Role for IgE in Airway Secretions: IgE Inducers Than Antigen Alone of Airway Inflammation in a Murine Model

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Role for IgE in Airway Secretions: IgE Immune Complexes Are More Potent Inducers Than Antigen Alone of Airway Inflammation in a Murine Model

Riaz I. Zuberi, John R. Apgar, Swey-Shen Chen, and Fu-Tong Liu

IgE is present in airway secretions from human patients with allergic rhinitis and bronchial asthma. However, the contribution of IgE present locally to the overall airway inflammation is not well understood. We hypothesize that Ag-specific IgE can capture airborne Ags and form immune complexes. These immune complexes may function as potent inducers of immune responses in the lung, contributing to the perpetuation of airway inflammation. BALB/c mice were first sensitized with OVA in alum systemically and then challenged with nebulized OVA. Bronchoalveolar lavage (BAL) fluid from these mice contained significant amounts of IgE, of which >50% was Ag specific. The IgE levels in airway secretions remained elevated for more than 15 days after the termination of Ag exposure. Significant amounts of IgE-OVA immune complexes were detected in BAL fluid from the OVA-challenged mice. For comparison of IgE immune complexes vs Ag alone, we treated OVA-immunized mice with intranasal administration of trinitrophenyl-OVA or trinitrophenyl-OVA-anti-DNP IgE. Those treated with the immune complexes showed significantly higher levels of IL-4 and more pronounced eosinophilia in BAL fluid than did those receiving the Ag alone. The IgE immune complexes did not augment the inflammatory response in high affinity FcεRI receptor (FcεRI)-deficient mice. We conclude that IgE present in the airways can capture the Ag and that the immune complexes thus formed may augment allergic airway response in an FcεRI-dependent manner. Thus, IgE present in airway secretions may facilitate Ag-mediated allergic airway inflammation. The Journal of Immunology, 2000, 164: 2667–2673.

Immunoglobulin E has long been regarded as a major molecular component of atopic diseases, including asthma (1). Clinical studies have found a close association between asthma and serum IgE levels as well as IgE-dependent skin test reactivity to allergens (2). In addition, allergen-specific IgE has been detected in nasal mucosa from allergic patients (3), and IgE synthesis and heavy chain class switching to IgE occur in B cells from the nasal mucosa of allergic individuals (4). Furthermore, IgE is consistently present in airway secretions from human patients with allergic rhinitis and bronchial asthma (5) and is detected in nasal washes from allergic patients receiving nasal challenge with specific allergens (6). Allergen-specific IgE is detected in bronchoalveolar lavage (BAL) fluids in asthmatic patients obtained with or without bronchial provocation with the allergen (7). It is believed that mast cells present in airway tissue are sensitized by the allergen-specific IgE and, on exposure to a specific allergen, are activated to produce potent mediators that are responsible for some of the symptoms in allergic airway diseases. The functional significance of IgE found in airway secretions is, however, not well understood. In this study, we hypothesize that Ag-specific IgE can capture airborne Ags and form immune complexes. Further, these immune complexes may function as more potent inducers of immune responses in the lung and thus contribute to the perpetuation of airway inflammatory responses. We have used a mouse model of asthma to test this hypothesis.

Animal models of human asthma have drawn considerable attention in recent years. A mouse model has been particularly popular, in which mice are primarily sensitized with an Ag systemically and then challenged with the same Ag through the airways. The animals develop a Th2 response in the lung with prominent eosinophilia, production of Th2 cytokines, and development of airway hyperresponsiveness (8–10). This model has been valuable, especially in conjunction with the use of genetically engineered mice lacking specific gene products, in the elucidation of molecular and cellular mechanisms responsible for allergic airway inflammation and airway hyperresponsiveness. It is evident that T cells play an important role (11, 12). The roles of various cytokines, including IL-4 (13, 14), IL-5 (15, 16), IL-6 (17), IL-13 (18–20), and IL-16 (21) in promoting, and IL-12 (22, 23) in suppressing inflammation have been demonstrated. Consistent with the presence of a Th2-like response, elevation of serum IgE is also a prominent feature of this mouse model.

