Eosinophils Are a Major Source of Nitric Oxide-Derived Oxidants in Severe Asthma: Characterization of Pathways Available to Eosinophils for Generating Reactive Nitrogen Species

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Eosinophils Are a Major Source of Nitric Oxide-Derived Oxidants in Severe Asthma: Characterization of Pathways Available to Eosinophils for Generating Reactive Nitrogen Species

Jennifer C. MacPherson,* Suzy A. A. Comhair,‡ Serpil C. Erzurum,§ Dennis F. Klein,‖ Mary F. Lipscomb,¶ Mani S. Kavuru,§ Michael K. Samoszuk,‖ and Stanley L. Hazen2*†¶

Eosinophil recruitment and enhanced production of NO are characteristic features of asthma. However, neither the ability of eosinophils to generate NO-derived oxidants nor their role in nitrination of targets during asthma is established. Using gas chromatography-mass spectrometry we demonstrate a 10-fold increase in 3-nitrotyrosine (NO2 Y) content, a global marker of protein modification by reactive nitrogen species, in proteins recovered from bronchoalveolar lavage of severe asthmatic patients (480 ± 198 μmol/mol tyrosine; n = 11) compared with nonasthmatic subjects (52.5 ± 40.7 μmol/mol tyrosine; n = 12). Parallel gas chromatography-mass spectrometry analyses of bronchoalveolar lavage proteins for 3-bromotyrosine (BrY) and 3-chlorotyrosine (ClY), selective markers of eosinophil peroxidase (EPO)- and myeloperoxidase-catalyzed oxidation, respectively, demonstrated a dramatic preferential formation of BrY in asthmatic (1093 ± 457 μmol BrY/mol tyrosine; 161 ± 88 μmol ClY/mol tyrosine; n = 11 each) compared with nonasthmatic subjects (13 ± 14.5 μmol BrY/mol tyrosine; 65 ± 69 μmol ClY/mol tyrosine; n = 12 each). Bronchial tissue from individuals who died of asthma demonstrated the most intense anti-NO2 Y immunostaining in epitopes that colocalized with eosinophils. Although eosinophils from normal subjects failed to generate detectable levels of NO, NO2−, NO3−, or NO2 Y, tyrosine nitration was promoted by eosinophils activated either in the presence of physiological levels of NO2− or an exogenous NO source. At low, but not high (e.g., >2 μM/min), rates of NO flux, EPO inhibitors and catalase markedly attenuated aromatic nitrination. These results identify eosinophils as a major source of oxidants during asthma. They also demonstrate that eosinophils use distinct mechanisms for generating NO-derived oxidants and identify EPO as an enzymatic source of nitrating intermediates in eosinophils. The Journal of Immunology, 2001, 166: 5763–5772.

Eosinophils play a specialized role in innate host defense. These granulocytes are recruited to tissues to combat invading pathogens, such as during parasitic and helminthic infections (1–5). However, eosinophil recruitment is also a characteristic feature of asthma and other allergic inflammatory disorders (3–7). Although the function of eosinophils during host defense scenarios seems apparent, whether they play a beneficial role in allergy and asthma is less certain. Rather, substantial evidence supports the notion that eosinophils contribute to tissue injury and many of the pathophysiological features of asthma (3–7). Clinically, elevated eosinophil counts and the presence of eosinophil-secreted granular proteins such as major basic protein, eosinophil cationic protein, or eosinophil peroxidase (EPO)3 in sputum and biopsy samples have been used as biomarkers for monitoring the severity of asthmatic events (3–10). Though high levels of granular proteins are indicative of eosinophilia, leukocyte activation, and degranulation, little is known about the precise chemical mechanisms through which eosinophils contribute to tissue damage in the asthmatic lung and airways.

Eosinophils are unique among circulating leukocytes in their prodigious capacity to wage chemical warfare. As mentioned above, they are endowed with numerous highly basic and cytotoxic granule proteins that are released upon activation or during cell necrosis. They also possess an arsenal of enzymes designed to inflict oxidative damage upon biological targets (1–3). During eosinophil activation, a respiratory burst occurs where O2− and its dismutation product, H2O2, are formed (1, 2, 11). The respiratory burst of human eosinophils produces anywhere from 3 to 10 times as much O2− as a corresponding number of similarly treated neutrophils (12, 13). Although reduced oxygen species such as O2− and H2O2 do not effectively oxidize biological targets, one of the most abundant proteins secreted upon eosinophil activation, EPO, amplifies the oxidizing potential of H2O2 by using it as cosubstrate

References

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3 Abbreviations used in this paper: EPO, eosinophil peroxidase; BAL, bronchoalveolar lavage; Br-HPA, 3-(3-bromo-4-hydroxyphenyl)propanoic acid; BrY, 3-bromotyrosine; C1HPA, 3-(3-chloro-4-hydroxyphenyl)propanoic acid; ClY, 3-chlorotyrosine; DTPA, diethylenetriaminepentaacetic acid; GC-MS, gas chromatography-mass spectrometry; hICAT, heat inactivated catalase; HPA, 3-(4-hydroxyphenyl) propanoic acid; NO2−, NO3−, NO2 Y, 3-nitrotyrosine; PAPA NONOate, (Z)-N-(3-aminopropyl)-N-(propylamino)diacene-1-imid-1,2-diolate; RNS, reactive nitrogen species; SOD, superoxide dismutase.

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to generate potent reactive oxidants and diffusible radical species (2, 14–16).

EPO is a member of the mammalian peroxidase superfamily (17) and has a high degree of sequence homology with myeloperoxidase (MPO), an abundant peroxidase in neutrophils, monocytes, and certain tissue macrophages (18). Both EPO and MPO share the unique ability to use halides and pseudohalides as substrates to make highly reactive oxidants, hypohalous acids (HOX) (Equation 1).

\[ \text{H}_2\text{O}_2 + X^- + \text{H}^+ \rightarrow \text{HOX} + \text{H}_2\text{O} \]  

(1)

where \( X = \text{Cl}^-, \text{Br}^-, \text{I}^-, \) or SCN\(^-\).

