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Dendritic Cells Pulsed with Fungal RNA Induce Protective Immunity to Candida albicans in Hematopoietic Transplantation

Angela Bacci,* Claudia Montagnoli,* Katia Perruccio,† Silvia Bozza,* Roberta Gaziano,‡ Lucia Pitzurra,* Andrea Velardi,† Cristiana Fe’ d’Ostiani,* Jim E. Cutler,§ and Luigina Romani**

Immature myeloid dendritic cells (DC) phagocytose yeasts and hyphae of the fungus Candida albicans and induce different Th cell responses to the fungus. Ingestion of yeasts activates DC for production of IL-12 and Th1 priming, while ingestion of hyphae induces IL-4 production and Th2 priming. In vivo, generation of antifungal protective immunity is induced upon injection of DC ex vivo pulsed with Candida yeasts but not hyphae. In the present study we sought to determine the functional activity of DC transfected with yeast or hyphal RNA. It was found that DC, from either spleens or bone marrow, transfected with yeast, but not hyphal, RNA 1) express fungal mannoproteins on their surface; 2) undergo functional maturation, as revealed by the up-regulated expression of MHC class II Ags and costimulatory molecules; 3) produce IL-12 but no IL-4; 4) are capable of inducing Th1-dependent antifungal resistance when delivered s.c. in vivo in nontransplanted mice; and 5) provide protection against the fungus in allogeneic bone marrow-transplanted mice, by accelerating the functional recovery of Candida-specific IFN-γ-producing CD4+ donor lymphocytes. These results indicate the efficacy of DC pulsed with Candida yeasts or yeast RNA as fungal vaccines and point to the potential use of RNA-transfected DC as anti-infective vaccines in conditions that negate the use of attenuated microorganisms or in the case of poor availability of protective Ags. The Journal of Immunology, 2002, 168: 2904–2913.

In humans, Candida albicans is the most frequently isolated fungal pathogen (1). The delicate balance between the host and this otherwise harmless commensal fungus turn into a parasitic relationship, resulting in the development of severe infections. Invasive candidiasis is a significant cause of morbidity and mortality in differently immunocompromised patients, including those undergoing bone marrow transplantation (BMT).3 It is recognized that the nature and extent of the impairment of host defense influence the manifestation and severity of infection (1).

Clinical evidence and experimental data indicate that both the innate and the adaptive immune systems regulate resistance to Candida infections (3–5). In murine experimental models of infection, it has been demonstrated that Th cell reactivity plays a central role in regulating immune responses to the fungus, Th1 reactivity being responsible for resistance and Th2 reactivity being associated with susceptibility (6). The development of protective anticyndal Th1 responses requires the concerted actions of several cytokines, including IFN-γ and IL-12, in the relative absence of Th2 cytokines, such as IL-4 and IL-10, which inhibit development of Th1 responses (6).

However, the fungus is not a mere passive participant in the infectious process, and a hypothetical set of virulence factors has been attributed to it (7). Among these, the ability to reversibly switch between budding yeast to the filamentous growth form or hypha, all of which can be found in infected tissues (1), is thought to be important for virulence (8–10).

Recent evidence indicates that dendritic cells (DC) are uniquely capable of decoding the fungus-associated information required to elicit the qualitative nature of the adaptive immune response (11). DC finely discriminated between the two forms of C. albicans in terms of type of immune responses elicited. By the production of IL-12 and IL-4 in response to the nonvirulent and virulent forms of the fungus, DC were uniquely capable of Th priming and education in vitro and in vivo (11). This finding is particularly relevant in candidiasis, because the fungus behaves as a commensal as well as a true pathogen of skin and mucosal surfaces (1), known to be highly enriched for DC.

DC are uniquely specialized to initiate T cell immunity in vitro and in vivo (12). When delivered in vivo, DC charged with peptide Ags, tumor lysates, or viral vectors that encode the relevant Ags induce immune responses, which include protective and therapeutic immunity to tumors and pathogens in animals (11, 13–18) and clinical antitumor responses in humans (19–22).

Studies have shown that DC transfected with tumor RNA could also serve as potent cancer vaccines (23–25). DC transfected with chicken OVA RNA stimulate primary anti-OVA CTL responses in vitro. Moreover, vaccination of mice with OVA RNA-transfected DC has been used to treat OVA-positive tumor metastases to lung.

1 Abbreviations used in this paper: BMT, bone marrow transplantation; DC, dendritic cell; BM, bone marrow; BM-DC, BM-derived DC; SP-DC, spleen-derived DC; RAPD, random amplified polymorphic DNA; DOTAP, N-[1-(2,3-dioleoyloxypropyl]-N,N-trimethylammonium methylsulfate; MFI, median fluorescence intensity.
These findings are particularly relevant, as they would extend vaccination to conditions of limited availability of protective Ags. However, whether or not induction of antimicrobial immunity with DC transfected with pathogen-derived RNA could represent a strategy of DC-based therapy in infections remains to be determined.

