Processing of Exogenous Antigens for Presentation by Class I MHC Molecules Involves Post-Golgi Peptide Exchange Influenced by Peptide-MHC Complex Stability and Acidic pH

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Vacuolar alternate class I MHC (MHC-I) pathway involves processing of protein Ags expressed within the cell (endogenous Ags) for presentation of peptides derived from endogenous Ags by MHC-I molecules. However, it is well established that endogenous Ags can also be processed for presentation by MHC-I molecules via alternate MHC-I Ag-processing mechanisms (1–9). Experimental evidence supports several different alternate MHC-I processing mechanisms.

Cytosolic alternate MHC-I processing mechanisms involve the egress of exogenous Ags from vacuolar (endocytic or phagocytic) compartments to enter the cytosol (10–13). These Ags merge with conventional MHC-I Ag processing mechanisms. Their processing is dependent on proteasomes for proteolytic production of antigenic peptides and the TAP for transport of these peptides into the endoplasmic reticulum (ER). In this pathway, peptides bind to newly synthesized MHC-I molecules in the ER, and the resulting peptide-MHC-I complexes are transported from the ER through the Golgi complex and eventually to the cell surface. This pathway is blocked by deficits in TAP or treatment of cells with proteasome inhibitors or brefeldin A (an inhibitor of anterograde ER-Golgi transport) (11).

Vacuolar alternate MHC-I processing mechanisms involve the processing of exogenous Ag in vacuolar compartments without penetration of Ag into the cytosol. Vacuolar processing produces antigenic peptides that bind to MHC-I molecules that have already exited from the Golgi complex (i.e., MHC-I molecules in post-Golgi compartments). Hence, this pathway is relatively resistant to treatment with proteasome inhibitors or brefeldin A. However, the supply of peptide-receptive MHC-I molecules in post-Golgi compartments can be depleted by long incubations with brefeldin A or deficits in TAP function. In some experimental systems, vacuolar alternate MHC-I Ag processing is indirectly dependent on TAP and is partially inhibited by deficits in TAP, even though TAP does not transport the peptides derived from exogenous Ags in these systems (5, 14, 15). In other systems, vacuolar alternate MHC-I Ag processing is completely TAP independent (16–20). The site where peptides bind to MHC-I molecules during vacuolar alternate MHC-I Ag processing is still unclear, and binding may take place within intracellular processing compartments that contain MHC-I molecules (e.g., phagosomes) (21) or on the cell surface following recycling and regurgitation of peptides (22).

Early reports suggested that TAP-deficient cells express “empty” MHC-I molecules under certain conditions (e.g., at 26°C) (23). However, recent evidence indicates that such MHC-I molecules do contain peptides, but these peptide-MHC-I complexes are less stable than most complexes expressed on TAP-replete cells at 37°C (24). Decreased ER peptide supply in TAP-deficient cells may limit availability of high affinity peptides. Peptide binding may be required for MHC-I molecules to exit from the ER (25–29). Limited peptide supply in TAP-deficient cells may result in exit of...
peptide-MHC-I complexes that have relatively low affinity peptide, high dissociation rates, and decreased stability at 37°C (24). TAP-deficient cells have decreased levels of both total (30) and peptide-receptive (5, 14, 31) MHC-I on the cell surface (and possibly in post-Golgi vacuolar compartments that communicate with the cell surface), limiting the availability of peptide-receptive MHC-I molecules available for vacuolar alternate MHC-I Ag processing.

In this study, we analyze the source and nature of post-Golgi peptide-receptive MHC-I molecules that are used in alternate MHC-I processing. Because TAP contributes indirectly to vacuolar alternate MHC-I Ag processing without transporting peptide derived from exogenous Ag (5, 14), TAP-dependent loading of MHC-I with endogenous peptides may enhance exit of MHC-I molecules from the ER and their availability for subsequent binding and presentation of peptides derived from exogenous Ags. This model suggests that vacuolar alternate MHC-I Ag processing involves dissociation of one peptide and binding of peptide derived from exogenous Ag (i.e., peptide dissociation/exchange). Our experiments address the role of prior peptide occupancy in controlling the level of peptide-receptive MHC-I molecules that contribute to alternate MHC-I Ag processing. Macrophages from TAP1−/− mice provide a good system to address this question. Macrophages mediate alternate MHC-I processing of particulate Ag (2). TAP-deficient cells express MHC-I molecules that are not optimally loaded and can bind exogenous peptide (30) to generate a large cohort of homogenous peptide-MHC-I complexes. Such cohorts can be assessed as potential precursors for peptide-receptive MHC-I molecules that function in processing and presentation of exogenous Ag. Overnight incubation of TAP1−/− macrophages with a first (“stabilizing”) peptide stabilized Kb molecules and enhanced subsequent binding and presentation of a second (“readout”) peptide by Kb molecules. This enhancement was influenced by the stability of stabilizing peptide-Kb complexes and was increased by exposure of Kb molecules to acidic pH (similar to that of intracellular vacuolar compartments). These observations suggest that peptide exchange contributes to alternate MHC-I Ag processing and is influenced by the stability of peptide-MHC-I complexes and the pH of the processing environment.

