Migration of Antigen-Presenting B Cells from Peripheral to Mucosal Lymphoid Tissues May Induce Intestinal Antigen-Specific IgA Following Parenteral Immunization

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Parenterally administered immunizations have long been used to induce protection from mucosal pathogens such as *Bordetella pertussis* and influenza virus. We previously found that i.m. inoculation of mice with the intestinal pathogen, rotavirus, induced virus-specific Ab production by intestinal lymphocytes. We have now used adoptive transfer studies to identify the cell types responsible for the generation of virus-specific Ab production by gut-associated lymphoid tissue (GALT) after i.m. immunization. Three days after i.m. immunization with rotavirus, cells obtained from the draining peripheral lymph nodes of donor mice were transferred into naive recipient mice. We found that intestinal lymphocytes produced rotavirus-specific IgAs (IgM, IgA, and IgG) 2 wk after transfer of either unfractionated cells, or unfractionated cells rendered incapable of cellular division by mitomycin C treatment. Additional studies demonstrated that rotavirus-specific IgA, but not IgG, was produced by intestinal lymphocytes after transfer of purified B cells. Ig allotype analysis revealed that rotavirus-specific IgA was produced by intestinal B cells of recipient origin, suggesting that migration of Ag-presenting B cells from peripheral lymphoid tissues to GALT may contribute to the generation of mucosal IgA responses after parenteral immunization. Strategies that promote Ag uptake and presentation by B cells may enhance mucosal IgA production following parenteral immunization.

F or decades parenteral immunization has been used to stimulate protection from a variety of infectious organisms whose site of replication is limited solely to mucosal surfaces (e.g., influenza virus and *Bordetella pertussis*). Ag-specific Abs (1–4) and CTL (5–7) that protect against mucosal challenge have been detected at intestinal and respiratory mucosal surfaces after parenteral inoculation.

Little is known about the mechanisms by which mucosal Ab-secreting cells (ASC) (3) are induced by nonmucosal immunization. Studies by several groups have shown that Ag-specific IgA-secreting cells that bear the mucosal homing receptor αββ can be detected in the circulation shortly after parenteral inoculation (8, 9). These findings demonstrate that nonmucosal immunizations can induce effector B cells capable of homing to mucosal tissues. However, how and where these ASC are induced by nonmucosal immunization remain obscure. Understanding the mechanisms by which nonmucosal immunizations induce IgA-secreting cells that bear mucosal homing receptors may allow for rational design of vaccines administered by the parenteral route.

This work was supported in part by Grants 1 K08 AI01367 (S.E.C.) and 1 R01 AI26251 (P.A.O.) from the National Institutes of Health.

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2 This work was supported in part by Grants 1 K08 AI01367 (S.E.C.) and 1 R01 AI26251 (P.A.O.) from the National Institutes of Health.

3 Abbreviations used in this paper: ASC, Ab-secreting cells; GALT, gut-associated lymphoid tissues; ILN, inguinal lymph nodes; LP, lamina propria; mlg, membrane Ig; MLN, mesenteric lymph nodes; PP, Peyer’s patches; RT, room temperature.

Materials and Methods

**Mice**

Conventionally reared, 5- to 8-wk-old BALB/c (H-2d, Ighb), C.B-17 (H-2d, Ighb), or C57BL/6 (H-2d, Ighb) female mice (Taconic Breeding Laboratories, Germantown, NY), or C.B-17-SCID (H-2d) female mice (Wistar Institute, Philadelphia, PA) were housed in individual isolation units. Before inoculation, sera from these mice did not contain rotavirus-specific Abs as determined by ELISA.
Virus

Murine rotavirus strain EDIM was originally obtained from Richard Ward (Children’s Hospital of Cincinnati, Cincinnati, OH) and passed in infant mice as previously described (10).

Immunization of donor mice

Adult BALB/c mice were inoculated i.m. bilaterally in the quadriceps femoris muscle with 200 μl per hind leg of EDIM. Mice were inoculated with 2.4 × 10^7 50% shedding dose (equivalent to 18 ng of viral Ag).