We have already shown that DC pulsed with viable Candida, but not hyphae, were capable of inducing protective Th1 immune responses when adoptively transferred in vivo (11). In the current study we tested whether transfrecting DC with Candida yeast RNA is an effective way to induce antifungal protective immunity in vivo. Ex vivo transfrected DC were adoptively transferred into recipient mice, and parameters of infection and immunity to it were evaluated upon the i.v. challenge with the fungus.

We also used a very stringent experimental system to assess the priming capacity of transfected DC, which includes the infusion of DC in an allogeneic T-depleted BMT experimental model (26). In this model, autologous reconstitution of host stem cells is greatly reduced to the benefit of a long-term, donor-type chimerism in >95% of the mice and low incidence of graft-vs-host disease (27–29). We found here that DC transfected with yeast, but not hyphal, RNA were capable of stimulating Candida-specific Th1 immunity that conferred antifungal resistance to susceptible mice, either untransplanted or after allogeneic hematopoietic transplantation.

Materials and Methods

Animals

Female, 8- to 10-wk-old, inbred C3H/HeJ and hybrid (BALB/c × DBA/2)F1 (CD2F1) mice were obtained from Charles River Breeding Laboratories (Calco, Italy). Breeding pairs of homozygous IFN-γ/H9253 mice were obtained from Charles River Breeding Laboratories (Calco, Italy). Breeding pairs of C57BL/6J mice were obtained from Charles River Breeding Laboratories (Calco, Italy). Breeding pairs of homozygous IFN-γ/H9253 mice were obtained from Charles River Breeding Laboratories (Calco, Italy). Breeding pairs of homozygous IFN-γ/H9253 mice were obtained from Charles River Breeding Laboratories (Calco, Italy). Breeding pairs of homozygous IFN-γ/H9253 mice were obtained from Charles River Breeding Laboratories (Calco, Italy). Breeding pairs of homozygous IFN-γ/H9253 mice were obtained from Charles River Breeding Laboratories (Calco, Italy).

Irradiation

C3H/HeJ mice were exposed to a single, lethal dose of 9 Gy from a gamma beam 150A, 60Co source (Clinac 600C Varian, Cenersus, Milan, Italy) with focus to skin distance of 75 cm and 0.7 Gy/min dose rate. Unless BM-transplanted, mice die within 14 days.

C. albicans strains and isolation of fungal RNA

The origin and characteristics of C. albicans highly virulent and live vaccine strains used in this study have already been described (11). Total RNA was isolated from actively growing yeasts and hyphae as described (30). Briefly, cells were disrupted by repeated cycles of thawing and freezing on dry ice, cells were disrupted by repeated cycles of thawing and freezing on dry ice, cells were disrupted by repeated cycles of thawing and freezing on dry ice, 75°C for 30 min at 37°C, the enzymes were inactivated by incubating samples with May-Grünewald-Giemsa reagents (Sigma-Aldrich) before analysis.

Preparation of T cell-depleted BM cells

Donor BM cells were prepared by differential agglutination with soybean agglutinin, as described (27). T cell-depleted soybean agglutinin-positive cells (containing <1% of contaminating T cells on FACs analysis) were injected at the concentration of ≥2 × 105/ml into recipient mice i.v. According to previous studies (27–29), >95% of the mice survived showing stable, donor-type hematopoietic chimerism, as revealed by donor-type MHC class I Ag expression on cells from spleens.