The airway inflammation developed in this model can be conveniently studied through analysis of BAL fluid obtained from the mice. Typically, allergic airway inflammation is accompanied by a prominent increase in the numbers of eosinophils and elevation of levels of Th2 cytokines, such as IL-4 and IL-5, in the BAL fluid. However, there is no report documenting the detection of IgE in the fluid in this mouse model. In this study, we found the presence of Ag-specific IgE in both free and Ag-complexed forms in BAL fluid from mice with allergic airway inflammation. We also identified a potential role for the IgE present in the airway secretions by demonstrating that in vitro-formed IgE immune complexes, when
administered through the airways, are more potent than Ag alone in inducing airway inflammatory responses in mice previously sensitized with the Ag.

Materials and Methods

Mice

Female BALB/cByJ mice (The Jackson Laboratory, Bar Harbor, ME) 6–8 wk of age were used in all experiments. High affinity IgE receptor (FceRI)-deficient (FceRI–/–) mice and Tg1101 mice with a defective murine FceRIα chain gene but carrying a human FceRIα gene were obtained from R.W. Johnson Pharmaceutical Research Institute, San Diego, CA. In the latter mice, mast cells express functional FceRI receptor that recognizes both murine and human IgE (24).

Reagents

Mouse monoclonal IgE specific for dinitrophenol (25) and that specific for a ragweed Ag (26) were obtained previously. Goat anti-mouse IgE and rabbit anti-mouse IgE were prepared as described previously (25), and were affinity purified by using mouse anti-ragweed Ag monocolonal IgE (26). Tritonxylated OVA (TNP-OVA) was prepared as described (27). The IgE immune complexes were prepared before use by mixing 20 μg anti-DNP IgE with either 2 or 4 μg TNP-OVA followed by incubation at room temperature for 30 min.

Immunization and challenge protocols

Both control and OVA mice were immunized with 10 μg of OVA (chicken egg albumin from Sigma, St. Louis, MO) in 2 mg aluminum hydroxide gel (alum) i.p. After 14 days, the mice were given aerosolized OVA for 30 min each day from days 1 to 6 on consecutive days. Control mice were exposed to aerosolized PBS, pH 7.2. This was accomplished by placing the mice in a Plexiglass chamber connected to a nebulizer that generates an aerosol mist. In other experiments, OVA-immunized mice were first anesthetized using metofane (Pitman-Moore, Washington Crossing, NJ) and then challenged by administering intranasally 25–50 μl OVA, TNP-OVA, or mixtures of TNP-OVA and IgE (the concentrations used for individual experiments are shown in Results).

Collection of BAL fluid

The BAL fluid was collected after various time intervals following airway Ag challenge. The mice were anesthetized by using pentobarbital (Sigma) at 100 μg per body weight, the trachea was cannulated, and a BAL was performed with 1 ml pyrogen-free PBS. The lavage fluid collected was centrifuged at 400 X g, and cells were separated from the fluid. The fluid was again centrifuged at 1000 X g to remove cellular debris and was then stored at −70°C until it was evaluated for its cytokine and Ig levels as well as the presence of IgE-OVA immune complexes.

Enumeration of cells

Total viable cell numbers were determined by trypsin blue exclusion using a hemocytometer. Differential cell counts were determined by cytocentrifuging cells onto slides and staining them with either Wright-Giemsa stain (Sigma) or Leukostat staining kit (Fisher Scientific, Pittsburgh, PA).