Materials and Methods

Animals and cells

TAP1−/− mice (32) were backcrossed for six generations with C57BL/6 mice and were generously provided by Dr. L. Van Kaer (Vanderbilt University, Nashville, TN). TAP1−/− and C57BL/6 mice (The Jackson Laboratory, Bar Harbor, ME) were bred and housed under specific pathogen-free conditions at Case Western Reserve University (Cleveland, OH). Activated peritoneal macrophages were harvested 4 days after i.p. inoculation with Con A (100 μg/mouse). The T hybridoma CD8OVA1.3 (2) was used to detect SIINFEKL-Kb complexes. The T hybridoma SV7B1.6 was used to detect FAPGNYPAL-Kb complexes. To obtain this hybridoma, C57BL/6 mice were infected intranasally with 103 PFU Sendai virus and sacrificed 19 days after infection. Spleen cells were stimulated in vitro with 1–5 nM Sendai virus nucleoprotein (324–332) (FAPGNYPAL) for 6 days, mixed 1:4 with BW5147 TCRαβ CD8+ cells (33), and fused with polyethylene glycol. Fusion products were screened and characterized, and SVB1B6 was selected for these studies. RMA-S cells are TAP-deficient (TAP2-mutant) Rauscher virus-transformed murine T cell lymphoma cells (H-2Kd) (30). Cells were cultured in standard medium consisting of DMEM (Life Technologies, Grand Island, NY) supplemented with 10% heat-inactivated FCS (HyClone Laboratories, Logan, UT), 5 × 10−5 M 2-ME, 1 mM sodium pyruvate, HEPES buffer, and penicillin-streptomycin (Life Technologies). Incubations were conducted in a 5% CO2 atmosphere at 37°C or 26°C.

Ag preparations

Peptides included SIINFEKL (OVA257−264), KVVRFDKL (OVA35−42), FAPGNYPAL (Sendai virus nucleoprotein 324–332), RGYVYQGL (ve-
derive from peptide-occupied MHC-I molecules via peptide dissociation/exchange as opposed to empty MHC-I molecules that have never bound peptide. Initial loading of MHC-I molecules may occur with an endogenous peptide in the ER, allowing export to post-Golgi compartments. Low affinity peptide-MHC-I complexes may subsequently participate in peptide dissociation/exchange, allowing MHC-I molecules to bind exogenous peptide or peptide derived from exogenous Ag by alternate MHC-I Ag processing.

In testing the role of peptide dissociation/exchange, it is difficult to manipulate the initial loading of MHC-I molecules with endogenous peptides in the ER. We overcame this difficulty by manipulating the loading of MHC-I molecules after their exit from the ER and studying their subsequent ability to bind and present new antigenic peptide. A large cohort of post-Golgi peptide-MHC-I complexes was generated with an exogenous (stabilizing) peptide to determine how these complexes affected or contributed to subsequent binding of a second exogenous (readout) peptide or binding of a peptide derived from alternate MHC-I processing of particulate Ag. TAP1\(^{-/-}\) cells were used to achieve this goal, because many of their MHC-I molecules are poorly loaded (probably containing low affinity peptides). Thus, exogenous peptide can load a greater proportion of MHC-I molecules and have greater impact on their cell surface expression and potential contribution to peptide dissociation/exchange. Brefeldin A was added before addition of readout peptide to focus the experiment on the defined cohort of stabilizing peptide-MHC-I complexes in post-Golgi compartments and to exclude MHC-I molecules synthesized after the removal of stabilizing peptide. Thus, incubation of TAP1\(^{-/-}\) macrophages with or without exogenous stabilizing peptide determined the presence or absence of a large cohort of post-Golgi peptide-MHC-I complexes that were potentially available for peptide dissociation/exchange.

