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Cutting Edge: Selection of B Lymphocyte Subsets Is Regulated by Natural IgM

Nicole Baker and Michael R. Ehrenstein

Natural IgM has a wide range of actions in the immune system. Here we demonstrate that mice lacking serum IgM have an expansion in splenic marginal zone B cells with a proportionately smaller reduction in follicular B cells. The increase in the marginal zone-follicular B cell ratio (and an expansion in peritoneal B1a cells) is fully reversed by administration of polyclonal IgM, but not by two IgM monoclonals. Mice engineered to have a secreted oligoclonal IgM repertoire with an endogenous membrane IgM also exhibited a similar expansion of marginal zone B cells. We propose that natural IgM, by virtue of its polyreactivity, enhances Ag-driven signaling through the B cell receptor and promotes the formation of follicular B cells. These results demonstrate that natural IgM regulates the selection of B lymphocyte subsets. The Journal of Immunology, 2002, 169: 6686–6690.

Natural serum IgM provides a first defense against infection and accelerates the development of the primary immune response (1, 2). Mice deficient in serum IgM (Sμ−) also have an elevated response to T-independent Ags, an expanded B1a cell subset, and an increased propensity to develop IgG autoantibodies (3, 4). Although B1 B cells have been characterized as a major contributor to natural IgM production, the mechanism underlying the elevation in B1 cells when serum IgM is absent is unclear. Splenic marginal zone (MZ) B cells, have been identified as having some similar properties to B1 cells including contributing to IgM secretion. Both MZ and B1 cells have been implicated in the rapid, early response to infection, including the T-independent response, in autoimmune and lymphomagenesis (5, 6).

Recent evidence has suggested that only a minority of newly formed B cells are selected for maturation through positive selection by Ag (7–9). Once the immature B cell has reached the spleen, it can develop into follicular (FO) or MZ B cells, and this decision may be governed by a number of factors including the strength of B cell receptor (BCR) signaling (10), and the drive to make a complete repertoire of natural Abs (6). Moreover, MZ and FO B cell numbers are also affected by other factors such as life span and retention and migration signals. B1 cell development also appears to be dependent on Ag selection (11). Whereas alterations in some molecules that affect B cell signaling result in parallel changes in B1 and MZ B cells, e.g., Aiolos (12), other examples demonstrate that these two B cell subsets are not always regulated in tandem as observed in CD22− mice in which B1 cells are increased but MZ B cells are reduced (13, 14).

Here we show that splenic MZ B cells, as well as B1 cells, are expanded in mice deficient in serum IgM, with a corresponding reduction in splenic FO B cells, and that replacement with polyclonal, polyreactive IgM, but not monoclonal or oligoclonal IgM, reverses the changes in these B cell subsets.

Materials and Methods

Mice

The generation of Sμ− mice has been described previously (2). Mice were backcrossed onto C57BL/6 mice for six generations and kept under specific pathogen-free conditions. Litter-matched controls were used for all experiments.

Generation of Vμ1s-transgenic mice

The Vμ1 construct (gift from Dr. M. Neuberger, Laboratory of Molecular Biology, Cambridge, U.K.) encodes a Vμ1 (α allotype) gene that binds to anti-4-hydroxy-3-nitrophenenacetyl under control of a Vμ1 promoter (15). It was modified by restriction enzyme digestion so that the membrane exon of the transgene was deleted but the secretory exon remained intact (Fig. 2A). The resulting 11-kb construct Vμ1s was used for pronuclear injection, and transgenic mice were generated by standard techniques. The mice were tested for the presence of the transgene by Southern blotting and PCR and were backcrossed six times onto C57BL/6 mice and subsequently onto Sμ− mice.

Flow cytometric analyses

Flow cytometric analyses were performed using standard techniques, stained cells were analyzed on FACScan, and files were plotted using CellQuest software (BD Biosciences, San Jose, CA). The MZ:FO ratio was calculated using absolute numbers of B cells in the CD21highCD23low MZ compartment and the CD21midCD23high FO compartment.

Injections and immune responses

Sμ− and litter-matched controls were injected i.p. on six occasions at regular intervals for 2 wk with 200 μg/mouse of a purified monoclonal (either TEP-183, unknown specificity [here designated monoclonal A] or MOPC-104E, specific for α-1,3-glucose [here designated monoclonal B]; Sigma-Aldrich, St. Louis, MO) or polyclonal IgM Ab (Rockland, Gilbertsville, PA). To induce a T-independent response, mice were immunized i.p. with 5 μg of phosphorylcholine-Ficoll (PC-Ficoll; Bioresearch Technologies, Novato, CA) in PBS. Mice were bled at regular intervals during the 2 wk after the immunization.
ELISA

Ig and Ag ELISAs were performed as previously described (2) using anti-class- or -allotype reagents (BD Pharmingen, San Diego, CA). The myeloperoxidase ELISA was performed with a Capita myeloperoxidase ELISA kit (Trinity Biotech, Bray, Ireland).