At normal plasma levels of halides and pseudohalides (100 mM \( \text{Cl}^- \), 20–150 \( \mu \text{M} \text{Br}^- \), 69 \( \mu \text{M} \text{SCN}^- \), 0.1–0.6 \( \mu \text{M} \text{I}^- \) (Ref. 19), EPO preferentially uses the pseudohalide SCN\(^-\) (20) and Br\(^-\) as substrates (15, 21), whereas MPO uses Cl\(^-\) (22, 23). Recent gas chromatography-mass spectrometry (GC-MS) studies demonstrate that eosinophils use EPO to generate oxidants in allergen-triggered asthma (24). BrY (3-bromotyrosine), a specific marker of protein modification by reactive brominating species (25), was markedly enriched in bronchoalveolar lavage (BAL) proteins recovered from asthmatic subjects following exposure to segmental allergen challenge (24). Thus, one chemical pathway used by eosinophils to promote oxidative modification of proteins during asthma is through EPO-generated reactive brominating species.

Another potential pathway for oxidative modification of tissues in asthma by eosinophils may involve formation of reactive nitrogen species (RNS). NO production is increased in asthma (26–28). Although NO is a relatively long-lived radical that does not nitrate biological targets directly, it can be converted into more potent RNS. The most commonly studied NO-derived oxidant is peroxynitrite (ONOO\(^-\)), a product formed by the near diffusion-limited interaction of NO and \( \text{O}_2^- \) (29). ONOO\(^-\) nitrates protein tyrosine residues (29–31), and immunohistochemical studies suggest that 3-nitrotyrosine (NO2Y) is produced in asthmatic airways (32, 33). At physiological levels of \( \text{CO}_2/\text{HCO}_3^- \), ONOO\(^-\) exists predominantly as the more potent nitrating species, peroxycarboxy-nitrite (ONOOCO2\(^-\)) (Refs. 34–36; Equation 2).

\[ \text{NO} + \text{O}_2^- + \text{H}_2\text{O} \rightarrow \text{ONO}^- + \text{CO}_2 + \text{ONOOCO}_2^- \]  

(2)

Concomitant production of NO and \( \text{O}_2^- \) almost invariably occurs at sites of inflammation; consequently, potential adverse effects of excess NO production have been primarily attributed to formation of ONOO\(^-\) or ONOOCO2\(^-\) (29, 32, 33, 36, 37).

However, recent studies have identified alternative pathways for generating NO-derived oxidants that involve leukocyte peroxidases, van der Vliet and colleagues first demonstrated that MPO could use nitrite (\( \text{NO}_2^- \)), a major end-product of NO metabolism, as a substrate to nitrate tyrosine residues (38). Subsequent studies with isolated neutrophils (39, 40) and monocytes (41–43) have demonstrated that MPO-generated RNS can play a significant, and even predominant, role in oxidative modification of biomolecules. We recently demonstrated that EPO is significantly more effective than MPO at promoting protein nitration at physiologically relevant concentrations of halides and \( \text{NO}_2^- \) (44). Although increased levels of NO, \( \text{NO}_2^- \), \( \text{H}_2\text{O}_2 \), and eosinophils are present in asthmatic airways, neither the ability of eosinophils to generate NO-derived oxidants nor their role in nitration of targets during asthma are reported.

In this study we demonstrate that RNS play a significant role in oxidative modification of proteins in asthma. Using a combination of approaches including mass spectrometry, immunohistochemistry, and studies with isolated eosinophils, we show that eosinophils generate NO-derived oxidants both in vitro and in vivo. Finally, we examine the chemical pathways available to human eosinophils for generating NO-derived oxidants and demonstrate that the leukocytes form RNS by at least two mechanistically distinct pathways.

**Materials and Methods**

**General materials**

All solvents were purchased from Fisher Scientific (Pittsburgh, PA) and were Optima or HPLC grade. PAPA NONOate (2-[N-(3-aminopropyl)]-N-(o-propyl)amino)diazien-1-ium-1,2-diolate was obtained from Alexis (San Diego, CA). Glucose oxidase (grade II) and catalase were acquired from Roche Molecular Biochemicals (Indianapolis, IN). \( \text{t}-\text{[Cl}^-\text{]CO}_2\)tyrosine and \( \text{t}-\text{[Br}^-\text{]CO}_2\)tyrosine were obtained from Cambridge Isotope Laboratories (Andover, MA). Unless otherwise stated, all other chemicals were purchased from Sigma (St. Louis, MO).

**Subjects and sample collection**

Human endotracheal/bronchial aspirates were obtained from patients \( n = 11 \) who required mechanical ventilatory assistance due to respiratory failure from a severe asthmatic exacerbation. All asthmatic subjects had a history of a \( >12\% \) increase in absolute forced expiratory volume in 1 s (FEV1) and a 200-cc increase in forced expiratory volume either spontaneously or after bronchodilator within 1 year of hospitalization, and satisfied the definition of severe asthma as defined by the National Institutes of Health guidelines (45). Samples were collected within 12 h of admission because asthmatic subjects received i.v. corticosteroids upon presentation to the emergency department and subsequent transfer to the intensive care unit, and the effects of corticosteroids on protein oxidation products is unknown. All asthmatic individuals had a history of using inhaled \( \beta_2 \)-agonists either regularly \( (n = 8) \) or on an as-needed basis \( (n = 3) \) in the month preceding admission. Several also used inhaled corticosteroids \( (n = 8) \) and/or had received oral corticosteroids within 1 month of presentation \( (n = 4) \). Control subjects \( (n = 12) \) were age- and sex-matched, nonsmokers, and had no prior history of asthma or other lung disease. Endotracheal/bronchial aspirates were obtained from several control subjects who were either undergoing elective surgery \( (n = 3) \) or were admitted to the intensive care unit for a noninflammatory, nonrespiratory process (i.e., airway protection secondary to either head trauma \( (n = 2) \) or drug overdose \( (n = 2) \)).