Quantitation of IL-4

IL-4 in the BAL fluid was quantitated by a sandwich ELISA. The buffers, diluents, and washes used between steps in this assay were according to the PharMingen Cytokine ELISA Protocol (PharMingen, San Diego, CA). Briefly, 96-well plates (Maxisorp, Nunc, Roskilde, Denmark) were coated overnight with the capture Ab 11B11 (National Cancer Institute, Biological Response Modifier Program) at 4°C and then blocked for 1 h with 1% BSA in PBS. Recombinant IL-4 standard (a gift from Dr. J. Ohara, Denver, CO.) and BAL fluid samples were added, and the plates were incubated for 2 h at room temperature. The bound cytokine was detected with a biotinylated secondary Ab (PharMingen), with an incubation period of 2 h at room temperature, followed by HRP-avidin (Becton Dickinson, Mountain View, CA), at a dilution of 1:3,000, with an incubation period of 30 min at room temperature. The plates were developed by addition of the substrate, o-phenylenediamine dihydrochloride (OPD), followed by incubation in the dark on a Variac shaker for 30 min before reading them in an ELISA plate reader (Spectramax 250, Molecular Devices, Sunnyvale, CA) at 490 nm. The calculations were done with the computer program provided with the plate reader.

Quantity of IgE

Total IgE in the BAL fluid was quantitated with a sandwich ELISA protocol. Briefly, the plates were coated with affinity-purified rabbit anti-IgE overnight at 4°C and then blocked with 1% BSA in PBS for 1 h at 37°C. The BAL fluid samples and appropriate dilutions of a standard IgE preparation were placed in the wells, and the plates were incubated for 3 h at 4°C. Sample blank wells did not receive BAL fluid but were otherwise treated similarly. The bound IgE was detected with polyclonal goat anti-IgE Abs (incubation for 1 h at 37°C), followed by HRP-conjugated rabbit anti-goat Abs (incubation for 1 h at 37°C). The plates were developed by addition of OPD and read in an ELISA plate reader at 490 nm.

An initial experiment, due to nonavailability of anti-OVA specific IgE Ab, the actual concentration (nanograms per milliliter) of OVA-specific IgE in BAL fluid was determined indirectly by batch absorption of 100 μl BAL fluid with either BSA-Sepharose (control) or OVA-Sepharose (test). The unabsorbed IgE level was determined by the total IgE ELISA as described above. The percent OVA-specific IgE present in BAL fluid was then calculated as a reciprocal value of (IgE not absorbed by OVA-Sepharose/IgE not absorbed by BSA-Sepharose) X 100.

This pooled BAL fluid contained 30 ng/ml OVA-specific IgE and was subsequently used as the standard in ELISA to determine OVA-specific IgE levels in BAL fluids from other experiments. For this, 96-well plates were coated with 10 μg/ml OVA overnight at 4°C and the plates were then blocked as for total IgE. The standard and experimental BAL fluids were placed at several dilutions and incubated for 2 h at room temperature. Then, plates were washed nine times in PBS, and 100 μl biotinylated anti-IgE (PharMingen) at 1 μg/ml were placed in each well. After 2 h of incubation at room temperature, the plates were washed as before and incubated with avidin-HRP conjugate for 30 min. After washing, the plates received the OPD substrate and were read 30 min later as described above for total IgE.

Quantitation of IgE immune complexes

IgE-OVA complexes in the BAL fluid were detected by a sandwich ELISA. The plates were coated with rabbit anti-OVA Abs overnight at 4°C and then blocked as described above. Samples were placed in the wells, the plates were incubated for 3 h at 4°C. The bound IgE was detected by goat anti-mouse IgE Abs followed by HRP-coupled rabbit anti-goat Abs (Zymed, San Francisco, CA) and substrate as described above. Experimental values were read off a standard curve prepared with serial dilutions of in vitro prepared anti-DNP IgE-TNP-OVA complexes (see Reagents).

Statistics

Data were analyzed using Statview software. An unpaired Student two-tailed t test was used to compare samples from different treatments as shown under individual experiments in Results.