Immunofluorescence analysis of tissue sections

Standard immunofluorescence staining procedures were used on frozen spleen sections stained with FITC-Conjugated anti-mouse IgM and Texas Red conjugated anti-mouse IgD (BD Pharmingen). Sections were viewed using a confocal microscope (Axiovert 100; Zeiss, Oberkochen, Germany).

Results

MZ B cells are expanded in Sµ− mice

Mature splenic B cells can be divided into MZ and FO B cells and can be distinguished by surface markers using FACS. MZ B cells (CD21<sup>high</sup>/CD23<sup>low</sup>) are expanded in 4-mo-old Sµ− mice, whereas FO B cells are reduced (Fig. 1A). Whereas the increase in MZ B cell numbers is 2.5-fold, the reduction in follicular B cells is less (a 25% reduction) based on proportion of absolute cell numbers for individual mice. Wild-type (WT) and Sµ− mice have the same number of splenic B220<sup>+</sup> B lymphocytes (WT, 1.27 ± 0.26 × 10<sup>7</sup>; Sµ−, 1.12 ± 0.17 × 10<sup>7</sup>). This contrasts with the cellular changes observed after a T-independent response to PC-Ficoll where MZ B cell numbers increase in both WT and Sµ− mice (compare immunized with naive) whereas absolute FO B cell numbers are unchanged. In older mice (12 mo) the MZ:FO B cell ratio is increased in WT mice and the difference in MZ/FO B cell ratio between Sµ− and WT mice is less striking at this age (Fig. 1, A and B). Spleen sections taken from naive 4-mo-old Sµ− mice stained with anti-IgM/anti-IgD demonstrated a larger IgM<sup>bright</sup>/IgD<sup>dim</sup> MZ area on the periphery of the follicle compared with WT controls, further demonstrating the expansion in MZ B cells in Sµ− mice (Fig. 1C). There is also an increase in the number of splenic CD21<sup>high</sup>/CD23<sup>low</sup> B cells which predominantly consist of newly formed B lymphocytes but would also contain splenic B1 cells. Although the numbers of splenic B1 cells are small compared with B2 cells, there is an increase in B1 cells (CD5<sup>+</sup>) in Sµ− mice (4.5 ± 0.7% within the splenic B lymphocyte gate) compared with WT mice (2.26 ± 0.3%), consistent with previous observations (1).

Polyclonal, but not monoclonal or oligoclonal, IgM fully reverses the MZ B cell expansion

To determine whether polyclonal IgM was critical in governing the B cell compartments, two approaches were used. The first was to generate a transgenic mouse line (Vµ<sub>1s</sub>) using absolute cell numbers in individual cell fractions. MZ and FO were formed.

Histogram showing splenic MZ:FO B cell ratios calculated from 10 mice and are gated on B220<sup>+</sup>– cells for antinuclear Ab staining demonstrated that the polyclonal and 12 mo) or 2 wk after immunization (4 mo) with PC-Ficoll, stained with F(ab)2 anti-B220, anti-CD21, and anti-CD23. The probe was to generate a transgenic mouse line (Vµ<sub>1s</sub>) mice back to that observed in control mice as shown by FACS (Fig. 2B). The serum concentration of these preparations during the 2-wk period were identical (Fig. 2C).

Effects of IgM reconstitution on B1 cells

Consistent with previous observations, peritoneal B1a cells were increased in Sµ− mice (1, 2). This expansion in B1a cells was reversed by polyclonal IgM and to a lesser extent by monoclonal A but not in those mice expressing the Vµ<sub>1s</sub> transgene or the mice that had received the monoclonal IgM B preparation (Fig. 3).

Polyclonal IgM is polyreactive

To determine whether the reactivity of the different IgM preparations was associated with their ability to influence MZ and B1 cells, binding to four different Ags was assayed. Analysis of the binding of these IgM preparations to the Ags tested confirmed the multireactivity of polyclonal IgM which bound all four Ags strongly and monoclonal A bound two Ags strongly, whereas monoclonal B bound only one Ag (Fig. 4). Analysis using HEP-2 cells for antinuclear Ab staining demonstrated that the polyclonal preparation and monoclonal A but not monoclonal B were positive for antinuclear Ab staining.
FIGURE 2. Effects of IgM reconstitution on B cell subsets. A, Transgene construct of V\(\mu\)1s. The secretory \(\mu\) tailpiece (tp) and the \(\mu\) polyadenylation site are located within the V\(\mu\)1s construct; the \(\mu\) constant region (C\(\mu\)M) and the \(\mu\) polyadenylation have been removed from the original V\(\mu\)1 construct and are shown for information only. B, FACS analysis of spleen cells taken from mice expressing the V\(\mu\)1s transgene using IgM allotype-specific markers. The IgM allotype of the V\(\mu\)1 transgene and the targeted S\(\mu\)- allele are \(\alpha\), whereas the BL/6 allotype is b. C, IgM serum concentration in S\(\mu\)-/N\(\mu\)1s mice and in mice that had received a polyclonal (poly) preparation or two monoclonal (mono A, mono B) preparations of purified IgM measured 2 days after the final injection of the different IgM preparations (three mice per group). Samples taken 1 or 2 days after each injection were not significantly different from results shown (data not shown).