The remaining clinical specimens from healthy nonasthmatic controls \( (n = 5) \) were obtained as residual specimens collected as baseline BAL samples for a separate clinical study (46). Healthy control subjects in that study all had a negative methacholine challenge test. Thus, all \( 11 \) specimens from asthmatic subjects were obtained as endotracheal/bronchial aspirates, and the \( 12 \) specimens from nonasthmatic controls were obtained as endotracheal/bronchial aspirates \( (n = 7) \) and baseline BAL \( (n = 5) \) specimens. No differences were noted in levels of NO2Y, BrY, or 3-chlorotyrosine (ClY) (per mol tyrosine) in endotracheal/bronchial aspirates vs BAL; consequently, specimens from the nonasthmatic subjects were combined as a single control group. Cells in clinical specimens were removed by centrifugation, the supernatants were supplemented with a mixture of antioxidants and peroxidase inhibitors (final concentration: 200 \( \mu \text{M} \) diethylthiencarboxylic-pentaacetic acid (DTPA), 100 \( \mu \text{M} \) butylated hydroxyltoluene, 50 mM sodium phosphate \((\text{pH} 7.0)\), 1 m\( \text{M} \) aminotriazole), capped in vials under argon atmosphere, snap frozen in liquid \( \text{N}_2 \), and stored at \(-80\)°C until time of sample preparation and mass spectrometry analysis.

**Histological analysis of lung and bronchial tissues**

Histological sections were cut from paraffin blocks of lung and bronchial tissues obtained from four individuals, who died from asthma, and four age-matched individuals who died of nonpulmonary processes. For NO2Y immunostaining, slides were initially incubated at 37°C with 0.01 mg/ml protease K for 15 min. Following wash with PBS containing 0.5 mM levamisole, the tissue was treated with 1% BSA in PBS to block nonspecific binding, then incubated for 2 h with immunoperoxidased polyclonal Ab directed against NO2Y (1:150 diluted in 1% BSA/PBS; Upstate Biotechnology, Lake Placid, NY). Following wash with PBS/0.5 mM levamisole, the tissue was incubated with a biotin-conjugated secondary Ab (Dako, Carpinteria, CA) for 10 min. Washing was followed by another 10-min incubation with alkaline phosphatase-labeled streptavidin (Dako). Immunostaining was visualized with an alkaline phosphate substrate solution containing naphthol AS-MX phosphate, Fast Red, and levamisole in Tris buffer \((\text{pH} 8.2)\) (Dako) and counterstained with the nuclear stain, hematoxylin. Negative control experiments involved either immunoadsorption of NO2Y Ab with 3.75 mM NO2Y before incubation with tissue sections or incubating the section with isotype-control nonimmunogenic Ig of the same subclass as the test Ab.
primary Ab. In separate studies, the specificity of the primary Ab was confirmed by observing loss of nitrotyrosine-specific recognition following reduction of nitrotyrosine-containing protein with dithionite. All biopsies were stained on the same day. Specific autofluorescence imaging of EPO in biopsies was performed as described (47–49). EPO-specific in situ peroxidase staining of tissues was performed on anti-nitrotyrosine-stained tissue sections following treatment of slides with 0.01 M KCN to inhibit MPO (50, 51).

Eosinophil and neutrophil isolation

Human eosinophils were isolated by negative selection using CD16 microbeads (Miltenyi Biotec, Auburn, CA), as described (52). Human neutrophils were isolated by buoyant density centrifugation (53), and low levels of contaminating eosinophils were then removed by fluorescence-activated cell sorting (54). The final purity of cell preparations was confirmed by flow cytometry using selective Abs for cell surface Ag on eosinophils (CD49d) and neutrophils (CD16), respectively (54). No detectable cross-contamination of peroxidase activity in detergent extracts of leukocyte preparations was observed following SDS-PAGE with in-gel tetramethylbenzidine staining (55). Trypan blue exclusion tests demonstrated over 97% viability in eosinophil and neutrophil preparations.

Cell experiments

These studies were performed in the presence of CO2 (5% gas phase) and HCO3− in the medium (4.2 mM) to more closely mimic a biologically relevant situation. Leukocytes (1 × 10⁶/ml) were incubated at 37°C under 95% air, 5% CO2, in Medium A (Ca2+/Mg2+/phenol-free HBSS; Life Technologies, Gaithersburg, MD) supplemented with 100 μM NaBr, 50 μM methionine (Met), or 1 mM L-[13C9]tyrosine as internal standards (24). The NO2− content of cells was determined by addition of PMA (200 nM) and incubated for either 1 h or the indicated time interval. In some experiments, eosinophils were activated in the presence of an exogenous NO source by addition of PAPA NONOate (Alexis) for 1 min following PMA addition. Rates of NO flux were determined spectrophotometrically by reaction of NO with oxyhemoglobin (56) under the identical conditions used for experiments, but in the absence of any added cells. To maintain a final pH of 7.4 during experiments with PAPA NONOate, incubations were performed in Medium B (Medium A containing only 100 mM NaCl and supplemented with 20 mM sodium phosphate, pH 7.4). Peroxidase activity was detected spectrophotometrically by reaction of NO with oxyhemoglobin (56) under the same conditions used. In some cases, leukocyte reaction mixtures also contained one of the following: 1 mM NaCl, 10 mM 3-aminoazirone (Atz), 300 mM catalase (Cat), 300 nM heat-inactivated catalase (hiCat), 10 μg/ml superoxide dismutase (SOD), 1 mM methionine (Met), or 1 mM N7-acetyl lysine.