Purification, Candida pulsed, and culture of DC

DC were purified from spleens (SP-DC) by magnetic cell sorting with MicroBeads (Miltenyi Biotec, Bergisch Gladbach, Germany) conjugated to hamster anti-mouse CD11c mAbs (clone N-418), as described (11). Briefly, after overnight plastic adherence to remove macrophages, 106 collagenase D (Sigma-Aldrich, St. Louis, MO)–treated nonadherent spleen cells were reacted with 100 μl of CD11c MicroBeads before magnetic separation. BM-derived DC (BM-DC) were prepared as described (31). Briefly, BM-DC, obtained from femurs of mice, were depleted of red cells and seeded (2 × 106/ml) in six-well plates (Falcon; BD Labware, Lincoln Park, NJ) in 3 ml IMDM (Life Technologies, Milan, Italy) with 10% FCS, 5 mm 2-ME, 50 μg/ml gentamicin sulfate, 2000 U/ml GM-CSF (Sigma-Aldrich), and 1 × 106 U/ml IL-4 (BD Pharmingen, San Diego, CA) for 6 days. On day 3 of culture, 106/ml cells were washed and fresh medium containing GM-CSF and IL-4 was added. On day 6 of culture, DC were isolated by transferring nonadherent cells to new culture plates and incubating at 37°C for at least 2 h. Consistent with previous reports (31, 32), ≥90% SP-DC (at ~1% yield recovery) routinely express high levels of CD11c integrin, while BM-DC (at ~10% of the BM population) uniformly showed low level of CD11c staining. More than 90% of BM-DC did not react with the 2.43 anti-CD8α mAb, while 25–30% of SP-DC expressed CD8α (data not shown). For pulsing, 2 × 106 DC were exposed to live yeasts or hyphae (at DC:yeast ratio of 1:1 and 1:10, respectively) for 2 h after addition of 2.5 μg/ml amphotericin B (Sigma-Aldrich) to prevent Candida overgrowth, as described (11). Pulsing of DC with RNA was performed in serum Opti-MEM medium (Life Technologies, Italy) (31). DC were washed twice in Opti-MEM and reseeded in Opti-MEM medium at 2–5 × 104 cells/ml and added to 15-ml polystyrene tubes (Falcon). The cationic lipid, N-[1-(2,3-dioléoléoyloxypropyl)]-N,N,N-trimethylammonium methysulfate (DOTAP; Boehringer Mannheim, Mannheim, Germany) was used to deliver RNA into cells (33). RNA (25 μg in 250 μl Opti-MEM medium) and DOTAP (50 μg in 250 μl Opti-MEM medium) was mixed in 12 × 75-mm polystyrene tubes at room temperature for 20 min at the RNA:DOTAP ratio of 1:2. The complex was added to DC in a total volume of 2 ml and incubated at 37°C in a water bath with occasional agitation for 2 h. After pulsing, the cells were washed and left for an additional 24 h in culture before FACS analysis, measurement of cytokine in the supernatants, and adoptive transfer. Some RNAs were treated with 400 μg/ml proteinase K (Sigma-Aldrich) or 100 U/ml RNase A (Sigma-Aldrich) or RNase-free DNase I (Life Technologies, Gaithersburg, MD) for 30 min at 37°C. The enzymes were inactivated by incubating samples at 65°C for 15 min.

Adoptive immunization, fungal challenge, and assessment of protection

DC (3 × 105/each injection) were injected either s.c. in 20 μl of PBS or i.v. in 0.5 ml of PBS following different protocols, as described in Results. In BM-transplanted recipients, DC were administered s.c. twice, 1 and 7 days after BMT. For infection, a week after the last DC administration, mice were i.v. infected with 106 C. albicans cells of the live vaccine or the virulent strain in 0.5 ml of PBS as described (29). After infection with the virulent strain was done 14 days after the primary infection with the live vaccine strain. Resistance to infection was assessed by quantifying the number of CFU (mean ± SE) per organ and production of Th1 and Th2 cytokines in culture supernatants of Ag-stimulated splenocytes (106 splenocytes stimulated with 105 heat-inactivated C. albicans cells for 48 h), and by enumerating the IFN-γ-producing cells, by ELISPOT assay, as described (11). Total and differential white blood cell counts were done by hemocytometry and by staining blood smears from transplanted mice with May-Grünwald Giemsa reagents (Sigma-Aldrich) before analysis.

Flow cytometry

For DC analysis, cell surface phenotype was assessed with the following mAb reagents: FITC-conjugated mAb anti-CD11c (N418), anti-CD8α (2.43), anti-IA-α (34-5-3), anti-CD80 (1G10), and anti-CD86 (GL1) (all from BD Pharmingen). Unrelated isotype-matched mAbs were used as control. For assessment of C. albicans protein expression, DC pulsed with viable yeasts or hyphae or transfected with RNA from yeasts or hyphae were sequentially reacted with the IgM B6.1 mAb, specifically reacting to the phosphomannan component of the fungal cell wall, β-1,2-mannanotriose (34, 35) or with polyclonal antiserum from vaccinated mice (36) and the FITC-conjugated goat anti-mouse IgA plus IgG plus IgM (H + L) (EuroClone, Milan, Italy). Before all labeling experiments, FcR blocking was performed by incubating cells with 5% normal mouse serum. For chimerism assessment in BMT, the analysis was done as described (26). For lymphocyte analysis, splenocytes were reacted with FITC-conjugated anti-
CD4+ cells (L3T4, clone GK1.5), anti-CD8α (Ly-2, clone 53-6.7), and anti-SE6NK (clone SE6). Staining with PE-conjugated anti-CD44 (Pgp-1, clone IM7) or anti-CD25 (7D4) mAb (BD Pharmingen) was done on gated CD4+ cells. Cells were analyzed with a FACScan flow cytometer (BD Biosciences, Mountain View, CA). Nonspecific binding was excluded from analysis by accepted procedures involving propidium iodide and narrow forward-angle light scatter gating. Control staining of cells with irrelevant Ab was used to obtain background fluorescence values. Data were evaluated both as the percentage of positive cells and as median fluorescence intensity (MFI).