These experiments assessed the effects of four different stabilizing peptides that bind K\(^b\) (SIINFEKL, KVVRFDKL, FAPGNYPAL, or RGYVYQGL) on the binding and presentation of K\(^b\)-restricted readout peptides (SIINFEKL or FAPGNYPAL) to T hybridoma cells. Relative to incubation in standard medium with no stabilizing peptide, overnight incubation of TAP1\(^{-/-}\) macrophages with RGYVYQGL or KVVRFDKL enhanced subsequent binding and presentation of FAPGNYPAL to SVB1.B6 T hybridoma cells (Fig. 1). As shown in Fig. 1B, the response to readout peptide was not altered by prior incubation with a peptide that does not bind to K\(^b\) (HIV reverse transcriptase 476–484 peptide, ILKEPVHG\(^b\), which binds HLA-A2). Addition of stabilizing peptide to wild-type C57BL/6 macrophages had little effect on production of readout peptide-K\(^b\) complexes (data not shown and below), presumably because wild-type cells have a sufficient supply of endogenous stabilizing peptides. These results suggest that RGYVYQGL and KVVRFDKL bound and stabilized K\(^b\) molecules, increasing expression of peptide-K\(^b\) complexes that subsequently provided peptide-receptive K\(^b\) molecules to bind and present the FAPGNYPAL readout peptide.

Enhancement of peptide-receptive MHC-I molecules by stabilizing peptide is influenced by the stability of stabilizing peptide-MHC-I complexes

In contrast to the results seen with RGYVYQGL or KVVRFDKL as stabilizing peptide, overnight incubation with SIINFEKL decreased subsequent binding and presentation of FAPGNYPAL (Fig. 1). We proposed that the stability of SIINFEKL-K\(^b\) complexes might limit their dissociation and production of peptide-receptive K\(^b\) molecules. Other studies have established the high affinity of SIINFEKL for K\(^b\) (\(K_d = 1.6–6.7 \times 10^{-9}\) M) (39–41) and high stability of SIINFEKL-K\(^b\) complexes (39, 42). In addition, we studied the ability of the peptides used in this study to increase expression of thermostable K\(^b\) molecules on TAP-deficient cells. This assay has been used by other groups as an indirect indication of relative peptide affinity and/or stability of peptide-MHC-I complexes, although this method does not allow calculation of true affinity (41, 43, 44). After overnight incubation with peptide at 26°C and incubation at 37°C for 2–4 h to down-regulate unstable MHC-I molecules, flow cytometry showed that SIINFEKL, KVVRFDKL, FAPGNYPAL, and RGYVYQGL all increased cell surface expression of K\(^b\) on RMA-S cells, confirming that these peptides bound and stabilized K\(^b\) (Fig. 2A). When DGSTDYGILQINSR (which does not bind to K\(^b\)) was used as the stabilizing peptide, enhancement of K\(^b\) expression was not seen (data not shown). SIINFEKL enhanced K\(^b\) expression substantially more than the other peptides, which appeared relatively similar in their ability to up-regulate K\(^b\). These data indicate that SIINFEKL formed a greater number of stable peptide-K\(^b\) complexes than the other peptides under these conditions.

To directly assess the stability of peptide-K\(^b\) complexes and to determine the half-life of different peptide-K\(^b\) complexes, we studied the decay of specific peptide-induced enhancement of K\(^b\) expression on RMA-S cells or TAP1\(^{-/-}\) macrophages. Cells were incubated overnight with peptide to stabilize K\(^b\) expression, washed, subjected to a chase incubation of varying length at 37°C to allow decay of specific peptide-K\(^b\) complexes, and stained for determination of K\(^b\) expression by flow cytometry. Under these conditions, the majority of K\(^b\) molecules were loaded with the specific peptide (see medium controls in Figs. 2, A and C) so that the half-life of K\(^b\) expression approximated the half-life of the specific peptide-MHC-I complex. On RMA-S cells, \(t_{1/2}\) was determined to be 8.9 h for SIINFEKL-K\(^b\) complexes, 3.4 h for FAPGNYPAL-K\(^b\) complexes, 3.1 h for RGYVYQGL-K\(^b\) complexes, and 1.6 h for KVVRFDKL-K\(^b\) complexes (Fig. 2B). Similar results were obtained with TAP1\(^{-/-}\) macrophages, with a \(t_{1/2}\) of 7.2 h for SIINFEKL-K\(^b\) complexes, 3.7 h for FAPGNYPAL-K\(^b\) complexes, and 1.6 h for RGYVYQGL-K\(^b\) complexes.
complexe:s, 2.8 h for RGYVYQGL-Kb complexes, and 1.6 h for KVVRFKDL-Kb complexes (Fig. 2D). These figures are similar to those produced by other studies, e.g., that of Chen et al. (39), which showed a t1/2 of 8.3 h for SIINFEKL-Kb complexes and 2.7 h for KVVRFKDL-Kb complexes. Thus, SIINFEKL-Kb complexes are more stable than the other peptide-Kb complexes examined here. The high stability of SIINFEKL-Kb complexes may limit their ability to serve as precursors for peptide-receptive MHC-I molecules that are used in alternate MHC-I Ag processing or for binding and presentation of exogenous peptide.