Experimental design

Three days after i.m. inoculation of donor mice, draining inguinal lymph nodes (ILN) were harvested and disrupted. Unfractionated or FACS-purified populations of cells were transferred either i.v. (via tail vein infusion) or i.p. in a volume of 200–300 μl to seronegative, adult recipient mice. Two or 6 wk after adoptive transfer, recipient mice were sacrificed and intestinal and nonintestinal lymphoid cultures were established. Supernatants from lymphoid cultures were subsequently tested for the presence of rotavirus-specific Abs by ELISA.

Isolation of cells

ILN were harvested and then disrupted with 21-gauge needles and 200-μm wire mesh (Small Parts, Miami Lakes, FL) in IMDM (Life Technologies, Grand Island, NY) with 10% FBS (Life Technologies). Cells were passed through a 125-μm cell sieve (Thomas Scientific, Swedesboro NJ), washed three times, and resuspended in IMDM with 10% FBS.

Cell preparation and sorting

Treatment of cells with mitomycin C to block cellular division. Unfractionated ILN cells were diluted to a concentration of 5 × 10^7 cells/ml in PBS and incubated with 50 μg/ml mitomycin C (Sigma, St. Louis, MO) for 20 min at 37°C. Cells were washed three times in IMDM with 10% FBS. Purification of B cells. Unfractionated ILN cells were diluted to a concentration of 2 × 10^7 cells/ml and incubated for 30 min at room temperature (RT) with 4 μl/ml of mouse anti-mouse Thy1.2 (Biosource International, Camarillo, CA). Cells were washed with IMDM with 10% FBS, 0.3% (w/v) BSA (Sigma), and 20 mM HEPES buffer solution (Life Technologies) (lysis buffer). Cells were then resuspended in 900 μl lysis buffer and 100 μl Low-Tox rabbit complement (Accurate Chemical and Scientific, Westbury, NY) were added per 2 × 10^7 cells and incubated for 1 h at 37°C. Cells were placed on ice for 5 min, washed with cold lysis buffer, and resuspended in IMDM with 5% FBS. The remaining cells were incubated for 30 min at RT with PE-conjugated rat anti-mouse CD19 (PharMingen, San Diego, CA) at a concentration of 10 μg/ml. The cells were washed and resuspended in IMDM with 5% FBS. CD19^+ cells were purified by FACS using an EPICS Elite Flow Cytometer (Coulter, Hialeah, FL). This method yielded ≥99.9% CD19^+ cells.

Depletion of macrophages

Macrophages were stained by incubating unfractionated ILN cells with PE-conjugated rat anti-mouse CD11b (PharMingen) for 30 min at RT at a concentration of 2 μg/10^7 cells. Cells were washed and resuspended in IMDM with 5% FBS. Non-CD11b^+ cells were purified by FACS. This method yielded ≥99.8% non-CD11b^+ cells.

Depletion of dendritic cells

Dendritic cells were stained by incubating unfractionated ILN cells with biotin-conjugated mAb 33D1 (a kind gift of Dr. Ralph Steinman, Rockefeller University, New York, NY) (11) for 30 min at RT at a concentration of 2 μg/10^7 cells. The cells were washed and incubated with PE-conjugated streptavidin (PharMingen) for 15 min at RT at a concentration of 0.5 μg/10^7 cells. Cells were washed and resuspended in IMDM with 5% FBS. Non-33D1^+ cells were purified by FACS. This method yielded ≥99.9% non-33D1^+ cells.

Intestinal or nonintestinal lymphoid cultures

To assess the production of virus-specific Abs by intestinal or nonintestinal tissues of recipient mice, lymphoid cultures of ILN, PP, MLN, or small intestinal LP fragments were established 2 or 6 wk after adoptive transfer as previously described (12) with the following modifications. In brief, under sterile conditions ILN, PP, MLN, and small intestines were isolated. MLN, ILN, and PP were washed twice in IMDM containing 50 μg/ml of gentamicin (JRH Bioscience, Lenexa, KS). Segments of small intestine 5 cm in size were opened and washed twice in calcium- and magnesium-free HBSS (Life Technologies) containing 50 μg/ml of gentamicin and 25 mM HEPES (Mediatech, Washington, DC), once in HBSS with 0.05% EDTA to remove villous epithelial cells and intraepithelial lymphocytes, and twice in HBSS. Under a dissecting microscope (×30 magnification) fat, mesentry, and connective tissue were removed from small intestinal segments. Eight 1 × 1 mm LP fragments from small intestinal segments of each animal were dissected. One LP fragment, MLN, PP, or ILN was placed in a well of a 48-well plate (Costar Scientific, Braintree, MA) containing 0.5 ml of GALT media (Kennet’s HY media (Life Technologies), 100 μg/ml of streptomycin (JRH Bioscience), 50 μg/ml of gentamicin and 0.25 μg/ml of amphotericin B (Fungizone, JRH Bioscience)). Samples were incubated at 37°C in an atmosphere of 95% O2 and 5% CO2 for 5 days. Supernatant fluids were collected and tested for the presence of rotavirus-specific Igs (IgM, IgA, and IgG) by ELISA.