Results

Ag-specific IgE is present in BAL fluid from mice with allergic airway inflammation

As shown in Fig. 1, IgE was detectable in BAL fluid from mice exposed to just one dose of aerosolized OVA, when the fluid was obtained 24 h after the Ag exposure. Significantly higher levels were detected after six Ag exposures, at both 3 and 24 h after the last exposure. Control mice that were sensitized to OVA but were exposed to aerosolized PBS did not show detectable levels of IgE. The levels of IgE in general correlated with eosinophilia. To determine whether the IgE present in the BAL fluid was Ag specific, samples of the BAL fluid taken 24 h after the last aerosol were adsorbed with OVA-Sepharose 4B, or BSA-Sepharose 4B as a control, and the concentrations of IgE in the supernatants were determined by ELISA. By this method, it was determined that >50% of the IgE secreted into the airways was Ag specific (Table I).

IgE persists in airway secretions after aerosolized Ag exposure

To determine whether IgE persists in airway secretions after cessation of Ag exposure, we treated OVA-sensitized mice with six doses of aerosolized OVA and then monitored the amounts of IgE in BAL fluid after various intervals following the last Ag challenge. Data are presented for two experiments with different and overlapping time intervals (Fig. 2). Both the total and ova-specific
IgE peaked at 3 days after the end of the last Ag exposure; the levels declined afterward, but significant amounts could still be detected 15 days after the challenge. In general, OVA-specific IgE was 50% of total IgE in the airways. Three days after the last aerosol, OVA-specific IgE reached peak levels and accounted for 50–90% of the total IgE. In contrast, although significant numbers of eosinophils could be detected in BAL fluid obtained from mice 1–3 days after the last aerosolized Ag challenge, the numbers declined precipitously afterward and reached a background level 9 days after the last Ag exposure.

IgE-OVA immune complexes are naturally formed in the airways in mice challenged with nebulized OVA. The Ag-specific IgE present in the airway secretions likely recognizes the same Ag to which the animals are exposed, thus forming immune complexes. To definitively establish the presence of such immune complexes, we challenged the OVA-immunized mice with nebulized OVA six times, obtained BAL fluids at various time points after the last challenge, and quantitated the amounts of the IgE immune complexes in the fluids. As shown in Fig. 3, significant levels of IgE-OVA complexes could be detected in BAL fluid harvested immediately after the Ag challenge. The levels in fluids obtained at later time points declined progressively, but the immune complexes remained readily detectable in the fluid obtained 30 min after the Ag challenge.

Administered IgE immune complexes are more potent than the Ag alone in inducing airway inflammation

Ag-Ab complexes are better recognized by lymphoid cells than Ag alone. We therefore decided to determine whether there is a difference in airway inflammation induced by Ag alone vs IgE-Ag immune complexes. To definitively establish the presence of such immune complexes, we challenged the OVA-immunized mice with nebulized OVA six times, obtained BAL fluids at various time points after the last challenge, and quantitated the amounts of the IgE immune complexes in the fluids. As shown in Fig. 3, significant levels of IgE-OVA complexes could be detected in BAL fluid harvested immediately after the Ag challenge. The levels in fluids obtained at later time points declined progressively, but the immune complexes remained readily detectable in the fluid obtained 30 min after the Ag challenge.

### Table I. OVA-specific IgE in BAL fluid of BALB/c mice

<table>
<thead>
<tr>
<th>No. Adsorbent (μl)</th>
<th>IgE (ng/ml) (unabsorbed)</th>
<th>% OVA-Specific IgE</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>BSA-Sepharose</td>
<td>OVA-Sepharose</td>
</tr>
<tr>
<td>1</td>
<td>12.5</td>
<td>56</td>
</tr>
<tr>
<td>2</td>
<td>25</td>
<td>58</td>
</tr>
<tr>
<td>3</td>
<td>50</td>
<td>56</td>
</tr>
</tbody>
</table>