Quantification of leukocyte-generated products in vitro

NO2− production by isolated eosinophils was quantified by reversed phase HPLC with photodiode array detection (44). Peak identity was routinely confirmed by demonstrating the appropriate UV-VIS absorbance spectrum of the peak that comigrated with authentic NO2−. In preliminary studies, NO2− production by eosinophils was also independently confirmed by HPLC with on-line electrospary ionization mass spectrometry, similar to prior studies using isolated EPO (44). The nitrated (NO2−-HPA; 3-(4-hydroxy-3-nitrophenyl)propanoic acid) and chlorinated (Cl-HPA; 3-(3-chloro-4-hydroxyphenyl)propanoic acid) and brominated (Br-HPA; 3-(3-bromo-4-hydroxyphenyl)propanoic acid) products of the tyrosine analog, HPA, were routinely quantified by reversed phase HPLC with electrospary (chemo)metric detection on an ESA CoulArray HPLC (Cambridge, MA) equipped with UV detector and electrochemical cells (eight channels) (44). Peak identity was established by demonstrating the appropriate retention time, mass, and potential of integrated currents in adjacent channels, and by the method of standard additions for each analyte. Authentic standards of NO2−-HPA, Br-HPA, and CI-HPA were prepared by reaction of HPA with a molar equivalent of ONOO−, HOBr, or HOCl, respectively. Standards were then isolated by reversed phase HPLC, and their structures were confirmed by electrospray ionization mass spectrometry by demonstrating the appropriate mass-to-charge ratio (and isotopic cluster, where applicable) of the anticipated molecular ion of the isolated product.

Sample preparation and mass spectrometry

The contents of Br and CI in proteins present in clinical specimens were determined by stable isotope dilution GC-MS using 3-bromotoluene as internal standard (24). The NO2− content of

All water used to prepare buffers and medium was pretreated with Chelex-100 resin (Bio-Rad, Hercules, CA) and supplemented with 100 μM DTPA to remove trace levels of potential redox-active transition metal ions. Superoxide generation by activated human eosinophils was measured as the SOD-inhibitable reduction of ferricytochrome c (59). Quantification of NO2− and NO− was performed by anion exchange HPLC with UV detection at 210 nm under argon atmosphere. Products were resolved on a Spherisorb SS SAX column (24 cm × 4.6 mm, 5 μm; Phase Separations, Norwalk, CT) under isocratic conditions using 45 mM sodium phosphate (pH 3.0) as the mobile phase.

Leukocytes were isolated from whole blood of healthy volunteers after obtaining informed consent. Tissue sections were obtained from the New Mexico Office of the Medical Examiner and the Anatomic Pathology Department at the Cleveland Clinic Foundation. Sections were anonymized by the forensic pathologists for use in these studies. All protocols were in accordance with institutional guidelines of either the University of New Mexico School of Medicine or the Cleveland Clinic Foundation and were approved by their respective Institutional Review Committees.

Statistics

Data represent the mean ± SD of the indicated number of samples. Statistical analyses were made using a paired Student’s t test. For all hypotheses the significance level was 0.05. When multiple comparisons were made, a Bonferroni correction to the significance criterion for each test was made.

Results

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(p < 0.0001) 10-fold increase in protein NO₂ Y levels was observed in samples from severe asthmatic patients compared with levels present in nonasthmatic subjects (480 ± 198 vs 52.5 ± 40.7 µmol/mol tyrosine; asthmatic vs nonasthmatic, respectively; Fig. 1).

**Immunohistochemical studies colocalize nitrotyrosine with eosinophils in bronchial tissues from severe asthmatics**

To identify the potential cellular source(s) of NO-derived oxidants in severe asthma, specimens from subjects who died from asthma (status asthmaticus) were examined using affinity-purified Ab specific for NO₂ Y. Intense staining that colocalized with eosinophils was typically observed in the majority of specimens (Fig. 2). Diffuse staining of epithelial cells was also commonly observed, as has previously been reported (32, 33). Both in situ fluorescence microscopy specific for the heme group of EPO (data not shown) and in situ peroxidase staining specific for EPO were also abundant in eosinophil-rich areas of specimens from asthmatics. Double staining of sections for both NO₂ Y- and EPO-specific in situ peroxidase staining confirmed colocalization of NO₂ Y with eosinophils in the submucosa of airways from severe asthmatic subjects (Fig. 2).

**Eosinophils are a major cellular mediator of protein oxidation in severe asthma**

To assess the relative contributions of eosinophils and neutrophils in the oxidative modification of proteins in severe asthma, the protein content of BrY and ClY, molecular fingerprints for eosinophil- and neutrophil-mediated tissue damage, respectively (24, 60), were determined in the same clinical specimens evaluated for protein NO₂ Y content in Fig. 1. There was a striking 84-fold elevation (p < 0.0001) in the content of BrY observed in proteins recovered from airways of asthmatic (1093 ± 457 µmol BrY/mol tyrosine) vs nonasthmatic subjects, whose levels were near the limit of detection (13 ± 14.5 µmol BrY/mol tyrosine) (Fig. 3A). There was also a significant 3-fold increase (p < 0.05) in ClY in airway proteins recovered from severe asthmatics (161 ± 88 µmol ClY/mol tyrosine) over nonasthmatics (65 ± 69 µmol ClY/mol tyrosine) (Fig. 3B). A comparison of the BrY/ClY ratios, an indication of the relative preferential contribution of eosinophils vs...
neutrophils toward oxidation of proteins, revealed a 30-fold difference in asthmatics compared with nonasthmatics (ratio of 6.8 vs 0.2 for asthmatic vs nonasthmatic, respectively).