**Stabilizing peptide enhances subsequent production of SIINFEKL-Kb complexes by alternate MHC-I processing of HB101.Crl-OVA to a greater degree than production of these complexes from soluble SIINFEKL.**

As observed in previous studies (2, 5, 14, 22), macrophages processed HB101.Crl-OVA (E. coli expressing an OVA fusion protein) to produce SIINFEKL-Kb complexes that were detected by CD80VA1.3 T hybridoma cells (Fig. 3). Incubation of macrophages with HB101.Crl-hen egg lysozyme, which expresses a different antigenic fusion protein, resulted in no detectable response by CD80VA1.3 T hybridoma cells (data not shown), confirming the antigenic specificity of this system.

Incubation of macrophages with stabilizing peptide (FAPGNYPAL, KVVRFKDL, or RGYVYQGL) produced substantial enhancement of subsequent alternate MHC-I processing of HB101.Crl-OVA to produce SIINFEKL-Kb complexes (Fig. 3B). This implies that stabilization of MHC-I molecules as peptide-MHC-I complexes provides precursors to the peptide-receptive MHC-I molecules that are used in alternate MHC-I Ag processing. In addition, enhancement of SIINFEKL-Kb complexes was more pronounced with addition of exogenous SIINFEKL (Fig. 3A), which can bind to cell surface Kb molecules (in some experiments the enhancement of exogenous SIINFEKL presentation was even less than in Fig. 3A). This suggests that the intracellular processing environment promotes peptide dissociation/exchange to a greater degree than the environment at the cell surface.

**Effects of stabilizing peptide on subsequent binding of readout peptide as quantified by flow cytometry detection of SIINFEKL-Kb complexes.**

Experiments using a T cell assay supported the hypothesis that stabilizing peptides generate peptide-MHC-I complexes that can subsequently dissociate or undergo peptide exchange to bind an additional readout peptide. A T cell assay is advantageous for this type of experiment because it allows detection of physiological levels of peptide-MHC-I complexes, as can be generated by alternate MHC-I Ag processing. However, T cell assays do not provide quantitative assessment of peptide-MHC complexes and are subject to some potential artifacts. The mAb 25-D1.16 is specific for complexes, 2.8 h for RGYVYQGL-Kb complexes, and 1.6 h for KVVRFKDL-Kb complexes (Fig. 2D). These figures are similar to those produced by other studies, e.g., that of Chen et al. (39), which showed a t1/2 of 8.3 h for SIINFEKL-Kb complexes and 2.7 h for KVVRFKDL-Kb complexes. Thus, SIINFEKL-Kb complexes are more stable than the other peptide-Kb complexes examined here. The high stability of SIINFEKL-Kb complexes may limit their ability to serve as precursors for peptide-receptive MHC-I molecules that are used in alternate MHC-I Ag processing or for binding and presentation of exogenous peptide.

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SIINFEKL-Kb complexes (38) and can be used to directly quantify these complexes by flow cytometry, although the lower sensitivity of this assay makes it useful only for measuring the production of complexes with relatively high concentrations of exogenous peptide (the physiological level of SIINFEKL-Kb complexes generated by alternate MHC-I Ag processing, e.g., of HB101.Crl-OVA, is not detectable by flow cytometry using this Ab).

Fig. 4 shows 25-D1.16 staining of cell surface SIINFEKL-Kb complexes on TAP1−/− macrophages that were first incubated with or without a stabilizing peptide (FAPGNYPAL, KVVRFDKL, or RGYYVQGL) and then incubated with or without SIINFEKL readout peptide. In each case, stabilizing peptide increased the production of SIINFEKL-Kb complexes. SIINFEKL-specific mean fluorescence value (MFV) was defined as the MFV with SIINFEKL minus the MFV without SIINFEKL. Incubation with stabilizing peptide increased SIINFEKL-specific MFV by 3.5- to 4.5-fold (see Fig. 4 legend). Staining of macrophages with 25-D1.16 was increased after incubation with KVVRFDKL even without addition of SIINFEKL. This indicates that 25-D1.16 also recognizes KVVRFDKL-Kb complexes, probably due to the similar carboxyl-terminal sequences of KVVRFDKL and SIINFEKL (FDKL vs FEKL). Even against the background with KVVRFDKL, addition of SIINFEKL produced an increase in 25-D1.16 staining consistent with a level of SIINFEKL-Kb complexes similar to that achieved after incubation with other stabilizing peptides. In conclusion, flow cytometry quantitatively demonstrated that prior incubation with stabilizing peptide caused substantial increases in the generation of SIINFEKL-Kb complexes.