Discussion
The major finding of this work is that a selective deficiency of serum IgM leads to an increase in splenic MZ B cells with a proportionally smaller reduction in FO B cells. Furthermore, the fact that the increased MZ:FO B cell ratio is fully reversed by adding back polyclonal, but not monoclonal or oligoclonal, IgM suggests that a full repertoire of IgM molecules is required to regulate these B cell subsets. The difference between these preparations of IgM on splenic B cell subsets may be due to the multiple Ag reactivity of polyclonal IgM because this preparation bound to all the Ags tested and had the greatest effect on the MZ:FO B cell ratio. Monoclonal B had the least reactivity of the IgM preparations and no effect on the MZ:FO ratio. Administration of polyclonal IgM also reversed the expansion in peritoneal B1 cells previously documented (1, 2). Although there are a number of mechanisms that might explain the increased MZ:FO B cell ratio seen in S\(\mu\)-, as discussed below, we favor the hypothesis that IgM is acting as an adjuvant to deliver enhanced (self) Ag-driven signals to the BCR.

It has been proposed that the balance between splenic MZ and FO B cells is regulated by B cell signaling and that increasing clonal B cell signals favors the formation of FO over MZ B cells (10). The pentameric structure of IgM, in contrast to IgG, can potentially yield a high avidity for Ag despite a low Ag-specific affinity. Moreover, one molecule of IgM can activate complement, whereas two IgG molecules are required; this may lead to enhanced complement coating of selected self Ags when complexed to IgM, resulting in more efficient engagement of coreceptor signals (16, 17). Consequently, BCR signaling could be reduced and MZ B cells preferentially formed in the absence of IgM, despite normal levels of serum IgG. A similar process may apply to B1 cells, which are also sensitive to Ag drive (11), because polyclonal IgM also reversed the expansion of this B cell subset. The MZ:FO B cell ratio increases in mice as they age (this work and Ref. 18), which may be explained by defective signaling through the BCR which is also associated with aging (19), because MZ B cells are preferentially formed when BCR signals are reduced (10). Thus, the less striking difference between older S\(\mu\)- and control mice could be attributed to the already dampened BCR signaling associated with age.

The expansion in MZ B cell numbers in S\(\mu\)- mice may be explained by a number of mechanisms apart from B cell signaling. Mice bearing Ig chain transgenes with a variety of Ag specificities have increased MZ B cell compartment. Because these mice have a restricted Ab repertoire, it has been proposed that the drive to make a complete repertoire increases MZ B cell numbers (6). Clearly, in S\(\mu\)- mice, the secreted IgM repertoire is absent whereas a full membrane IgM repertoire is present. If this proposed drive arises from the serum Ab repertoire then one would predict, as is observed here, that in S\(\mu\)- mice MZ B cell numbers would be increased and that polyclonal but not monoclonal IgM would reverse the changes. The transgenic mouse V\(\mu\)1s crossed onto the S\(\mu\)- mouse also has a restricted secreted IgM repertoire with a normal membrane BCR repertoire, supporting the notion that it is the secreted repertoire that is important in governing B1 and MZ B cell numbers. This line of reasoning, however, presumes that the serum IgG repertoire does not constitute a full Ab repertoire. A loss of diversity in the Ab repertoire is also observed in aging...
feedback mechanism that simply senses the IgM concentration. However, polyclonal IgM may have an advantage even in this situation if a putative FcµR binds preferentially to the altered confirmation arising out of IgM complexed with Ag. An FcµR has been described, although not with these properties (21). However, a recent report has demonstrated that serum IgM is a ligand for CD19 (22), and B1 and MZ B cells are affected by alterations in CD19 expression (8, 23).

One apparent paradox of these observations is the short time (2 wk) required to reverse the MZ:FO ratio coupled with the evidence that MZ and FO B cells have a relatively long life span (24). This long life span, however, is somewhat dependent on the numbers of immature B cells entering the mature B cell pool. The finding that newly formed B cells increase in Su- mice may suggest either that these cells are being retained in this pool longer or that there are more immature B cells exiting from the marrow. It is known, and confirmed here, that MZ B cell numbers change during a 2-wk period after encountering T-independent Ags (6). Thus, changes in these B cell subsets could occur through a variety of mechanisms including alterations in cell proliferation or apoptosis, or by interchange between the MZ and FO B cell pools, although how IgM influences these processes remains to be investigated. It is tempting to speculate that IgM, through regulation of these B cell subsets, may have a role in suppressing the autoimmunity that occurs in mice strains where the MZ and B1 B cells are expanded.

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References