**Human eosinophils nitrate tyrosine in the presence of physiological levels of halides and NO₂⁻**

We recently demonstrated that isolated EPO effectively uses NO₂⁻ at concentrations comparable to those observed in inflammatory tissues and fluids (≤50 μM; Ref. 38 and references therein) in cell-free systems to generate a RNS capable of promoting tyrosine nitration (44). Based upon these studies, we hypothesized that eosinophils might use EPO to contribute to protein oxidation through nitration in asthma. However, the ability of human eosinophils to generate NO-derived oxidants has not yet been demonstrated. To determine whether eosinophils can generate NO-derived oxidants, we isolated peripheral cells from normal healthy donors and incubated them in medium containing L-tyrosine, plasma levels of halides (100 mM Cl⁻), and the agonist PMA. Analysis of medium revealed that no significant NO₂⁻Y was formed (Fig. 4A). Moreover, we were also unable to detect any endogenous NO production or significant (i.e., >1 μM) NO₂⁻ /NO₃⁻ accumulation by human eosinophils freshly isolated from peripheral blood, with or without phorbol ester activation, under the conditions and time course used (data not shown). In contrast, eosinophils activated in Medium A supplemented with pathophysiologically relevant levels of NO₂⁻ (50 μM) readily produced NO₂⁻Y (Fig. 4A). The time course for NO₂⁻Y formation paralleled the time course for O₂⁻ production during a respiratory burst (Fig. 4B). Finally, in separate studies, eosinophils were activated with an alternative agonist, N-formyl-methionyl-leucyl-phenylalanine (100 nM). Cell-dependent NO₂⁻Y formation again demonstrated an absolute requirement for exogenous NO₂⁻, although NO₂⁻Y levels produced were ~8-fold less than that observed with eosinophils stimulated with PMA (data not shown).

The absolute requirement of NO₂⁻ for NO₂⁻Y formation suggested that, under the conditions used, eosinophils were generating RNS via the EPO-H₂O₂-NO₂⁻ system. To further explore the reaction mechanism for eosinophil-mediated aromatic nitration reactions, isolated human eosinophils were incubated in medium containing plasma levels of halides (100 mM Cl⁻, 100 μM Br⁻), levels of NO₂⁻ observed in epithelial lining fluid of severe asthmatics (50...
Eosinophils promote nitrotyrosine formation in asthma

Figure 7. Eosinophil and neutrophil modification of HPA at physiologically relevant levels of nitrite. Isolated human (A) eosinophils and (B) neutrophils (1 × 10⁶ cells/ml) were incubated for 1 h at 37°C in magnesium-, calcium-, and phenol-free HBSS supplemented with 100 μM HPA, 100 μM NaBr, and the indicated concentrations of NaN₃. Cells were then pelleted and the content of NO₂-HPA, Br-HPA, and CI-HPA generated by eosinophils and neutrophils was then determined as described in Materials and Methods. Data represent the mean ± SD of triplicate determinations for a characteristic experiment performed at least three times.

Figure 8. Investigation of the potential pathways available to eosinophils for generating NO-derived oxidants. Phorbol ester-stimulated human eosinophils (1 × 10⁶ cells) were incubated in calcium-, magnesium-, and phenol-free HBSS containing 100 μM NaBr, 100 μM HPA, and the indicated rates of NO release from an exogenous NO source, PAPA NONOate, and plasma-brain HCl (Equation 2). Based on the inability to detect NO, NO₂-, or NO₃- accumulation in cell medium (see above), this likely reflects the limited capacity of freshly isolated peripheral blood eosinophils from healthy (nonallergic) donors to generate NO, particularly within the brief time period of the respiratory burst (≈1 h, Fig. 4B). However, enhanced levels of NO in expired breath of asthmatics is well documented, and expression of NO synthase isoforms in the airways is likewise established (26–28). Therefore, we performed a series of experiments in which eosinophils were activated in the presence of an exogenous NO-generating system, PAPA NONOate, and plasma-brain HCl (Equation 2). Aromatic nitration by eosinophils required cell activation, consistent with a requirement for reduced oxygen species (O₂⁻ and/or H₂O₂) for oxidation. Moreover, nitration was the favored biochemical pathway for oxidative modification at all but the lowest levels of NO flux examined. At the higher fluxes of NO (≥2 μM/min) examined, the overall extent of NO₂-HPA formation diminished (Fig. 8).

Eosinophils generate NO-derived oxidants by at least two mechanistically distinct pathways

One striking feature of the results thus far described was that we could find no evidence that isolated eosinophils generate nitrating intermediates through formation of ONOO⁻ (Equation 2). Based on the inability to detect NO, NO₂-, or NO₃- accumulation in cell medium (see above), this likely reflects the limited capacity of freshly isolated peripheral blood eosinophils from healthy (nonallergic) donors to generate NO, particularly within the brief time period of the respiratory burst (≈1 h, Fig. 4B). However, enhanced levels of NO in expired breath of asthmatics is well documented, and expression of NO synthase isoforms in the airways is likewise established (26–28). Therefore, we performed a series of experiments in which eosinophils were activated in the presence of an exogenous NO-generating system, PAPA NONOate, and plasma-brain HCl (Equation 2). Aromatic nitration by eosinophils required cell activation, consistent with a requirement for reduced oxygen species (O₂⁻ and/or H₂O₂) for oxidation. Moreover, nitration was the favored biochemical pathway for oxidative modification at all but the lowest levels of NO flux examined. At the higher fluxes of NO (≥2 μM/min) examined, the overall extent of NO₂-HPA formation diminished (Fig. 8).