Detection of rotavirus-specific and total Igs by ELISA

Sera and supernatant fluids from intestinal and nonintestinal lymphoid cultures were tested for the presence of rotavirus-specific and total IgM, IgA, and IgG (13). Quantities of total IgM, IgA, and IgG were determined to assure the viability of each lymphoid culture. To determine quantities of rotavirus-specific Abs, individual wells of 96-well, flat-bottom plates (Costar) were coated with either 100 μl PBS (Life Technologies) or 200 ng purified rotavirus diluted in 100 μl of PBS. To determine quantities of total Abs, individual wells of 96-well, flat-bottom plates were coated with either 100 μl PBS or 100 μl goat anti-mouse IgM, IgA, or IgG (Cappel, West Chester, PA) diluted in 1:1,000 in PBS. Plates were incubated for 18 h at 4°C in a humid chamber. Wells were washed 5 times with PBS plus 0.05% Tween-20 (Sigma), blocked with 300 μl of 1% BSA plus 0.025% Tween-20 (BSA-T), and incubated for 1 h at RT. Wells were washed again as above and 50 μl of sera or supernatant fluids diluted in BSA-T were added. Plates were gently rocked for 2 h at RT and washed as above. Fifty microliters of HRP-conjugated goat anti-mouse IgM, IgA, or IgG (Southern Biotechnology Associates, Birmingham, AL) diluted 1:2,000 in BSA-T were added to each well. Plates were incubated for 1 h at RT and then washed as above. Fifty microliters of 0.04% tetramethylbenzidine peroxidase solution (Kirkegaard & Perry, Gaithersburg, MD) were added to each well. Washed plates were incubated with 0.1 OD units and 2-fold greater than OD values for the corresponding uncoated well. Supernatants of lymphoid cultures obtained from BALB/c or C.B-17 mice inoculated with rotavirus were used as controls. Quantities of virus-specific and total Igs were determined using an isotype-specific standard curve that was constructed for each assay based on serial dilutions of purified mouse IgM, IgA, or IgG (Sigma).

Detection of allotype-specific, rotavirus-specific IgA by ELISA

Quantities of rotavirus-specific IgA α or IgAβ were determined by ELISA using a modification of the technique described above. Following coating, blocking, and addition of sample as described above, 25 ng of biotinylated-mAb specific for mouse IgA α (HY16) or IgA β (HISM2) (14) diluted in 50 μl of BSA-T were added to each well and incubated for 1 h at RT. Wells were washed as above and 50 μl of streptavidin-HRP conjugate (PharMingen) diluted 1:2,000 in BSA-T was added. Plates were developed, and quantities of Abs were determined as described above.

Results

Adaptive transfer of unfractionated cells from draining lymph nodes of i.m.-immunized mice induces rotavirus-specific Ab production by GALT of recipient mice

To determine whether cells resident in draining peripheral lymph nodes after i.m. immunization with rotavirus were capable of inducing mucosal humoral immune responses, 2 × 10^7 unfractionated cells harvested from draining ILN were transferred into naive syngeneic mice. Repeated analysis by flow cytometry demonstrated that unfractionated cell populations contained ∼58–73%
To evaluate the role of APCs in the generation of intestinal humoral immunity following i.m. immunization, unfractionated ILN cells from recently i.m.-immunized BALB/c mice were treated with the radiomimetic agent, mitomycin C. Virus-specific IgA was produced by intestinal lymphocytes following adoptive transfer into naive BALB/c recipient mice (Table III). Similarly, no virus-specific IgA was produced by intestinal lymphoid tissues of SCID mice following adoptive transfer of 2 × 10^7 unfractionated cells harvested from the ILN of recently i.m.-immunized BALB/c mice (data not shown).