Acidic pH increases subsequent binding and presentation of readout peptide

Stabilizing peptide caused a greater increase in presentation of SIINFEKL derived from alternate MHC-I processing of HB101.Crl-OVA than presentation of exogenous SIINFEKL peptide (Fig. 3). This suggested the hypothesis that peptide dissociation/exchange is promoted more by the acidic environment of phagosomes and phagolysosomes than the neutral pH at the cell surface. To test the role of pH in controlling peptide dissociation/exchange, freshly isolated C57BL/6 macrophages were exposed to buffers of varying pH for 20 min, neutralized, incubated with SIINFEKL for 2 h in the presence of brefeldin A, and then fixed. The resulting level of SIINFEKL-Kb complexes was assessed using CD80VA1.3 T hybridoma cells. Exposure of macrophages to acidic conditions (pH 5.0–6.5, similar to the pH of endocytic or phagocytic compartments) substantially increased subsequent binding and presentation of SIINFEKL peptide (Fig. 5). Macrophages exposed to pH 4.5 exhibited markedly decreased subsequent binding and presentation of SIINFEKL, presumably due to cell injury at this very acidic pH. Because C57BL/6 macrophages should be replete with diverse endogenous peptide-Kb complexes, these data suggest that acidic pH enhances dissociation of previously associated peptides (endogenous stabilizing peptides), thereby increasing the number of peptide-receptive MHC-I molecules.

To further test the roles of both acidic pH and stabilizing peptide on availability of peptide-receptive MHC-I molecules, C57BL/6 and TAP1−/− macrophages were incubated overnight at 26°C with FAPGNYPAL stabilizing peptide, subsequently exposed to acidic pH, and then incubated with SIINFEKL readout peptide. SIINFEKL-Kb complexes were detected with CD80VA1.3 T hybridoma cells. With C57BL/6 macrophages, addition of FAPGNYPAL stabilizing peptide had little effect on production of SIINFEKL-Kb complexes, presumably because these TAP-replete cells have a sufficient supply of endogenous stabilizing peptides. Incubation of C57BL/6 macrophages at acidic pH markedly enhanced subsequent production of SIINFEKL-Kb complexes (Fig. 6A, consistent with Fig. 5), suggesting that dissociation of endogenous stabilizing peptides was enhanced at acidic pH. With TAP1−/− macrophages, both FAPGNYPAL stabilizing peptide and incubation at acidic pH promoted formation of SIINFEKL-Kb complexes, and a combination of FAPGNYPAL stabilizing peptide and incubation at acidic pH provided the highest presentation of SIINFEKL-Kb complexes.

FIGURE 4. Effects of stabilizing peptide on subsequent binding of readout peptide as quantified by flow cytometry detection of SIINFEKL-Kb complexes. TAP1−/− macrophages were treated as described in Fig. 3 (with stabilizing peptides at 50 μM) except that readout peptide SIINFEKL was present at 5 μM for 2 h, and cells were subsequently stained for flow cytometry with 25-D1.16 anti-SIINFEKL-Kb Ab. Results are expressed in the graph as MFV of events gated to include intact cells. SIINFEKL-specific MFV was defined as (MFV with SIINFEKL) − (MFV without SIINFEKL). SIINFEKL-specific MFV was 53 after incubation without stabilizing peptide, 183 after incubation with FAPGNYPAL, 238 after incubation with KVVRFDKL, and 194 after incubation with RGYYVQGL.

FIGURE 5. Incubation of C57BL/6 macrophages at acidic pH increases subsequent binding and presentation of SIINFEKL. C57BL/6 macrophages were incubated in citrate-buffered saline at varying pH for 20 min, washed, incubated with SIINFEKL for 2 h in the presence of brefeldin A (to exclude nascent Kb molecules that were not exposed to the acidic buffer), fixed, and incubated with CD80VA1.3 T hybridoma cells as in Fig. 3. Results are expressed as means of quadruplicate wells ± SD.
To assess the relationship between the stability of stabilizing peptide-MHC-I complexes and the role of pH in promoting peptide dissociation/exchange, a similar experiment was performed with a switch in the stabilizing and readout peptides. TAP1−/− macrophages were incubated overnight at 26°C with or without 5 μM FAPGNYPAL stabilizing peptide and then processed as in Fig. 5. Results are expressed as means of quadruplicate wells ± SD.

FIGURE 6. Stabilizing peptide and exposure to acidic pH can increase subsequent binding and presentation of readout peptide. C57BL/6 or TAP1−/− macrophages were incubated overnight at 26°C with or without 5 μM FAPGNYPAL stabilizing peptide and then processed as in Fig. 5. Results are expressed as means of quadruplicate wells ± SD.