Eosinophils are more efficient than neutrophils at promoting aromatic nitration reactions

In the next series of experiments we compared the ability of purified human eosinophils and neutrophils to nitrate, chlorinate, and brominate the tyrosine analog HPA in the presence of plasma levels of halides over the (patho)physiologically relevant range of NO₂⁻ (0–50 μM). Eosinophils generated significantly more NO₂-HPA (>5-fold) than an equivalent number of neutrophils at all concentrations of NO₂⁻ examined (Fig. 7). At levels of NO₂⁻ observed in epithelial lining fluid from normal individuals (i.e., <10 μM NO₂⁻), eosinophils were more effective at oxidation of phenolic groups through bromination. As NO₂⁻ levels became elevated into the pathophysiological range (≥10 μM), nitrination of targets predominated (Fig. 7A). In stark contrast to neutrophils, no significant oxidation of HPA through chlorination was observed under all conditions examined (Fig. 7). Finally, neutrophils were ineffective at oxidizing the tyrosine analog through bromination under all conditions examined (Fig. 7).

Materials and Methods

In contrast, nitration reactions were not blocked by the presence of the hydrogen peroxide scavenger methionine to affect nitration, or the EPO-H₂O₂-NO₂⁻ accumulation in cell medium (see above), this likely reflects the limited capacity of freshly isolated peripheral blood eosinophils from healthy (nonallergic) donors to generate NO, particularly within the brief time period of the respiratory burst (≈1 h, Fig. 4B). However, enhanced levels of NO in expired breath of asthmatics is well documented, and expression of NO synthase isoforms in the airways is likewise established (26–28). Therefore, we performed a series of experiments in which eosinophils were activated in the presence of an exogenous NO-generating system, PAPA NONOate, and plasma-brain HCl (Equation 2). Aromatic nitration by eosinophils required cell activation, consistent with a requirement for reduced oxygen species (O₂⁻ and/or H₂O₂) for oxidation. Moreover, nitration was the favored biochemical pathway for oxidative modification at all but the lowest levels of NO flux examined. At the higher fluxes of NO (≥2 μM/min) examined, the overall extent of NO₂-HPA formation diminished (Fig. 8).

Aromatic nitration reactions mediated by eosinophils in the presence of an exogenous NO donor might occur via formation of either ONOO⁻ (and/or ONOO⁻CO⁻) or the EPO-H₂O₂-NO₂⁻...
system. To gain insights into the pathway(s) used by eosinophils to generate NO-derived oxidants, we examined the effects of EPO inhibitors, H$_2$O$_2$ scavengers, and SOD on NO$_2$-HPA formation. At low rates of NO flux (<2 μM/min), NO$_2$-HPA formation was inhibited by azide, a heme poison that inhibits EPO catalysis (Fig. 9). Under these same conditions, catalase, but not hiCat, significantly attenuated NO$_2$-HPA production, consistent with a role for H$_2$O$_2$ in the aromatic nitration reaction. In contrast, at high rates of NO flux (>2 μM/min), aromatic nitration by activated eosinophils became increasingly less sensitive to inhibition by either azide or catalase. Finally, while addition of SOD to reactions demonstrated a modest increase in NO$_2$-HPA formation at low levels of NO flux, inhibition in nitration was observed at the higher rates of NO flux examined (Fig. 9).

**Discussion**

Enhanced production of NO is widely observed in the exhaled breath of asthmatics (26–28). Because of the ability of NO to generate potent nitrating intermediates, it has been suggested that RNS contribute to inflammatory tissue injury in asthma. However, remarkably few studies have actually directly examined the role of RNS in oxidation of biological targets in human asthma. Evidence thus far reported is primarily limited to studies that rely upon immunohistochemical detection of NO$_2$Y (32, 33). Because of the inherent uncertainties in the precise cognate epitope recognized by immunohistochemical methods and their nonquantitative nature, we sought to use more direct chemical methods to establish that RNS contribute to oxidative modification of proteins in asthma.

The results of this study directly demonstrate that RNS contribute to oxidative modification of proteins in asthma. They also strongly support a role for eosinophils in generating NO-derived oxidants in severe asthma. Using a specific and sensitive mass spectrometric method, we observe a 10-fold increase in the content of NO$_2$Y in proteins recovered from airways of patients with severe asthma compared with nonasthmatic subjects (Fig. 1). Eosinophil activation and subsequent protein oxidation in the asthmatic airways is also supported by the dramatic increase in BrY content observed in airway proteins recovered from asthmatic subjects admitted to the intensive care unit (Fig. 3). Moreover, immunohistochemical studies implicate eosinophils as a major cellular generator of NO-derived oxidants in severe asthma because epitopes demonstrating NO$_2$Y-specific immunostaining substantially colocalize with the leukocyte (Fig. 2). Finally, studies with isolated human eosinophils reveal that aromatic nitration reactions are a preferred oxidation pathway mediated by the leukocytes at plasma concentrations of halides and levels of nitrate observed in airway lining fluid of asthmatics (Figs. 6 and 7).

