ng/ml IL-13.

Eosinophil chemotactic activity in culture supernatant

To determine whether LTC₄-induced eosinophil sequestration is attributed to LTC₄ itself or to eosinophil chemotactic factors, including eotaxin produced by fibroblasts stimulated by LTC₄, eosinophil chemotactic activity in the culture supernatant was evaluated with the use of anti-eotaxin Ab and a CysLT1R antagonist.
Culture supernatant from HFL-1 stimulated with LTC₄ had higher eosinophil chemotactic activity than did unstimulated control supernatant, and this increased chemotactic activity was abolished by the treatment of HFL-1 with pranlukast (Fig. 8A). Because LTC₄ added to the fresh medium showed equivalent chemotactic activity to that of the culture supernatant of HFL-1 stimulated with LTC₄, and the direct chemotactic effect of LTC₄ on eosinophils was inhibited by pranlukast (Fig. 8B), LTC₄ was considered to directly possess chemotactic activity. A combination of LTC₄ (10⁻⁷ M) and IL-13 (10 ng/ml) showed greater eosinophil chemotactic activity than did either LTC₄ or IL-13 alone (Fig. 8A). Treatment of HFL-1 with pranlukast inhibited the eosinophil chemotactic activity induced by the combination of LTC₄ (10⁻⁷ M) and IL-13 (10 ng/ml) (Fig. 8A). In this case, pranlukast was considered to be effective both in inhibition of eosinophil migration to LTC₄ and in inhibition of eotaxin production from HFL-1.

Furthermore, the addition of anti-eotaxin Ab significantly decreased eosinophil chemotactic activity induced by IL-13 alone or by a combination of LTC₄ and IL-13 (Fig. 9). Nonspecific Ab had no effect on eosinophil chemotactic activity. These findings suggest that, in addition to its own chemotactic effect, LTC₄ can augment eotaxin production by fibroblasts stimulated with IL-13, thereby inducing eosinophil migration.

### Discussion

Our study clearly showed that LTC₄ plays an important role in the pathogenesis of eosinophilic inflammation in vitro. LTC₄ alone had weak chemotactic activity for eosinophils, consistent with previous reports (9). In addition to the direct effect of LTC₄, we showed that LTC₄ could stimulate eotaxin production by IL-13-treated fibroblasts, thereby indirectly inducing eosinophil sequestration. Whereas unstimulated fibroblasts did not respond to LTC₄, coinoculation or preincubation with IL-13 altered the response of fibroblasts to LTC₄. HFL-1 stimulated with IL-13 responded to LTC₄ and produced eotaxin. Such production was inhibited by the CysLT1R antagonists.
We investigated the mechanism of the IL-13-induced increase in HFL-1 responsiveness to LTC4. Unstimulated HFL-1 constitutively expressed low levels of CysLT1R mRNA as compared with levels expressed by human bronchial SMC or AM. IL-13 induced CysLT1R expression at both the transcript and protein levels. This up-regulation of CysLT1R expression by IL-13 was considered responsible for the effect of LTC4 on eosinotaxis produced by HFL-1. Recently, Lynch et al. (18) reported the cloning and characterization of high-affinity cell surface LTD2R (CysLT1R). Expression of CysLT1R mRNA has been identified in bronchial SMC, macrophages, and peripheral blood leukocytes (18, 23). However, to our knowledge, no study has examined CysLT1R expression in fibroblasts. Therefore, we believe that this is the first report describing CysLT1R expression in fibroblasts and regulation of such expression by IL-13. Lynch et al. (18) reported that the in situ hybridization pattern of CysLT1R in normal human lung is characterized by strict localization to SMC and some macrophages. Although the expression of CysLT1R in patients with bronchial asthma remains unclear, fibroblasts might express CysLT1R in the TH2 cytokine-rich milieu.