**Rotavirus-specific IgA, but not IgG, is produced by GALT lymphocytes after adoptive transfer of purified B cells**

To evaluate the production of rotavirus-specific IgM and IgA by intestinal lymphocytes is short-lived following adoptive transfer; conversely, production of virus-specific IgG persists for at least 6 wk

To determine the duration of virus-specific Ab production by GALT after adoptive transfer, we established lymphoid cultures 6 wk after transfer of 2 × 10^7 unfractionated cells into naive recipient mice. Six weeks after cell transfer, virus-specific IgG was still produced by intestinal lymphocytes, although at reduced quantities than observed at 2 wk (Table II). However, neither virus-specific IgM nor IgA were produced by LP, PP, or MLN 6 wk after transfer.

**Rotavirus-specific Abs are produced by GALT lymphocytes after adoptive transfer of APCs**

To verify that free rotavirus was not transferred with unfractionated cells, mice were inoculated with 200 μl of supernatant fluids obtained from the first, second, or third cell washings. Two weeks after inoculation, rotavirus-specific Abs were not detected in sera or supernatant fluids from either intestinal or nonintestinal lymphoid tissues (data not shown).

Production of rotavirus-specific IgM and IgA by intestinal lymphocytes is short-lived following adoptive transfer; conversely, production of virus-specific IgG persists for at least 6 wk

To determine the duration of virus-specific Ab production by GALT after adoptive transfer, we established lymphoid cultures 6 wk after transfer of 2 × 10^7 unfractionated cells into naive recipient mice. Six weeks after cell transfer, virus-specific IgG was still produced by intestinal lymphocytes, although at reduced quantities than observed at 2 wk (Table II). However, neither virus-specific IgM nor IgA were produced by LP, PP, or MLN 6 wk after transfer.

Table I. Intestinal and nonintestinal lymphoid tissues of formerly naive recipient mice produced rotavirus-specific Abs 2 wk after adoptive transfer of unfractionated cells obtained from draining peripheral lymph nodes of i.m.-immunized donor mice

<table>
<thead>
<tr>
<th>Site</th>
<th>IgM (ng/ml)</th>
<th>IgA (ng/ml)</th>
<th>IgG (ng/ml)</th>
</tr>
</thead>
<tbody>
<tr>
<td>PP</td>
<td>11.49 ± 1.59</td>
<td>24.53 ± 3.27</td>
<td>29.16 ± 2.92</td>
</tr>
<tr>
<td>MLN</td>
<td>18.22 ± 3.72</td>
<td>30.08 ± 6.21</td>
<td>39.22 ± 7.19</td>
</tr>
<tr>
<td>LP</td>
<td>&lt;0.50</td>
<td>8.09 ± 1.61</td>
<td>12.41 ± 2.70</td>
</tr>
<tr>
<td>ILN</td>
<td>2.60 ± 0.99</td>
<td>&lt;0.50</td>
<td>3.37 ± 1.33</td>
</tr>
</tbody>
</table>

*Draining peripheral lymph node cells were harvested from BALB/c mouse 3 days after i.m. immunization with murine rotavirus strain EDIM. A total of 2 × 10^7 unfractionated cells per mouse were transferred i.p. to seven naive, adult BALB/c mice. Two weeks after adoptive transfer, intestinal and nonintestinal lymphoid cultures from recipient mice were established. Supernatants were tested by ELISA for the presence of rotavirus-specific Abs. This table includes data from three consecutive experiments. Intravenous transfer of cells yielded similar results.

† Mean quantity of Ab detected (in ng/ml) ± SEM.

Table II. Intestinal and nonintestinal lymphoid tissues of formerly naive recipient mice produced rotavirus-specific IgG, but not IgA, 6 wk after adoptive transfer of unfractionated cells obtained from draining peripheral lymph nodes of i.m.-immunized donor mice

<table>
<thead>
<tr>
<th>Site</th>
<th>IgM (ng/ml)</th>
<th>IgA (ng/ml)</th>
<th>IgG (ng/ml)</th>
</tr>
</thead>
<tbody>
<tr>
<td>PP</td>
<td>&lt;0.50</td>
<td>&lt;0.50</td>
<td>1.49 ± 0.57</td>
</tr>
<tr>
<td>MLN</td>
<td>&lt;0.50</td>
<td>&lt;0.50</td>
<td>9.58 ± 4.17</td>
</tr>
<tr>
<td>LP</td>
<td>&lt;0.50</td>
<td>&lt;0.50</td>
<td>5.13 ± 1.74</td>
</tr>
<tr>
<td>ILN</td>
<td>&lt;0.50</td>
<td>&lt;0.50</td>
<td>3.06 ± 2.04</td>
</tr>
</tbody>
</table>