Cytokine assays

The levels of IL-2, IL-4, IL-6, IL-10, and IL-12 p70 in culture supernatants were determined by ELISA (R&D Systems, Space Import-Export, Milan, Italy). Cytokine titers were calculated by reference to standard curves constructed with known amounts of recombinant cytokines (from BD Pharmingen). The detection limits of the assays were <5 pg/ml for IL-2, <3 pg/ml for IL-4, <15 pg/ml for IL-6, <8 pg/ml for IL-10, and <16 pg/ml for IL-12 p70.

ELISPOT assay

IFN-γ-producing cells were enumerated by ELISPOT assay, as described (37). Briefly, freshly isolated splenocytes or purified CD4+ T cells were cultured (1 × 10^5-1 × 10^6 cells/well) in complete medium (RPMI 1640 with 10% FCS, 50 μM 2-ME, and 50 μg/ml gentamicin sulfate) for 18 h in 96-well plates previously coated with rat anti-murine R4-6A-2 mAb. Biotinylated AN-18.17.24 mAb was used as the detecting reagent, avidin-alkaline phosphatase conjugate (Vector Laboratories, Burlingame, CA) was used as the enzyme, and 5-bromo-4-chloro-3-indolyl phosphate-toluidine salt (Life Technologies, Gaithersburg, MD) was used as the substrate. Results were expressed as the mean number of IFN-γ-producing cells (± SE) per 10^6 cells, calculated using replicates of serial 2-fold dilutions of cells.

Rapid-PCR

Fungal RNA was isolated as described above. SP-DC transfected with yeast or fungal RNA were subjected to RNA extraction by the guanidium thiocyanate-phenol-chloroform procedure, as previously described (11). Before cDNA synthesis, all RNA samples were treated with RNase-free DNase I (Life Technologies). cDNA was synthesized from 1 μg of total RNA according to the manufacturer’s directions (Superscript Moloney murine leukemia virus reverse transcriptase; Life Technologies). cDNAs were amplified with random primers derived from the M13 phage core sequence (5'-GAGGGTGGCGGTTCT-3'), known to amplify hypervariable interrepeat DNA sequences from Candida strains (38). Amplifications were performed in volumes of 25 μl containing 10 ng of CDNA, 10 mM Tris-HCl (pH 8.3), 50 mM KCl, 1.5 mM MgCl2, 0.2 mM each dATP, dCTP, dGTP, and dTTP, 3 mM magnesium acetate, 10 ng of primer, and 1.5 U of AmpliTaq DNA polymerase. The PCR was performed in a PerkinElmer thermal cycler (model 480; PerkinElmer, Wellesley, MA) with an initial denaturation of 97°C for 3 min, followed by 40 cycles of 20 s at 93°C, 60 s at 50°C, and 20 s at 72°C, and a final cycle of 5 min at 72°C. Amplification products were separated by electrophoresis in 1.5% agarose gels in 1× Tris-acetate-EDTA buffer for 13 h at 2 V/cm. A 100-bp ladder (New England Biolabs, Cebio, Milan, Italy) was run in parallel for approximate PCR product band sizing. Amplification products were analyzed by 1.5% agarose gel electrophoresis and visualized by ethidium bromide staining.

Statistical analysis

Student’s t test was used to determine significance of values among experimental groups (significance was defined as p < 0.05). In vivo groups consisted of six to eight animals. The data reported were pooled from three to five experiments.

Results

Maturation and cytokine production by DC upon exposure to Candida yeasts or transfected with yeast or hyphal RNA

To assess whether DC undergo functional maturation upon exposure to viable yeasts or RNA from yeasts or hyphae, the surface expression of MHC class II Ags and of CD80 and CD86 costimulatory molecules was assessed in SP-DC and BM-DC exposed to either type of stimuli. The results showed that the expression of

![Figure 1](http://www.jimmunol.org/DownloadedFrom)

