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Blockade of Chemokine Activity by a Soluble Chemokine Binding Protein from Vaccinia Virus

Antonio Alcamí,* Julian A. Symons,* Paul D. Collins,† Timothy J. Williams,† and Geoffrey L. Smith2*

Chemokines direct migration of immune cells into sites of inflammation and infection. Chemokine receptors are seven-transmembrane domain proteins that, in contrast to other cytokine receptors, cannot be easily engineered as soluble chemokine inhibitors. Poxviruses encode several soluble cytokine receptors to evade immune surveillance, providing new strategies for immune modulation. Here we show that vaccinia virus and other orthopoxviruses (cowpox and camelpox) express a secreted 35-kDa chemokine binding protein (vCKBP) with no sequence similarity to known cellular chemokine receptors. The vCKBP binds CC, but not CXC or C, chemokines with high affinity ($K_d = 0.1–15$ nM for different CC chemokines), blocks the interaction of chemokines with cellular receptors, and inhibits chemokine-induced elevation of intracellular calcium levels and cell migration in vitro, thus representing a soluble inhibitor that binds and sequesters chemokines. The potential of vCKBP as a therapeutic agent in vivo was illustrated in a guinea pig skin model by the blockade of eotaxin-induced eosinophil infiltration, a feature of allergic inflammatory reactions. Furthermore, vCKBP may enable the rational design of antagonists to neutralize pathogens that use chemokine receptors to initiate infection, such as HIV or the malarial parasite.


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3 Abbreviations used in this paper: MIP-1$\alpha$, macrophage inflammatory protein-1$\alpha$; GRO-\$\beta\$, growth-related oncogene \$\beta\$, CRK, chemokine receptor; VV, vaccinia virus; vCKBP, virus chemokine binding protein; MCP, monocyte chemoattractant protein; NAP-2, neutrophil activating peptide-2; LTB$_4$, leukotriene B$_4$; WR, Western Reserve; EDC, 1-ethyl-3-(3-dimethylaminopropyl)-carbodiimide; EGS, ethylene glycol-bis-succinamyl succinate.

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that the virus chemokine binding protein (vCKBP) binds CC chemokines with high affinity and inhibits their ability to induce signal transduction and cell migration in vitro. However, vCKBP does not bind or inhibit CXC or C chemokines. We demonstrate that the vCKBP inhibitory mechanism is the blockade of the interaction of chemokines with cellular receptors, thus representing the first soluble protein identified that sequesters chemokines and blocks their activity. Furthermore, the therapeutic application of this novel soluble chemokine inhibitor is illustrated in a guinea pig skin model by the ability of vCKBP to block eosinoid-induced eosinophil infiltration in vivo, a feature of allergic inflammatory reactions.

Materials and Methods

Viruses

The growth conditions of human U937 and TK143B cells, and the sources of VV strains have been described previously (20). The Evans and Lister strains used in this study and a Lister virus lacking the gene encoding the 35-kDa protein (Lister35K or V357) (21) were provided by A. H. Patel (Institute of Virology, Glasgow, U.K.).

Reagents

Radioiodinated recombinant human IFN-γ (90 μCi/μg) and RANTES, MIP-1α, IL-8, and GRO-α (2200 Ci/mmol) were obtained from DuPont-New England Nuclear (Boston, MA). Recombinant human [125I]MIP-1α (2000 Ci/mmol) used for determination of affinity constants was obtained from Amersham (Little Chalfont, U.K.). Recombinant human RANTES, MIP-1α, monocyte chemotactic protein-1 (MCP-1), MCP-2, MCP-4, eotaxin, 2137, 2-met-28-aminopropyl)-carbodiimide (EDC) or ethylene glycol-bis-succinamidyl peptide-2 (NAP-2), stromal cell-derived factor-1 (SDF-1), α- and β-integrin, IFN-γ-inducible protein-10, platelet factor 4, neutrophil-activating peptide-2 (NAP-2), stromal cell-derived factor-1α, and lymphoactin, mouse eosinotactin, and rat RANTES and MIP-1α were purchased from PeproTech (Rocky Hill, NJ). Recombinant mouse MIP-1α and human eosotactin, GRO-α, and F309 were obtained from R&D Systems (Minneapolis, MN). Recombinant human IFN-γ was purchased from Genzyme (Cambridge, MA). Recombinant B15R and AcB8R were purchased from CLONTECH (Millsboro, CA) (Reading, U.K.). The following were gifts: guinea pig eosinotactin from Drs. G. Andrews and H. J. Showell (Pfizer Central Research, Groton, CT), human IL-5 from Dr. T. N. C. Wells (Glaxo-Wellcome College, Geneva, Switzerland), and human C5a from Dr. J. Van Oostrum (Ciba Geigy, Summit, NJ). Heparin (6 kDa) and heparan sulfate from porcine or bovine intestinal mucosa, respectively, were purchased from Sigma Chemical Co. (St. Louis, MO). Rabbit antiserum to purified VV Lister 35-kDa protein, provided by A. H. Patel (Institute of Virology), or VV Western Reserve (WR) B15R protein expressed in baculovirus have been described previously (21, 22). Preparations of medium for binding and biologic assays