FIGURE 7. Production of FAPGNYPAL-Kb complexes is increased by prior exposure of macrophages to acidic pH but decreased by prior exposure to SIINFEKL stabilizing peptide. Macrophages were processed as described in Fig. 6 except the stabilizing peptide was SIINFEKL (5 μM), the readout peptide was FAPGNYPAL and Kb-FAPGNYPAL complexes were detected with SVB1.B6 T hybridoma cells. Results are expressed as means of quadruplicate wells ± SD. Where error bars are not visible, they are smaller than the symbol width.

Discussion

The results of this study establish that peptide dissociation/exchange contributes significantly to vacuolar alternate MHC-I Ag processing. Some previous models do not include a role for MHC-I peptide dissociation/exchange and indicate that cell surface or post-Golgi peptide-receptive MHC-I molecules are empty, having escaped from the ER without binding a peptide. These models predict that incubation of cells with stabilizing peptide should reduce both the number of empty peptide-receptive MHC-I molecules and subsequent binding of readout peptide by MHC-I. In contrast, our results demonstrate that binding of stabilizing peptides can substantially enhance subsequent binding and presentation of exogenous readout peptide or peptide derived from exogenous Ag via alternate MHC-I Ag processing, implying a process of peptide dissociation/exchange. We propose that the role of stabilizing peptide is to bind MHC-I molecules in a reversible manner (depending on peptide affinity), increase the egress of MHC-I molecules from the ER (in the case of endogenous stabilizing peptides), increase the half-life of MHC-I molecules in post-Golgi compartments, and thereby increase the supply of precursors to peptide-receptive MHC-I molecules. Dissociation of stabilizing peptide-MHC-I complexes may then allow the formation of readout peptide-MHC-I complexes.

Two independent assay systems were used to establish this point. A T hybridoma assay was used to detect presentation of the SIINFEKL and FAPGNYPAL readout peptides. The advantages of this assay include the availability of T hybridomas with different antigenic specificities and its sensitivity to physiological levels of peptide-MHC-I complexes that are generated by alternate MHC-I Ag processing. Disadvantages of the T cell assay include the difficulty of directly quantifying peptide-MHC-I complexes and the possibility that variations in the numerous aspects of T cell activation may complicate the experiment. To complement the T hybridoma assay, SIINFEKL-Kb complexes were also detected by flow cytometry and staining with the 25-D1.16 mAb (38). The advantages of this approach include a more direct and quantitative detection of peptide-MHC-I complexes, but its disadvantages include availability of only a single Ab specificity for this study, a much lower sensitivity relative to T hybridoma assays, and a consequent inability of flow cytometry to detect the levels of peptide-MHC-I complexes generated during alternate MHC-I Ag processing (14). Thus, data presented here for experiments involving processing of exogenous Ag (e.g., HB101.Crl-OVA) exclusively reflect T hybridoma assays, whereas both T hybridoma and flow cytometry assays were used in experiments involving the presentation of exogenous peptide. Protocols for these two types of experiments provided very different concentrations of exogenous...
peptides available to bind K^b molecules, but the results of both supported identical conclusions, indicating that these observations may be applied over a wide range of peptide concentration. Thus, two independent approaches both supported the conclusion that binding of stabilizing peptide could increase subsequent binding and presentation of a different readout peptide.

Our model is consistent with studies that indicate a requirement for peptide loading to promote efficient egress of MHC-I molecules from the ER and other studies that demonstrate the existence of MHC-I peptide exchange in post-Golgi compartments (including the cell surface). MHC-I molecules are retained in the ER by a calnexin- and tapasin-dependent mechanism until they bind a peptide (25–29). Even the thermostable MHC-I molecules expressed on the surface of TAP-deficient cells (previously considered empty) are actually loaded with peptides, most of which are presumably of relatively low affinity (24). Following exit from the ER, some peptide-MHC-I complexes may dissociate to participate in peptide exchange or exist as short-lived empty MHC-I molecules (MHC-I H chain plus β2-microglobulin or free MHC-I H chains). Ojcius et al. reported that the dissociation of peptide-MHC-I complexes produced peptide-receptive MHC-I molecules that bound exogenous peptide (45, 46). Several studies from Hansen and colleagues have indicated that L^d can undergo extensive peptide exchange (42, 47–49). Empty MHC-I molecules (50) or free MHC-I H chains (51–55) have been detected on the cell surface, where they may arise by peptide dissociation, and some studies have demonstrated that these molecules reassemble with peptide and β2-microglobulin to form peptide-MHC-I complexes. Thus, several populations of potentially peptide-receptive MHC-I molecules may arise via peptide dissociation/exchange.