*Draining peripheral lymph node cells were harvested from donor BALB/c mice 3 days after i.m. immunization with murine rotavirus strain EDIM. A total of 2 × 10^7 unfractionated cells per mouse were transferred i.p. to four naive, adult BALB/c mice. Six weeks after adoptive transfer, intestinal and nonintestinal lymphoid cultures from recipient mice were established. Supernatants were tested by ELISA for the presence of rotavirus-specific Abs. Intravenous transfer of cells yielded similar results.

‡ Mean quantity of Ab detected (in ng/ml) ± SEM.

Table III. Virus-specific Abs were produced by intestinal lymphoid tissues after adoptive transfer of nondividing, unfractionated cells obtained from draining peripheral lymph nodes of i.m.-immunized donor mice

<table>
<thead>
<tr>
<th>Site</th>
<th>IgM (ng/ml)</th>
<th>IgA (ng/ml)</th>
<th>IgG (ng/ml)</th>
</tr>
</thead>
<tbody>
<tr>
<td>PP</td>
<td>2.72 ± 0.76</td>
<td>6.73 ± 2.89</td>
<td>2.10 ± 0.82</td>
</tr>
<tr>
<td>MLN</td>
<td>8.82 ± 1.81</td>
<td>0.80 ± 0.71</td>
<td>17.21 ± 9.45</td>
</tr>
<tr>
<td>LP</td>
<td>0.50</td>
<td>12.31 ± 6.74</td>
<td>4.81 ± 1.51</td>
</tr>
<tr>
<td>ILN</td>
<td>4.05 ± 0.78</td>
<td>&lt;0.50</td>
<td>11.91 ± 8.48</td>
</tr>
</tbody>
</table>

*Draining peripheral lymph node cells were harvested from donor BALB/c mice 3 days after i.m. immunization with murine rotavirus strain EDIM. Cell division was blocked by incubating unfractionated cells with mitomycin C. A total of 2 × 10^7 cells per mouse were transferred i.p. to four naive, adult BALB/c mice. Two weeks after adoptive transfer, intestinal and nonintestinal lymphoid cultures from recipient mice were established. Supernatants were tested by ELISA for the presence of rotavirus-specific Abs. This experiment was performed twice with similar results.

‡ Mean quantity of Ab detected (in ng/ml) ± SEM.
Table IV. Virus-specific IgA, but not IgG, Abs were produced by intestinal lymphoid tissues 2 wk after adoptive transfer of purified B cells obtained from draining peripheral lymph nodes of i.m.-immunized donor mice*.

<table>
<thead>
<tr>
<th>Site</th>
<th>IgM</th>
<th>IgA</th>
<th>IgG</th>
</tr>
</thead>
<tbody>
<tr>
<td>PP</td>
<td>&lt;0.50&lt;sup&gt;a&lt;/sup&gt;</td>
<td>1.23 ± 0.59</td>
<td>&lt;0.50</td>
</tr>
<tr>
<td>MLN</td>
<td>&lt;0.50</td>
<td>4.02 ± 1.31</td>
<td>&lt;0.50</td>
</tr>
<tr>
<td>LP</td>
<td>&lt;0.50</td>
<td>1.84 ± 0.71</td>
<td>&lt;0.50</td>
</tr>
<tr>
<td>ILN</td>
<td>&lt;0.50</td>
<td>&lt;0.50</td>
<td>&lt;0.50</td>
</tr>
</tbody>
</table>

* Draining peripheral lymph node cells were harvested from donor BALB/c mice 3 days after i.m. immunization with murine rotavirus strain EDIM. Unfractionated cells were incubated with mouse anti-mouse Thy1.2 and rabbit complement. The remaining cells were stained with PE-conjugated rat anti-mouse CD19 and CD19<sup>+</sup> cells were purified by FACS (purity >99.9%). A total of 2 × 10<sup>6</sup> cells per mouse were transferred i.p. to three naive, adult BALB/c mice. Two weeks after adoptive transfer, intestinal and nonintestinal lymphoid cultures from recipient mice were established. Supernatants were tested by ELISA for the presence of rotavirus-specific Abs. This table includes data from three consecutive experiments.