One of the remarkable findings of these studies is that human eosinophils can use distinct chemical mechanisms for generating NO-derived oxidants and that the relative preference of eosinophils for promoting aromatic nitration vs halogenation reactions varies depending upon their environment during activation. Moreover, the environment in which eosinophil activation occurs (e.g., rate of NO production) also influences the relative contribution that chemically distinct oxidation pathways play in contributing to protein nitration. On the basis of these results and recently published studies (24, 25, 44), we have generated the following model (Fig. 10) of pathways available to human eosinophils for oxidizing proteins through formation of reactive halogen and RNS. During eosinophil activation, such as following allergen challenge, the phagocyte undergoes a respiratory burst, generating O$_2^-$ and its dismutation product, H$_2$O$_2$. Concomitantly, the contents of the secretory granules are released, including EPO. EPO (15 μg/10^6 eosinophils) constitutes ~25% of the total protein mass of specific granule protein, and is thus one of the most abundant proteins in eosinophils (61). When human eosinophils are activated in medium possessing plasma levels of halides and low levels of NO (and its autoxidation product, NO$_2^+$), eosinophils preferentially use the EPO-H$_2$O$_2$ system to generate brominating oxidants (Figs. 6 and 8). Aromatic bromination reactions mediated by the activated cells are inhibited in the presence of the HOBr scavenger methionine, a thioether, but not by large molar excesses of primary amines (Fig. 6). These results are consistent with our prior observations that N-mono-bromoamines are preferred brominating intermediates for protein tyrosine residues in cell-free systems at neutral pH (44). The ability of N-mono-bromoamines to promote phenolic bromination reactions across the (patho)physiological pH range also likely contributes to the relatively high levels of protein BrY observed in endotracheal/bronchial aspirates recovered from patients with severe asthma (Fig. 3).

Under these same conditions (i.e., low levels of NO flux (<2 μM/min), plasma levels of halides), the predominant pathway for

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**FIGURE 9.** Effect of EPO inhibitor, H$_2$O$_2$ scavenger, and SOD on eosinophil-mediated aromatic nitration reactions. Phorbol ester-stimulated human eosinophils (1 × 10^6 cells) were incubated in calcium, magnesium, and phenol-free HBSS containing 100 μM NaBr, 100 μM HPA, and the indicated rates of NO release from an exogenous NO source, PAPA NONOate, as in Fig. 8 (control condition). In parallel reactions, cells were also incubated with the indicated additions. Reactions were stopped by removal of cells by centrifugation at 4°C, and the content of NO$_2$-HPA was determined. Data represent the mean ± SD of triplicate determinations of a representative experiment and are expressed as the percentage of NO$_2$-HPA formed in the presence of the indicated additions relative to that observed under control conditions. Similar results were observed in three independent experiments. Concentrations of additives, when present, were as follows: 1 mM azide (NaN$_3$), 300 nM catalase (+Cat), 300 nM hiCat (+hiCat), 10 μg/ml SOD.

**FIGURE 10.** Model of potential pathways used by eosinophils for the generation of NO-derived oxidants and reactive halogen species.
EOSINOPHILS PROMOTE NITROTYROSINE FORMATION IN ASTHMA

generating NO-derived oxidants by eosinophils appears to be via oxidation of NO$_2^-$ by the EPO-H$_2$O$_2$ pathway (Fig. 10). The ability of human eosinophils to use this pathway is strongly supported by the demonstration that activated eosinophils (this study) and isolated EPO (44) use physiologically relevant levels of NO$_2^-$ to promote nitration of phenolic targets as a preferred activity. A primary role for EPO- and H$_2$O$_2$-dependent formation of RNS by eosinophils is further supported by 1) the absolute requirement of NO$_2^-$ for aromatic nitration reactions (Figs. 4 and 7); and 2) the ability of EPO inhibitors (NaCl) and H$_2$O$_2$ scavengers (catalase, but not hiCat) to block phenolic nitration (Figs. 6 and 9). The RNS formed by EPO-catalyzed oxidation of NO$_2^-$ has not been identified. Studies with the related hemoprotein MPO suggest that the one electron oxidation product, nitrogen dioxide ($\cdot$NO$_2$), is a likely product (38, 40, 62). It should also be noted that isolated human EPO promotes nitration of free and protein-bound tyrosine residues at least 4- to 5-fold more efficiently than human MPO (on an equimolar basis, which corresponds to a 8- to 10-fold difference per heme group) (44).

Another remarkable finding of this study is the overall high yield of tyrosine nitration by eosinophils via the EPO-H$_2$O$_2$-NO$_2^-$ pathway. If one assumes that 2 mol of O$_2^-$ are required to form 1 mol of H$_2$O$_2$, during the respiratory burst, and 2 mol of H$_2$O$_2$ are required per mol of NO$_2^-$ formed (one to generate NO$_2^-$ and another to generate tyrosyl radical; Ref. 63), then the overall yield of eosinophil-dependent nitration of tyrosine via the EPO-H$_2$O$_2$-NO$_2^-$ system was ~6.6% under the conditions used in Fig. 4. Finally, although formation of a nitrating and halogenating species through secondary oxidation of NO$_2^-$ by hypohalous acids has been suggested for the MPO-generated product HOCl (64), eosinophil-mediated nitration of HPA in the presence of NO$_2^-$ was not blocked by addition of scavengers of halogenating oxidants (thiolethers and amines, Fig. 6). Thus, eosinophil-generated HOBBr does not appear to play a significant role in subsequent formation of NO-derived oxidants by these cells.

Isolated peripheral blood eosinophils from normal healthy donors failed to generate detectable levels of NO, NO$_2^-$, or NO$_3^-$ during the brief incubation periods used in this study. Consistent with these results, eosinophil activation in the absence of an exogenous source of either NO$_2^-$ or NO failed to nitrate phenolic targets (Figs. 4, 7, and 8). However, eosinophil activation in vivo occurs in an environment replete in NO, whether generated by cytokine-stimulated eosinophils or adjacent cells. As noted above, in the presence of an exogenous NO donor, eosinophils readily promoted aromatic nitration reactions (Figs. 8 and 9). At high rates of NO flux (>2 µM/min), eosinophils appear to use a pathway for generating RNS that is chemically distinct from the EPO-H$_2$O$_2$-NO$_2^-$ system because EPO inhibitors and H$_2$O$_2$ scavengers do not significantly inhibit phenolic nitration reactions under these conditions, yet addition of SOD does (Fig. 9). These results are consistent with interaction of NO with cell-generated O$_2^-$ forming ONOO$^-$ and ONOOCO$_2^-$ (Equation 2; Fig. 10). The relative contribution of the ONOO$^-$/ONOOCO$_2^-$ vs the EPO-H$_2$O$_2$-NO$_2^-$ system for eosinophil-dependent generation of NO-derived oxidants during asthma is unknown. However, the high levels of BrY observed in BAL proteins from severe asthmatics suggest that the EPO pathway may play a significant role in protein oxidation and, therefore, NO$_2^-$Y formation. At the levels of NO$_2^-$ typically observed in epithelial lining fluid during severe asthma (>10 µM), nitration of phenolic targets by the EPO system of eosinophils is preferred (Fig. 7). Finally, it should be noted that a wide range of steady-state NO levels are observed in vivo (20 nM-2 µM; Ref. 30), and NO-dependent protein nitration was mediated by activated eosinophils over this range (Fig. 8).