**FIGURE 1.** Murine DC undergo maturation upon exposure to Candida albicans yeasts or to RNA from yeasts or hyphae. SP-DC or BM-DC were generated as described in Materials and Methods from CD2F1 mice and puleased with viable yeasts (at DC:fungi ratio of 1:1) or 25 μg total RNA plus 250 μl DOTAP for 2 h at 37°C. Pulsing with viable yeasts was stopped with amphotericin B. Cells were assessed 22 h later for MHC class II Ags, CD80, and CD86 costimulatory Ag expression by immunofluorescence analysis with FITC-conjugated mAb to I-A (clone 34-5-3), to CD80 (clone IGI0), and to CD86 (clone GL1) molecules. Unrelated isotype-matched mAbs were used as a control (gray lines). Analysis was performed on a FACScan. Numbers indicate the MFI values.
class II Ags and of costimulatory molecules was greatly enhanced in both types of DC upon exposure to yeasts or yeast RNA (Fig. 1). Treatment of yeast RNA with DNase or proteinase did not abolish the induction of costimulatory molecules, as did treatment with RNase (Fig. 2), a finding suggesting that sensitization of DC was mediated by RNA.

In contrast, transfecting with hyphal RNA did not result in upregulated expression of costimulatory molecules (Fig. 1), a finding suggesting that maturation of DC is a specific effect of yeast RNA and not a consequence of a nonspecific effect of the transfection method itself. The inability of hyphal RNA to sensitize DC was not due to a defective entry of RNA into the cells, as both yeast and hyphal RNA could be detected inside the cells by random amplified polymorphic DNA (RAPD) analysis (Fig. 3). An 867-bp band that was similar in size to that amplified in fungal cDNAs (Fig. 3) or in genomic DNAs from Candida strains (38) was detected in DC transfected with either yeast or hyphal RNA. As higher amounts of hyphal RNA still failed to up-regulate costimulatory molecule expression on DC (data not shown), all together, these data suggest that the induction of DC maturation is a property of yeast RNA.

We have already shown that myeloid DC produce opposite sets of Th cytokines upon exposure to yeasts (IL-12 p70 and IL-6) or hyphae (IL-4) (11). In this study we found that both types of DC produce IL-12 p70 and IL-6, but not IL-4 or IL-10, upon exposure to either yeasts or yeast RNA (Table I). Therefore, these data suggest that both live yeasts and yeast RNA are effective stimuli capable of inducing functional maturation and cytokine production by myeloid DC.

Expression of fungal Ags on DC upon exposure to Candida cells or Candida RNA

To assess whether effective transduction of fungal RNA occurs in DC after RNA transfection, we measured the surface expression of fungal proteins by reacting cells with a polyclonal antiserum from vaccinated mice, previously shown to contain Candida-specific Abs (36). We also assessed the levels of expression of mannoproteins, known to induce protective immune responses against disseminated candidiasis (39), with the use of the B6.1 mAb directed to the β-1,2 mannotriose of the phosphomannan protein complex (34, 35). The results show that DC stained positive with either the polyclonal or the mAbs after pulsing with viable yeasts. DC transfected with yeast RNA stained positive with polyclonal Abs and less with the mAb (Fig. 4). In contrast, DC pulsed with hyphae or transfected with hyphal RNA did not show positive staining with either type of Abs. Therefore, these data suggest that the transduction of yeast RNA in DC is responsible for the surface expression of fungal Ags, which may include those endowed with protective efficacy in vivo.

Table I. Production of cytokines by SP- or BM-DC upon exposure to Candida albicans yeasts or RNA from yeasts or hyphae

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<td>IL-12 p70</td>
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*SP- and BM-DC were obtained from CD2F1 mice as described in Materials and Methods. DC (2 × 10^6 cells) were pulsed with either live Candida yeasts (at DC: fungus ratio of 1:1) or 25 μg fungal RNA in 250 μl of DOTAP for 2 h at 37°C. After 2 h of pulsing, amphotericin B was added to the cultures of DC pulsed with viable yeasts, and the cells were cultured for an additional 22 h before measurement of cytokine contents in the culture supernatants. <. Below the detection limit of the assays.

**Cytokine levels (measured in picograms per milliliter ± SE) as determined by specific ELISA.
Immunization with yeast RNA-transfected DC induces Th1-mediated antifungal resistance