Table VI. Virus-specific IgA produced by intestinal tissues was of recipient, not donor, origin after adoptive transfer of congenic purified B cells obtained from draining peripheral lymph nodes of i.m.-immunized donor mice*.

<table>
<thead>
<tr>
<th>Site</th>
<th>IgA&lt;sup&gt;a&lt;/sup&gt;</th>
<th>IgA&lt;sup&gt;b&lt;/sup&gt;</th>
</tr>
</thead>
<tbody>
<tr>
<td>PP</td>
<td>&lt;0.50&lt;sup&gt;a&lt;/sup&gt;</td>
<td>0.61 ± 0.19</td>
</tr>
<tr>
<td>MLN</td>
<td>&lt;0.50</td>
<td>&lt;0.50</td>
</tr>
<tr>
<td>LP</td>
<td>&lt;0.50</td>
<td>1.20 ± 0.13</td>
</tr>
<tr>
<td>ILN</td>
<td>&lt;0.50</td>
<td>&lt;0.50</td>
</tr>
</tbody>
</table>

* Draining peripheral lymph node cells were harvested from donor BALB/c (IgA<sup>a</sup>) mice 3 days after i.m. immunization with murine rotavirus strain EDIM. Unfractionated cells were incubated with mouse anti-mouse Thy1.2 and rabbit complement. The remaining cells were stained with PE-conjugated rat anti-mouse CD19 and CD19<sup>+</sup> cells were purified by FACS (purity >99.9%). A total of 4 × 10<sup>6</sup> cells per mouse were transferred i.p. into three naive C.B-17 (IgA<sup>b</sup>) mice. Two weeks after adoptive transfer, intestinal and nonintestinal lymphoid cultures from recipient mice were established. Supernatants were tested by ELISA for the presence of rotavirus-specific IgA<sup>a</sup> and IgA<sup>b</sup>. This table includes data from two consecutive experiments.

Migration of virus-containing macrophages and dendritic cells may lead to the generation of rotavirus-specific IgG-secreting cells in GALT

To determine the contribution of macrophages or dendritic cells to the generation of intestinal humoral immune responses after i.m. immunization, ILN cells from i.m.-immunized mice were depleted of either macrophages (CD11b-bearing cells) or dendritic cells (33D1-bearing cells) by FACS. A total of 1.7 × 10<sup>6</sup> non-CD11b- or 1.4 × 10<sup>6</sup> non-33D1-bearing cells were transferred into naive recipient mice, and 2 wk later lymphoid cultures were established. Virus-specific IgG was not detected after transfer of cells depleted of either macrophages (Table VII) or dendritic cells (Table VIII). However, small quantities of virus-specific IgA were produced by GALT of recipient mice despite depletion of either macrophages or dendritic cells from adoptively transferred cells.

Discussion

We found that adoptive transfer of cells from the draining peripheral lymph nodes of mice recently immunized i.m. with rotavirus induced virus-specific Ab production by intestinal lymphocytes of naive recipient mice. In addition, we demonstrated that populations of rotavirus-specific intestinal lymphocytes that produced virus-specific IgA were transferred into intestinal lymphoid tissues 2 wk after adoptive transfer of purified B cells obtained from draining peripheral lymph nodes of i.m.-immunized donor mice. This transfer of B cells from the draining peripheral lymph nodes of i.m.-immunized donor mice is limited to the production of virus-specific IgA, but not IgG, Abs. Two weeks after cell transfer, virus-specific IgA<sup>a</sup> (recipient phenotype), but not IgA<sup>b</sup> (donor phenotype), was produced by intestinal tissues after adoptive transfer of 2 × 10<sup>6</sup> purified B cells from i.m.-immunized BALB/c mice into naive BALB/c mice (Table V, columns 1 and 2). Similarly, virus-specific IgA<sup>b</sup> (recipient phenotype), but not IgA<sup>a</sup> (donor phenotype), was produced by intestinal tissues after adoptive transfer of 4 × 10<sup>6</sup> purified B cells from i.m.-immunized BALB/c mice into naive C.B-17 mice 3 days after i.m. immunization with murine rotavirus strain EDIM. Unfractionated cells were incubated with mouse anti-mouse Thy1.2 and rabbit complement. The remaining cells were stained with PE-conjugated rat anti-mouse CD19 and CD19<sup>+</sup> cells were purified by FACS (purity >99.9%). A total of 10<sup>7</sup> unfractionated cells per mouse were transferred i.p. to three naive C.B-17 (IgA<sup>b</sup>) mice. Two weeks after adoptive transfer, intestinal and nonintestinal lymphoid cultures from recipient mice were established. Supernatants were tested by ELISA for the presence of rotavirus-specific IgA<sup>a</sup> and IgA<sup>b</sup>. This table includes data from two consecutive experiments.