*BSA and OVA were separately coupled to CNBr-activated Sepharose 4B (Pharmacia-Upjohn) at 2 ml protein (5 mg/ml) to 0.4 g Sepharose. The BAL fluid from OVA-sensitized and challenged mice from a representative experiment of three experiments was pooled. The IgE concentration in the BAL fluid was 60 ng/ml. Then, 100-μl aliquots of the BAL fluid were adsorbed with 12.5, 25, or 50 μl of Sepharose 4B beads coupled to Ag (OVA) or control (BSA). The unbound IgE in the supernatants was quantitated by ELISA. The percentage of OVA-specific IgE present in BAL fluid was calculated as a reciprocal of percent value of IgE not absorbed by OVA-Sepharose/IgE not absorbed by BSA-Sepharose.

### FIGURE 1
Detection of IgE and eosinophils in BAL fluid (BALF) from immunized mice with airway inflammation. BALB/c mice were immunized with OVA in alum and 14 days later were challenged with nebulized OVA 1, 3, or 6 times, 30 min each day. Control mice received aerosolized PBS challenge. The BAL fluid was collected either 3 h or 24 h after aerosol challenge, and the numbers of eosinophils and levels of IgE were determined as described in Materials and Methods. Data are presented as mean ± SEM from three mice for each data point, and results are representative of more than five experiments.

### FIGURE 2
Detection of IgE and eosinophils (EOS) in BAL fluid (BALF) from mice with airway inflammation over various intervals after termination of airway Ag exposure. BALB/c mice were sensitized with OVA and then challenged with nebulized OVA six times. The BAL fluid was collected from mice 1, 2, 3, and 7 (Experiment 1) and 3, 6, 9, and 15 (Experiment 2) days after the last challenge. The numbers of eosinophils and levels of total and OVA-specific IgE were determined as described in Materials and Methods. Data are presented as mean ± SEM from three mice for each data point.

### FIGURE 3
Detection of IgE-OVA immune complexes in BAL fluid from mice challenged with nebulized OVA. OVA-immunized BALB/c mice were challenged with nebulized OVA six times as described in Fig. 1. BAL fluids were obtained immediately (0 min), 30 min, or 24 h after the last challenge. The amounts of the IgE-OVA complexes were determined as described in Materials and Methods and are expressed as the quantity (nanograms per milliliter) of IgE in the complexes. Data are presented as mean ± SEM. from three mice.
complexes. For this purpose, we used TNP-OVA as the immunogen, which can form immune complexes with a previously well-characterized mouse monoclonal anti-DNP IgE Ab. To conserve the amount of IgE to be used, we administered the Ag or the immune complexes intranasally, rather than giving them in an aerosolized form. First, we established that OVA-alum-immunized mice developed similar levels of airway inflammation (cellular infiltration and IL-4 levels) on intranasal challenge with either OVA or TNP-OVA (data not shown). We then compared airway inflammation induced by intranasally administered TNP-OVA and IgE-TNP-OVA, in mice previously immunized with OVA. As shown in Fig. 4, the BAL fluid from mice challenged with IgE-TNP-OVA contained significantly higher numbers of eosinophils (Fig. 4A) and higher levels of IL-4 (Fig. 4C), as compared with that obtained from mice challenged with TNP-OVA alone. In addition, BAL fluid from mice challenged with IgE immune complexes contained significantly higher numbers of neutrophils as compared with those from mice challenged with the Ag alone (Fig. 4B). To ascertain that it was the presence of immune complexes rather than the mere addition of IgE that contributed to the augmented inflammatory response, we also challenged mice with TNP-OVA plus ragweed Ag-specific monoclonal IgE. As shown in Fig. 5, only TNP-OVA mixed with anti-TNP IgE, and not with anti-ragweed Ag IgE, induced heightened inflammatory responses.