DC pulsed with *Candida* yeasts, but not hyphae, were capable of inducing Th1 priming and antifungal resistance when adoptively transferred into naive recipients (11). To assess whether DC transfected with yeast RNA also induce antifungal resistance in vivo, SP-DC or BM-DC were exposed to yeasts or to RNA from yeasts or hyphae in vitro and injected in vivo into naive recipients following different experimental protocols, which include different routes and timing of DC administration. Mice were subsequently infected i.v. with virulent *C. albicans* and fungal growth in the kidneys was recorded a week after the infection. The results show that the fungal growth was significantly decreased in mice who had received either type of yeast-pulsed DC administered s.c. 14 days before challenge (Fig. 5). However, consistent with previous data (11), the fungal growth was even more restrained when DC were administered twice, 14 and 7 days before the infection. In contrast, no effect was observed when DC were given 14 days before by the i.v. route. Results with yeast or hyphal RNA-transfected DC showed that the adoptive transfer of yeast, but not hyphal, RNA-transfected DC s.c., twice, also resulted in a significant inhibition of fungal growth (Fig. 5). To understand whether, similar to what had been observed with yeast-pulsed DC (11), the antifungal resistance correlates with the priming of Th1 cells, the production of Th1 (IFN-γ) and Th2 (IL-4 and IL-10) cytokines was evaluated in the spleens of mice who had received multiple s.c. injections of RNA-transfected SP- or BM-DC. The results (Fig. 6) show that production of IFN-γ was higher and that of IL-4/IL-10 was lower in mice who had received multiple s.c. injections of yeast RNA-transfected DC as compared with either mice who had received DC transfected with hyphal RNA or control mice not receiving DC. Together, these data suggest that yeast RNA-transfected DC, similar to yeast-pulsed DC, are capable of inducing Th1 priming and antifungal resistance in vivo, and that the priming capacity is strictly dependent on mode of DC delivery in vivo.

Yeast RNA-transfected DC induce antifungal resistance in allogeneic T-depleted BM-transplanted mice

In a mouse model of allogeneic T-depleted BMT, we have recently shown that susceptibility or resistance to candidiasis correlates with the temporal occurrence of Th2 and Th1 cell responses, with Th2 reactivity accounting for susceptibility to the infection in the early engraftment period (26). To assess whether the infusion of *Candida*-pulsed DC would accelerate Th1 cell recovery and antifungal resistance in transplanted mice, recipients mice received at 1 and 7 days post-BMT donor DC and antifungal resistance in vivo, and that the priming capacity is strictly dependent on mode of DC delivery in vivo.

![Image](http://www.jimmunol.org/)

**FIGURE 4.** DC express fungal Ags upon exposure to *Candida albicans* yeasts or yeast RNA. Splenic DC from CD2F1 mice were pulsed with viable yeasts or hyphae (at DC:fungi ratio of 1:1 or 1:10, respectively) or 25 μg total RNA plus 250 μl DOTAP for 2 h at 37°C. Pulsing with viable fungi was stopped with amphotericin B. Cells were assessed 22 h later for fungal Ag expression by immunofluorescence analysis with the IgM B6.1 mAb (thick line) or the polyclonal antiserum from vaccinated mice (thin line) and subsequent reaction with the FITC-conjugated goat anti-mouse IgA plus IgG plus IgM (H + L) (Euroclone). Before all labeling experiments, FcR blocking was performed by incubating cells with 5% normal mouse serum. Unrelated isotype-matched mAbs were used as a control (filled histogram). Analysis was performed on a FACSScan.

![Image](http://www.jimmunol.org/)

**FIGURE 5.** Adoptively transferred DC pulsed with *Candida* yeasts or transfected with yeast but not hyphal RNA induce resistance to *Candida albicans* infection. SP-DC or BM-DC were isolated from CD2F1 mice and pulsed as in Fig. 1. A total of 5 × 10^6 DC were administered to recipient CD2F1 mice, either s.c. (□) or 14 and 7 (■) days before infection, or i.v. (●) 14 days before i.v. challenge with 5 × 10^6 *C. albicans* virulent cells. Seven days after infection, quantification of fungal growth was done in the kidneys and expressed as CFU. Mice not receiving DC; checked box, mice receiving DC transfected with hyphal RNA; *, p < 0.05, mice receiving DC vs mice not receiving DC.
received RNA-transfected DC as compared with mice not receiving DC and were similar to those of donor-vaccinated mice. Interestingly, CD4+ T cells also stained positive for the CD44 and CD25 activation surface markers (Fig. 8).