Table VII. Virus-specific IgG production by intestinal and nonintestinal lymphoid tissue was eliminated by adoptive transfer of draining lymph node cells depleted of macrophages obtained from i.m.-immunized donor mice*.

<table>
<thead>
<tr>
<th>Site</th>
<th>IgM</th>
<th>IgA</th>
<th>IgG</th>
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</thead>
<tbody>
<tr>
<td>PP</td>
<td>1.39 ± 1.05&lt;sup&gt;a&lt;/sup&gt;</td>
<td>0.80 ± 0.45</td>
<td>&lt;0.50</td>
</tr>
<tr>
<td>MLN</td>
<td>7.85 ± 4.30</td>
<td>&lt;0.50</td>
<td>&lt;0.50</td>
</tr>
<tr>
<td>LP</td>
<td>&lt;0.50</td>
<td>4.86 ± 0.65</td>
<td>&lt;0.50</td>
</tr>
<tr>
<td>ILN</td>
<td>2.70 ± 1.26</td>
<td>&lt;0.50</td>
<td>&lt;0.50</td>
</tr>
</tbody>
</table>

* Draining peripheral lymph node cells were harvested from donor BALB/c mice 3 days after i.m. immunization with murine rotavirus strain EDIM. Unfractionated cells were stained with PE-conjugated rat anti-mouse CD19. Non-CD11b-bearing cells were purified by FACS (purity >99.9%). A total of 1.7 × 10<sup>6</sup> cells per mouse were transferred i.p. to three naive, adult BALB/c mice. Two weeks after adoptive transfer, intestinal and nonintestinal lymphoid cultures from recipient mice were established. Supernatants were tested by ELISA for the presence of rotavirus-specific Abs.

<sup>a</sup> Mean quantity of Ab detected (in ng/ml) ± SEM.
Table VIII. Virus-specific IgG production by intestinal and nonintestinal lymphoid tissues was eliminated by adoptive transfer of draining lymph node cells depleted of dendritic cells obtained from i.m.-immunized donor mice

<table>
<thead>
<tr>
<th>Site</th>
<th>IgM (ng/ml)</th>
<th>IgA (ng/ml)</th>
<th>IgG (ng/ml)</th>
</tr>
</thead>
<tbody>
<tr>
<td>PP</td>
<td>0.75 ± 0.50</td>
<td>&lt;0.50</td>
<td>&lt;0.50</td>
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<tr>
<td>MLN</td>
<td>1.99 ± 1.41</td>
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<td>&lt;0.50</td>
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<td>LP</td>
<td>&lt;0.50</td>
<td>2.85 ± 0.66</td>
<td>&lt;0.50</td>
</tr>
<tr>
<td>ILN</td>
<td>2.95 ± 1.04</td>
<td>&lt;0.50</td>
<td>&lt;0.50</td>
</tr>
</tbody>
</table>

* Draining peripheral lymph node cells were harvested from donor BALB/c mice 3 days after i.m. immunization with murine rotavirus strain EDIM. Unfractionated cells were stained with biotinylated-rat anti-mouse 33D1. Non-33D1-bearing cells were purified by FACSort (purity > 99.9%). A total of 1.4 × 10^7 cells per mouse were transferred i.p. to two naive, adult BALB/c mice. Two weeks after adoptive transfer, intestinal and nonintestinal lymphoid cultures from recipient mice were obtained. Supernatants were tested by ELISA for the presence of rotavirus-specific Abs.

* Mean quantity of Ab detected (in ng/ml) ± SEM.