IgE-Ag complexes and Ag produce comparable airway inflammatory responses in FcεRI-deficient mice

To better understand the mechanisms by which the IgE-Ag complexes affect heightened airway inflammatory responses over the Ag alone, we compared the responses induced by intranasal administration of TNP-OVA-IgE complexes vs TNP-OVA in OVA-sensitized FcεRI(−/−) mice. As a control, we used a mouse strain that is also deficient in the murine FcεRI-a chain but expresses the human FcεRI-a chain (mast cells in these mice contain functional FcεRI, that recognizes endogenous murine IgE). As shown in Fig. 6, the BAL fluid IL-4 levels in FcεRI(−/−) mice challenged with either IgE immune complexes or Ag alone were similar. In contrast, in Tg1101 mice, the immune complexes induced higher levels of IL-4 than Ag alone.

Discussion

The studies presented herein demonstrate that: 1) significant amounts of both free IgE as well as IgE immune complexes are

FIGURE 4. Comparison of airway inflammation induced by IgE immune complexes and Ag alone. OVA-immunized BALB/c were challenged with either 2 μg TNP-OVA alone or 20 μg anti-DNP IgE three times intranasally. The BAL fluid was collected 3 h after last challenge, and the numbers of leukocytes and levels of IL-4 were determined as described in Materials and Methods. Each data point represents mean ± SEM of data from three mice, and the results are representative of five separate experiments. The p values of TNP-OVA vs TNP-OVA-anti-DNP IgE in eosinophil (A), neutrophil (B), and IL-4 (C) determinations are all <0.001.

FIGURE 5. Comparison of airway inflammation induced by TNP-OVA in combination with anti-DNP IgE or anti-ragweed IgE. Mice immunized with OVA were challenged 14 days later intranasally with 2 or 4 μg TNP-OVA mixed with 20 μg anti-DNP IgE or TNP-OVA mixed with 20 μg anti-ragweed IgE. The results are mean ± SD of data from three mice. Similar results were obtained in two separate experiments.

FIGURE 6. Comparison of airway inflammation induced by TNP-OVA-IgE and TNP-OVA in FcεRI(−/−) mice. FcεRI(−/−) mice and Tg1101 mice, which express functional human FcεRI in the place of murine FcεRI, were immunized with OVA in alum. After 14 days, they were challenged with TNP-OVA alone or TNP-OVA-IgE immune complexes intranasally for 3 consecutive days. Three hours after the last challenge, BAL fluid was collected from each mouse, and IL-4 levels were determined by ELISA. The data are average values obtained from three mice ± SEM. Similar results were obtained in two independent experiments.
IgE immune complexes may be more effective than the Ag in mediating the production of cytokines and chemokines (32). Thus, IgE produced by B cells may become involved in forming more T cells (34). These events can lead to a domino effect where more associated lung tissue (BALT) areas of lung tissue independent of cells can in turn up-regulate IgE synthesis by B cells in bronchus-associated tissues and their production of various potent inflammatory mediators (35). IgE Ab results in an attenuated response to airway challenge, supporting a contributory role of IgE in airway inflammation in this mouse model (42). On the other hand, IgE-deficient mice generated by gene targeting showed eosinophilia and airway hyperresponsiveness after airway challenge that are comparable to the wild-type mice (43). It is possible that several pathways are involved in the overall response observed in the mouse model of allergic airway inflammation and that IgE is involved in one or more of these pathways. Genetically IgE-deficient mice might be able to mount a full allergic response comparable to that found in the wild-type mice through IgE-independent pathways only. Also relevant to the present discussion is the role of mast cells in allergic airway inflammation. Based on their localization in airway tissues and their production of various potent inflammatory mediators, mast cells are believed to play an important role in allergic airway inflammation (44). However, there are studies that show that mast cell-deficient mice develop airway inflammation comparable to their normal congenic mice upon airway Ag challenge (45), although other studies found that mast cell-deficient mice developed significantly reduced eosinophilia in the airways as compared with the normal congenic mice (46). Again, the results demonstrate the complexity and redundancy in the mechanisms.
References