On assaying the pattern of cytokine production, we found that the frequency of IFN-γ-producing cells was increased in both freshly isolated T splenocytes and purified CD4+ T cells from mice receiving either yeast-pulsed or RNA-transfected DC as compared with recipients not receiving DC and was similar to that of donor-vaccinated mice (Fig. 9a). Upon Ag stimulation in vitro, splenocytes from DC-infused mice produced levels of IL-4 that were lower than those of mice not receiving DC and were similar to those of vaccinated donor mice (Fig. 9b). Interestingly, however, levels of IL-2 were also lower and those of IL-10 higher in DC-infused mice, as compared with not only untreated recipients but also donor-vaccinated mice. Therefore, the activation of CD4+ lymphocytes producing IFN-γ occurs together with IL-10 but not IL-2 production in hematopoietic BM recipients upon adoptive transfer of Candida-pulsed DC. Experiments in which RNA-transfected DC were infused into recipient mice who had received T-depleted BM cells from IFN-γ−/− mice demonstrated that the efficacy of RNA-transfected DC was mediated by the occurrence of donor IFN-γ-producing cells. Fig. 10 shows that the fungal growth in these mice was comparable to that observed in mice not receiving DC and significantly higher than that of mice transplanted with T-depleted BM cells from IFN-γ-sufficient mice. As a result, similar to what we observed in mice not receiving DC, all the mice succumbed to the infection as opposed to the long-term survival, with no signs of diseases, of mice who had received T-depleted BM cells from IFN-γ−/− mice.

**Discussion**

The present study shows that, consistent with previous results obtained with myeloid DC pulsed with Candida yeasts (11), DC transfected with yeast RNA are fully competent to induce Th1 priming upon adoptive transfer in vivo. Induction of antitumor immunity with tumor RNA-transfected DC represents a recently described approach for DC-based vaccines (25). In this study we show that this approach could be used to induce antimicrobial immunity as well.

It has been shown that DC transfected with MUC1 RNA expressed MUC1 tumor Ag along with costimulatory and adhesion molecules (24), a finding suggesting that newly synthesized molecules are potential targets for CTL recognition of RNA-transfected cells (24). We found in this study that DC from both BM and spleens underwent functional maturation in vitro upon pulsing with either live yeasts or yeast RNA, but not with hyphal RNA, as revealed by the up-regulated expression of MHC class II Ags and costimulatory molecules. No variation in CD8α expression was observed upon pulsing with either type of stimulus (data not shown), a finding suggesting that, in our system, myeloid DC do not convert into lymphoid DC upon activation, as suggested (40). Moreover, similar to yeast-pulsed DC, DC transfected with yeast but not hyphal RNA produced IL-12 p70 along with IL-6. As the ability of yeast-pulsed DC to induce Th1 priming in vivo was lost in conditions of IL-12 deficiency (11), these data suggest that DC transfected with yeast and not hyphal RNA would be fully competent to induce Th1 activation.

For this to occur, a prerequisite would be the expression of fungal Ags capable of inducing protective immunity on the surface of DC upon transfection with yeast RNA. This appeared to be the case as DC transfected with yeast but not hyphal RNA positively reacted with polyclonal Abs specifically reacting with Candida cells (36) and, partially, with mAb binding to a specific component of the acid-labile portion of the phosphomannan complex, which is known to be endowed with the ability to induce protective immunity in experimental candidiasis (39).
Pathways and mechanisms of fungal RNA and Ags processing by DC are largely unknown at the moment and remain to be explored.

Upon adoptive transfer in vivo, yeast RNA-transfected DC were capable of inducing the activation of *Candida*-specific Th1 cells, as revealed by the predominant production of IFN-γ over that of IL-4.
IL-4/IL-10. Moreover, the IL-12/β2R, a marker of CD4+ Th1 cells in candidiasis (5), was expressed on CD4+ T lymphocytes (data not shown). Associated with the occurrence of Candida-specific Th1 reactivity was the induction of a state of antifungal resistance, as revealed by the significant decrease of fungal growth in the target organs, such as the kidneys and the brain (data not shown).

Previous work in vivo in mice and in humans has suggested that, in addition to the maturation state (41), the mode of delivery of DC is important in eliciting T cell responses (42–45). DC given i.v. localized in the reticuloendothelial system and lung, while those given s.c. preferentially home to the T cell areas of the draining lymph nodes (42). In particular, induction of IFN-γ production, but not IL-4, was seen only with DC injected intradermally or intralymphatically but not i.v. (43). We found a similar result, as Th1 cell activation was observed after s.c. but not i.v. administration of DC. Although multiple injections of DC could lead to clonal exhaustion of the responding T cell population (43), the optimal protection against C. albicans was observed after two s.c. injections of Candida-pulsed DC. Labeling studies will clarify whether DC loaded with Candida home to different organs upon administration by different routes and whether the extent to which the ability of DC to migrate to relevant lymphoid organs will regulate the type and magnitude of the anticanidal immune response.