Several previous reports have examined the regulation of CysLT1R expression. Thivierge et al. (19) showed that IL-5 up-regulates CysLT1R expression and enhances responsiveness to LTD4 in HL-60 cells. They also reported that IL-13 and IL-4 enhance the expression of CysLT1R in monocytes/macrophages (20). These findings suggest that TH2 cytokines, including IL-4, IL-5, and IL-13, have a regulatory role in CysLT1R expression. In our study, IL-13 up-regulated CysLT1R expression on fibroblasts. The inflammatory process associated with asthma is thought to be mediated by TH2 lymphocytes and their cytokines. Elevated expression of IL-13 mRNA and protein has been observed in BAL cells of patients with atopic asthma after allergen challenge (24). In a TH2 cytokine-rich environment, CysLT1R expression on fibroblasts is thought to be up-regulated, promoting the proinflammatory effects of CysLTs.

In practice, inhaled LTD4 or LTE4 has been shown to induce airway eosinophilia in asthmatic subjects (6, 25). However, there is no report about the effect of CysLTs on eosinophilic airway inflammation in nonallergic subjects. In an animal study, LTD4 inhalation did not induce eosinophil sequestration into airway in control nonallergic mice. Conversely, eosinophil airway inflammation was not induced in 5-lipoxygenase knockout mice, even though an asthmatic state was made by OVA sensitization and challenge. However, LTD4 inhalation replenished eosinophil sequestration in 5-lipoxygenase knockout mice to the level of wild-type allergic mice (26). These findings suggest that both TH2 cytokines and CysLTs are essential for eosinophilic inflammation, and they also support our results that LTC4 alone stimulated no eosinotaxis production but it could induce eosinotaxis production with IL-13.

Eotaxin is a C-C chemokine that has potent and specific chemoattractant activity for eosinophils (27–30). Eotaxin plays an important part in the pathophysiology of eosinophilic airway inflammation. A variety of cell types are responsible for eotaxin production, including macrophages, T lymphocytes, bronchial epithelial cells, endothelial cells, and fibroblasts (31–35). However, fibroblasts are considered the major source of eotaxin (34–36). Recently, IL-4 and TNF-α were reported to synergistically promote the secretion of eotaxin in human skin and nasal fibroblasts (14, 34). IL-13 shares many biological properties with IL-4 and has similar effects in human airway epithelial cells (37, 38) and fibroblasts (17). Synergism between IL-4 and TNF-α is ascribed to the finding that transcription factor STAT-6 activated by IL-4 signals and NF-κB activated by TNF-α stimulation bind to an overlapping binding site on the eotaxin promoter proximal region, thereby amplifying response (39). In our study, STAT-6-dependent pathways were also presumably involved in the IL-13-induced up-regulation of CysLT1R expression.

The synergistic effect of IL-13 and LTC4 might not involve a CysLT1R-dependent mechanism. Recently, Panettieri et al. (40) reported that LTD4 augments proliferation of human airway smooth muscle induced by epidermal growth factor (EGF). The precise mechanism underlying this synergy is unknown, but can be explained by convergence between LTD4 signals via G protein-coupled receptor and EGF signals via an EGFR tyrosine kinase. Such signal convergence might participate in the synergism between IL-13 and LTD4, but this possibility is unlikely, because our study showed that eotaxin production was increased not only by simultaneous stimulation with LTC4 and IL-13 but also by stimulation with LTC4 alone after 24-h pretreatment with IL-13.

Pranlukast and montelukast are specific LTRAs that can inhibit CysLT-induced bronchospasm. Recent studies have demonstrated that a single exposure of guinea pigs to aerosolized LTD4 produces a significant accumulation of eosinophils in the lung and BAL fluid, which persists for at least 4 wk after challenge. Pranlukast significantly inhibited both the bronchoconstriction and the immediate (24 h) and sustained (4 wk) eosinophil influx into the airways (41). Pranlukast also reduces infiltration of inflammatory cells, including eosinophils, in the bronchial mucosa of patients with asthma (42). In the present study, LTRAs inhibited LTC4-induced eotaxin production by fibroblasts and reduced eosinophil migration. These findings support the therapeutic usefulness of LTRA for eosinophilic inflammation.

In conclusion, we have shown that IL-13 synergistically augmented eotaxin production by LTC4 via up-regulation of CysLT1R expression on lung fibroblasts and that a CysLT1R antagonist can suppress eosinophilic recruitment to the airway through both direct and indirect mechanisms.

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References