IgA, as compared with IgG, were generated by different classes of APCs and had different life spans. Following i.m. immunization with murine rotavirus, B cells derived from peripheral lymph nodes functioned as APCs and stimulated a short-lived virus-specific IgA response in GALT. Conversely, either macrophages or dendritic cells or both appeared to induce a longer-lived intestinal IgG response.

The quantities of virus-specific IgGs produced by GALT following transfer of purified cell populations were less than those produced following transfer of unfractionated cells. Likewise, smaller quantities of virus-specific Abs were produced following adoptive transfer of mitomycin C-treated, as compared to untreated, cells. These findings suggest that multiple cell types may be responsible for the generation of mucosal virus-specific IgA following i.m. immunization. Although we have shown that adoptively transferred B cells harvested from donor lymph nodes three days after i.m. immunization can function as APCs and induce intestinal IgA production, the time of cell harvest may have biased against identification of activated B cells that were also capable of inducing mucosal IgA responses. We are currently exploring the contributions of donor-derived CD4+ T cells and activated B cells in this system.

Using IgA-allotype-specific reagents we found that intestinal rotavirus-specific IgA responses were induced by Ag-presenting B cells present in adoptively transferred cells. Following adoptive transfer of congenic purified B cells, virus-specific IgA produced by intestinal tissues was of the recipient, not donor, phenotype. In addition, adoptive transfer of MHC-incompatible B cells failed to induce production of virus-specific Abs, indicating that rotavirus does not bind nonspecifically to murine B cells. In this system, B cells, resident in draining peripheral lymph nodes 3 days after primary i.m. immunization, appeared to be capable of virus uptake, processing, and presentation to naive CD4+ T cells. To be effective APCs, B cells must meet three requirements: uptake of Ag, presentation of peptide in the context of MHC molecules, and expression of costimulatory molecules. Although the efficiency of Ag presentation by Ag-prime B cells has been described (16–20), naive B cells may also function as APCs (21). Naive B cells may take up Ag either specifically, by endocytosis of Ag bound to membrane Ig (mlg), or nonspecifically, by pinocytosis. Although in a naive animal the B cell precursor frequency for a given Ag may be presumed to be low, Milich et al (21) found that less than 3 × 10^3 activated, or 2 × 10^4 resting, splenic B cells were capable of activating hepatitis B core Ag-specific T cell hybridomas in the presence of hepatitis B core Ag. Low-affinity binding of multiva-
Ag (16, 33, 34), dose of the Ag (35), route of Ag administration (35), and number of CD4+ T cell divisions stimulated by antigenic exposure (36) have all been shown to influence the T cell cytokine repertoire. Second, the anatomic source of dendritic cells (37, 38) may influence the outcome of APC-T cell interactions. Schrader et al. (39) demonstrated that both mucosally and systemically derived dendritic cells induced IgA production in vitro, whereas Spaulding et al. (40) found IgA production was preferentially supported by PP as compared with splenic dendritic cells. Finally, differences in cytokine profiles may be influenced by the anatomic site where APC-CD4+ interactions occur (39).

We found that production of intestinal rotavirus-specific IgA, as compared with IgG, was short-lived following adoptive transfer of unfraccionated cells. Although controversial, persistence of Ag may be necessary to maintain Ag-specific, ASC populations (41, 42). Thus, the transient presence of intestinal virus-specific IgA-secreting B cells may reflect a relative inability of B cells displaying rotavirus peptide-MHC complexes to persist in intestinal lymphoid tissues as compared with either macrophages or dendritic cells. Although Ag-bearing dendritic cells have been shown to persist for over 100 days in germinal centers (43), the life span of Ag-presenting B cells is not known.

Pathogen-specific mucosal IgA plays a critical role in mucosal defense. Nonmucosal inoculation has previously been shown to induce mucosal IgA responses (11, 15, 44, 45). We have extended our understanding of how nonmucosal immunization induces mucosal immunity by demonstrating that intestinal IgA production can be induced by nonimmune, Ag-presenting B cells found in the draining peripheral node after i.m. immunization. However, the transient nature of Ag-specific, intestinal IgA induced by parental immunizations may explain a major shortcoming of these vaccines, namely the need for booster dosing to maintain mucosal protection. Strategies that increase the number or functional life span of Ag-presenting B cells may augment the magnitude and duration of mucosal IgA responses and thus enhance the efficacy of parenterally administered vaccines.

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