An interesting observation of the present study was that the infusion of Candida-pulsed DC accelerated the recovery of functional antifungal Th1 responses in a murine model of T cell-depleted allogeneic BMT. Patients receiving T cell-depleted BMT are unable to develop Ag-specific T cell responses soon after transplant (46). It has been demonstrated that T cell depletion of allogeneic BMT is associated with a slow recovery of CD4+ and CD8+ T cells (46). However, functional recovery of the T cell system after T cell-depleted allogeneic BMT has been demonstrated (47, 48), and both donor and recipient DC may contribute to the reconstitution of the T cell repertoire in transplantation through distinct pathways of Ag presentation (49). Interestingly, it has been reported that T cell reactivity to Candida readily recovered after T cell-depleted allogeneic BMT, and the fact that recovery was related to the absence of antifungal prophylactic measures indicates that anticanidal reactivity of residual T cells could be maintained through continuous exposure to the fungus (50).

We have recently demonstrated that an unbalanced production of Th1 and Th2 cytokines was responsible for the susceptibility to candidiasis observed in our BMT model. However, readressing the balance between Th1 and Th2 subsets, as by treatment with Th2 cytokine antagonists, accelerated the recovery of Th1-mediated antifungal resistance (26). In this study we show that recovery of functional NK and T cells producing IFN-γ could also be accelerated by the infusion of Candida-pulsed DC. This, along with the decreased IL-4 production, translated in the occurrence of a state of antifungal resistance as revealed by the impaired fungal growth in different target organs, including those most frequently associated with fungal disease and pathology in BMT settings, such as the gut, liver, and lungs. Therefore, although conflicting data exist as to whether donor DC are “friends” or “foes” in transplantation (49), our findings suggest that DC may contribute to the educational program of T cells in BMT during reconstitution, in line with the recent observation of the immunization capacity of tumor-pulsed (51) or Aspergillus-pulsed (18) DC in BM-transplanted mice and humans.

The observation that the occurrence of a protective Th1 reactivity coexisted with the detection of significant levels of IL-10 is intriguing. It is known that high levels of IL-10 are associated with tolerance to HLA-mismatched BM stem cells (52) and IL-10 is required for the induction of regulatory T cells mediating tolerance to alloantigens in vivo (52–55). Different regulatory T cells have been reported and they are known to differentiate upon the encounter with immature DC (53, 55). However, fully competent DC were also found to be inducers of T cell anergy when presenting a self-epitope with altered peptide ligand features (56). Whether IL-10 produced in BM-transplanted mice infused with Candida-pulsed DC may serve to support the growth of regulatory T cells preventing donor Th1 alloreactivity remains a working hypothesis, although it may help to explain the long-term, disease-free survival of the mice. In terms of interference with the development and activity of antifungal Th1 reactivity, it is well established that the effect of IL-10 is dependent upon the dose of the cytokine. At high doses, IL-10 had a negative effect on Th1-mediated antifungal resistance (57), but at low doses IL-10 was required for optimal induction and maintenance of anticanidal Th1 immune response (58).

A recent study showed that yeast cells could act as an efficient vaccine and elicited protective cell-mediated immunity by virtue of their ability to induce DC maturation, IL-12 production, and efficient priming for MHC class I- or II-restricted Ag-specific T cell responses (59). We show in this work that not only the yeasts but yeast RNA could be an efficient trigger of DC maturation, IL-12 production, and induction of protective Ag-specific responses to Candida. Interestingly, inactivated yeasts failed to induce DC maturation in vitro, and DC pulsed with inactivated yeasts failed to promote Th1 immunity upon adoptive transfer in

FIGURE 10. Fungal RNA-transfected DC fail to induce antifungal resistance in mice receiving allogeneic T-depleted BM cells from IFN-γ−/−mice. Lethally irradiated C3H/HeJ mice, transplanted with ≥ 2 × 10⁶ T cell-depleted BM cells from wild-type (■) or IFN-γ−/− (□) mice, received 5 × 10⁸ RNA-transfected DC 1 and 7 days post BMT. A week after the last DC administration, mice were infected with 10⁸ C. albicans live vaccine strain and reinfected with virulent Candida cells 14 days later. □. BM-transplanted mice not receiving DC. At 3 days after reinfection, the fungal growth in different organs was evaluated and expressed as CFU per organ. *, p < 0.05, mice receiving DC vs mice not receiving DC.
vivo (data not shown). Therefore, these data may account for the long-standing observation of the inability of inactivated Candida to induce memory antifungal protective immune responses.

It has been shown that human DC phagocytose and process C. albicans (60, 61) as well as other fungi (18, 62), and ex vivo-generated DC pulsed with Aspergillus conidia restored antifungal immunity in BM-transplanted patients (18). This work suggests that DC could act as effective vaccines against fungal infections and that RNA-transfected DC could be of vaccinating potential in conditions that negate the use of attenuated microorganisms, such as immunosuppression, or in the case of poor availability of protective Ags.

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2912 DC VACCINATION IN MURINE CANDIDIASIS

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