Our data suggest an interpretation different from that proposed by Schirmbeck and Reimann based on studies of L^d function in vacuolar alternate MHC-I processing of hepatitis B surface Ag particles (56). These authors present evidence that cell surface-derived empty L^d molecules are involved in alternate MHC-I Ag processing. The detection of empty L^d molecules is based on their binding by mAb 64-3-7, as established by Hansen and colleagues (42, 47–49). L^d molecules that bind 64-3-7 do not bind the 30-5-7 mAb that recognizes peptide-associated L^d. However, it is difficult to completely exclude the possibility that L^d molecules recognized by 64-3-7 are actually associated with some peptides (perhaps of relatively low affinity) that could play a role in their stability. In contrast, Myers et al. used immunoprecipitation to demonstrate that 30-5-7-negative L^d molecules are not associated with β2-microglobulin (57). One explanation for the differences in results between our studies and those of Schirmbeck and Reimann lies in the specific MHC-I molecules involved (K^b vs L^d). Different MHC-I molecules may have differing stabilities in the absence of peptide. Thus, some MHC-I molecules may require stabilizing peptides in the fashion shown for K^b in our studies, whereas others may achieve some stability in the absence of peptide.

In our studies, exogenous stabilizing peptides enhanced the expression of peptide-receptive MHC-I molecules by TAP-deficient cells. Previous studies have shown that TAP-deficient cells have a paucity of peptide-receptive MHC-I molecules (5, 31), presumably due to a decreased supply of endogenous stabilizing peptides. In the present studies, TAP^−/− macrophages provided low binding of exogenous peptide by MHC-I without prior addition of stabilizing peptide. In contrast, TAP-replete C57BL/6 macrophages constitutively expressed higher levels of peptide-receptive MHC-I molecules, presumably due to a supply of endogenous stabilizing peptides, and the addition of exogenous stabilizing peptide did not enhance the subsequent binding and presentation of a readout peptide by C57BL/6 macrophages.

The deficit of endogenous stabilizing peptide and consequently decreased levels of peptide-receptive MHC-I molecules explains the decreased efficiency of vacuolar alternate MHC-I Ag processing in TAP^−/− macrophages. The decrease in alternate MHC-I Ag processing associated with TAP deficit is reversed by conditions that stabilize poorly loaded MHC-I molecules, such as incubation at 26°C (5, 14) with β2-microglobulin (5) or with stabilizing peptide (as shown in this study). Addition of exogenous peptide or β2-microglobulin promotes stability of MHC-I at the cell surface, but these agents or MHC-I molecules from the cell surface can also be internalized into post-Golgi vacuolar compartments to participate in alternate MHC-I Ag processing. We propose that the role of TAP in vacuolar alternate MHC-I Ag processing is not to transport readout peptides processed from exogenous Ag, but rather to deliver stabilizing peptides to MHC-I molecules in the ER, promoting the egress of effective precursors to peptide-receptive MHC-I molecules to post-Golgi compartments.

The mechanism of peptide dissociation/exchange remains uncertain. Stabilizing peptides may dissociate to generate truly empty MHC-I molecules that undergo separate interactions to bind readout peptides. Empty MHC-I molecules would be relatively unstable, but their continuous production from endogenous stabilizing peptide-MHC-I complexes could provide a continuous source of peptide-receptive MHC-I for alternate MHC-I Ag-processing mechanisms. Stabilizing peptides would increase the lifetime of precursors to empty MHC-I molecules, which might sequentially bind several low affinity stabilizing peptides before binding a high affinity peptide to create a stable peptide-MHC-I complex (a process akin to the “peptide editing” proposed for class II MHC (MHC-II) molecules). Alternatively, binding of readout peptide may be mechanistically linked to dissociation of a stabilizing peptide, allowing stabilizing peptide to occupy and stabilize MHC-I molecules up to the moment of readout peptide binding.

Peptide dissociation/exchange depends on the dissociation rate of stabilizing peptide-MHC-I complexes. Stabilizing peptides with very high affinity may create stable peptide-MHC-I complexes with such low dissociation rates that they do not contribute to peptide exchange. Increasing the loading of MHC-I with one such stabilizing peptide (SIINFEKL) decreased subsequent binding and presentation of readout peptide (Figs. 1 and 7). Other studies have also indicated that peptide exchange is influenced by the stability of preexisting peptide-MHC-I complexes (47). The exact half-life of endogenous stabilizing peptide-MHC-I complexes that normally serve as precursors to peptide-receptive MHC-I molecules is not clear. Vacular alternate MHC-I Ag processing is maintained in the presence of brefeldin A (2, 5, 22), implicating a post-Golgi pool of stabilizing peptide-MHC-I complexes that are precursors to peptide-receptive MHC-I molecules. However, long incubations with brefeldin A inhibit alternate MHC-I Ag processing (5), suggesting that the subset of stabilizing peptide-MHC-I complexes that contribute to peptide dissociation/exchange eventually decays with a half-life shorter than that of high stability peptide-MHC-I complexes.