One interesting finding in this study is that the overall yields of nitration and bromination decline at higher fluxes (>2 µM/min) of NO (Fig. 8). Similar bell-shaped curves for the overall extent of protein and lipid oxidation mediated by leukocytes (e.g., neutrophils and monocytes) activated in the presence of varying levels of NO recently have been reported (39, 41). One possible explanation is that NO modulates the extent of oxidation by interacting with critical heme proteins involved in O$_2^-$ formation (e.g., NADPH oxidase) or RNS formation (e.g., EPO). Recent studies demonstrate that NO modulates the catalytic activity of the related peroxidase MPO (65). At low levels, NO serves to enhance catalytic rates by accelerating the rate limiting step in the peroxidase cycle, reduction of Compound II (65, 66). At higher levels of NO, MPO activity is inhibited by formation of a ferric-nitrosyl complex (65). Similar behavior may occur during NO interactions with EPO. Another potential mechanism accounting for the inhibition in the extent of oxidation at higher fluxes of NO is that it may partially act as an antioxidant under these conditions by scavenging reactive intermediates though radical-radical interactions (67). Alternatively, decreased formation of nitrotyrosine at high fluxes of NO may be due to lower overall yields with ONOO$^-$/ONOOCO$_2^-$ compared with the RNS formed by the EPO-H$_2$O$_2$-NO$_2^-$ system.

Recent studies by Gaston and colleagues demonstrate that airway vapor condensates from severe asthmatic subjects are acidic (68). This has led to the suggestion that elevated levels of NO observed in asthmatic subjects may arise in part from protonation of NO$_2^-$ (68, 69). Although the pH of airway lining fluids in the subjects examined in this study were not determined, it is likely that they were acidic as well. Therefore, one might speculate that some of the NO$_2^-$Y formed on BAL proteins during asthma arose from nonenzymatic formation of RNS. However, several lines of evidence suggest that eosinophils are a major source of NO-derived oxidants in vivo and that EPO plays an active role in NO$_2^-$Y formation during asthma. Perhaps the strongest evidence is the intense focal staining observed in bronchial biopsies probed with Abs specific for NO$_2^-$Y where the immunostaining predominantly colocalized with eosinophils (Fig. 2). Moreover, using mass spectrometry, we recently demonstrated that significant levels of bromination (25) and nitration (44) of target proteins incubated in medium containing plasma levels of halides and pathophysiologically relevant levels of NO$_2^-$ does not occur through nonenzymatic processes to any significant degree over the pH range observed in normal and asthmatic human airways (pH 5.5–7.5). Moreover, analysis of halogenated tyrosine adducts in protein present in lavage fluids from this study revealed a much higher proportion of BrY than CIY in severe asthmatics vs controls subjects (Fig. 3), despite the 1000-fold higher concentration of Cl$^-$ observed in plasma. Given that eosinophils selectively brominate while neutrophils chlorinate proteins at plasma levels of halides (Fig. 7 and Ref. 24), these in vivo data strongly support the notion that eosinophils are likely a major leukocyte responsible for promoting oxidative modification of proteins in asthmatic patients. The calculation of the BrY/Ciy ratio may thus represent an objective and quantifiable index to estimate the relative contributions of eosinophils vs neutrophils in oxidative modification of proteins in tissues. Finally, these studies with isolated human eosinophils clearly demonstrate that the leukocytes readily form NO-derived oxidants under physiologically relevant conditions.

Measurement of NO$_2^-$Y is now widely used as a marker for protein oxidation by RNS. A multitude of techniques for quantification of NO$_2^-$Y in biological tissues and fluids are used, including immunossay and a variety of HPLC and mass spectrometry-based methods. The validity of many of these methods has been
questioned because of the ease with which artificial phenolic nitration occurs in the presence of nitrite and acid pH. Only mass spectrometry combined with isotope-labeling techniques permits simultaneous monitoring of authentic oxidized amino acids (e.g., NO2Y) and in vitro reactivity of EPO-generated oxidants undoubtedly plays a significant role in the cytotoxic properties of these agents on patho-
gens and eukaryotic cells. Thus it is reasonable to speculate that oxidative modification of critical biological targets in asthmatic subjects in this study are extremely low, similar to those recently observed on induced sputum of normal subjects (72) and in other tissues and fluids examined using mass spectrometry-based methods that permit development of sample preparation methods that minimize artifactual nitrination (70, 71).

It should be noted that the biological consequences of protein oxidation via nitration or bromination during asthma are not known. However, it should also be appreciated that the stable covalent adducts of tyrosine monitored represent only a fraction of the total modifications incurred during exposure of proteins to re-active nitrogen or halogenating species. Other nucleophilic targets on proteins including thiol (cysteine), triothiole (methionine), im-
idaazole (histidine), and indole (tryptophan) groups are all targets for oxidation by either brominating or nitrating agents. The chemical reactivity of EPO-generated oxidants undoubtedly plays a sig-
ficant contribution of ex vivo nitration (as well as bromination and chlo-
ration in human asthma.

References