The enhancement of readout peptide presentation by stabilizing peptide was greater for presentation of SIINFEKL derived from HB101.Cr1-OVA, which is processed in acidic phagolysosomal compartments, than for presentation of exogenous SIINFEKL, which can bind to K^b at the cell surface at neutral pH. This suggested the hypothesis that peptide exchange is promoted by the acidic pH. Brief exposure of C57BL/6 or TAP^−/− macrophages to acidic pH increased subsequent binding of readout peptide to surface MHC-I molecules, supporting this hypothesis. Other studies have also indicated that optimum peptide binding or exchange is obtained under acidic conditions (58–60). For example, Stryhn
et al. (58) suggested that acid-dependent dissociation of peptide-MHC-I complexes results in peptide exchange that may contribute to the presentation of exogenous Ags by MHC-I molecules, although their studies also indicated that the effect of pH and the effective range of pH varied for different peptide-MHC-I combinations and with the presence of exogenous β2-microglobulin (these factors may explain some variation in the pH optima observed in different studies). Overall, these observations suggest that phagolysosomes or other acidified intracellular vacuolar compartments are sites where peptides derived from exogenous Ag bind to MHC-I molecules during alternate MHC-I Ag processing.

There are interesting parallels between the vacuolar alternate MHC-I and conventional MHC-II Ag processing pathways. Both MHC-I and MHC-II molecules require chaperones for stabilization before peptide binding and interact with other proteins to promote proper peptide loading (e.g., tapasin and calnexin for MHC-I and invariant chain and HLA-DM for MHC-II). MHC-I and MHC-II molecules are unstable without bound peptide, and both are occupied by stabilizing peptides before binding of readout peptides (MHC-II uses invariant chain and invariant chain-derived peptides, whereas MHC-I uses stabilizing peptides as described in this study). Furthermore, two recent studies of MHC-II molecules (61, 62) illustrate some of the same principles that we have observed with MHC-I molecules. Elegant peptide binding studies demonstrated that binding of low affinity peptides to MHC-II maintains or promotes a MHC-II conformation that is “active” (61) or “peptide friendly” (62), i.e., peptide receptive. These studies indicate that the rate of peptide binding to empty MHC-II is much slower than peptide binding to MHC-II molecules that have previously bound a low affinity variant peptide, implying that stabilizing peptides promote a peptide-receptive MHC conformation. This parallels our observation that stabilizing peptide can promote the subsequent binding of readout peptide to MHC-I molecules. Furthermore, the studies of MHC-II showed that the enhancement of readout peptide binding was linked to the rate of stabilizing peptide dissociation, similar to our observation that highly stable complexes of stabilizing peptide with MHC-I did not enhance (or even decreased) subsequent readout peptide binding. Thus, both MHC-I and MHC-II molecules use stabilizing peptides to maintain a peptide-receptive state and enhance their subsequent ability to bind readout peptides. As peptide exchange occurs, both MHC-I and MHC-II molecules may undergo peptide editing until a very high affinity peptide generates a highly stable complex for presentation at the cell surface.

We conclude that peptide dissociation/exchange plays an important role in the genesis of peptide-receptive MHC-I molecules used for alternate MHC-I Ag processing. Endogenous stabilizing peptides may be transported by TAP to promote assembly of stabilizing peptide-MHC-I complexes in the ER, their exit to post-Golgi compartments, and their persistence as precursors to peptide-receptive MHC-I molecules. Alternate MHC-I Ag processing, then, involves the dissociation of stabilizing peptide and its replacement by readout peptide derived from exogenous Ag. This process is optimized at acidic pH, similar to conditions in post-Golgi vacuolar compartments that process exogenous Ags (e.g., phagolysosomes), suggesting that alternate MHC-I Ag processing involves binding of peptides derived from exogenous Ags to MHC-I molecules in these compartments.

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CORRECTIONS


An error was made in describing the protocol used for experiments shown in Figs. 5, 6, and 7. Text in the Materials and Methods and Results sections, along with figure legends for Figs. 5, 6, and 7 implied that antigenic readout peptide was only added subsequent to the incubation of macrophages in citrate-buffered saline at varying pH, but peptide was actually added at the indicated concentrations both during the incubation in citrate-buffered saline at varying pH and in the subsequent incubation for 2 h in normal medium. This correction does not change the conclusion of the paper, but is important for executing future studies.


In Fig. 1A, the last lane in the lower right panel was misplaced. It should have appeared as the first lane in the left side of the lower panel. The corrected figure is shown below.