Supernatants from orthopoxvirus-infected TK143B cells or baculovirus-infected SF2 cells were harvested 1 or 3 days postinfection, respectively, and prepared as previously described (22). Baculovirus supernatants were concentrated and dialyzed against PBS as described previously (18). Supernatants were inactivated with 4,5-dimethylpyrazole before the hinge region of human IgG1, which was provided by pCOSFcink (see below). Samples were analyzed by SDSPAGE in 16% or 20% long gels with 12 or 14% acrylamide. In the competition assays with U937 cells, supernatants were preincubated with 100 μM [125I]chemokine in 100 μl at 1 h at 4°C. Subsequently, 2.5 × 106 U937 cells were added in 50 μl and incubated for 2 h at 4°C. Bound [125I]MIP-1α was determined by scintillation proximity assay in a beta counter. Nonspecific binding, determined in the presence of a 1000-fold excess of unlabeled MIP-1α or as binding to 200 pg of purified B8R-Fc, was subtracted and represented as 3.8 to 7.5% of the total counts per minute bound. Binding data were analyzed by the LIGAND program (26). The number of binding sites for MIP-1α per molecule of VV was calculated from the number of binding sites deduced from the Scatchard analysis and from the concentration and m.w. of recombinant 35K-Fc.

Construction of recombinant baculovirus

The 35-kDa gene was obtained by PCR using VV Lister DNA and oligonucleotides 5'-ATCGGTATCAATGAGAACAATATACGTCC-3', which inserts a KpnI site, and 5'-GTTGATCTCAGACACCGTGT GAGTTTG-3', containing a BamHI site. The 782-bp fragment was sequenced and cloned into pAcCL29-1 (27), generating pAc35K. The sequence was confirmed and contained the Arg148 variant of the Lister 35-kDa protein. The recombinant baculoviruses Ac35K and Ac35K.His were produced from pAc35K and pAc35K.His, respectively, as described previously (20). The 35K.His protein was purified by metal chelate affinity chromatography (NiNTA resin; Qiagen, Inc., Chatsworth, CA). The recombinant baculoviruses AcB15R and AeB8R have been described (20, 22).

Construction of 35K-Fc fusion and expression

The VV Lister 35-kDa gene fused to the Fc region of human IgG1 was constructed in pCOSEFCLINK (a gift from Dr. P. R. Young, SmithKline Beecham Pharmaceuticals, King of Prussia, PA). The insert was obtained by PCR using VV Lister DNA and the oligonucleotides 5'-TCAGAATTCATTATGAGAACAATATACGTCC-3', which introduces an EcoRI site, and 5'-ATCCCTCGAGACACCGTGTGAGTTTG-3', which inserts Oly and Thr and a KpnI site after amino acid 235 of the 35-kDa protein before the hinge region of human IgG1, which was provided by pCOSEFCLINK, generating pCox35K-Fc. The sequence was confirmed and contained the Arg148 variant of the Lister 35-kDa protein. The 35K-Fc protein was purified from supernatants of CHO cells stably transfected with pCOB35K-Fc on a protein A-Sepharose column (Pharmacia, Piscataway, NJ). The B8R-Fc fusion protein will be described elsewhere (J. A. Symons and G. L. Smith, unpublished).

Measurement of calcium mobilization in human eosinophils and neutrophils

Changes in intracellular calcium flux were determined in purified human neutrophils and eosinophils as described previously (28). Briefly, granulocytes were separated from mononuclear leukocytes, RBCs, and platelets using a combination of dextran sedimentation and density centrifugation over Percoll-plasma gradients. For neutrophil experiments, granulocytes comprised >95% neutrophils, and the remaining cells were a mixture of eosinophils and mononuclear cells. For eosinophil experiments, blood was obtained from healthy atopic volunteers, and eosinophils were purified (>98%) from the isolated granulocytes by immunomagnetic separation using anti-human CD16 microbeads (Midimacs, Miltenyl Biotec, Teddington, U.K.) (29). Purified cells were loaded with fura-2/AM (neutrophils, 1 μM; eosinophils, 2.5 μM) and, after two washes, resuspended at 106 cells/ml in Ca2+-free PBS containing 10 mM HEPES (pH 7.4), 10 mM glucose, and 0.25% BSA. Aliquots of cells were taken, and the extracellular calcium concentration was adjusted to 1 mM. Changes in fluorescence were monitored at 37°C using a spectrometer (LS50, Perkin-Elmer Corp., Norwalk, CT) at excitation wavelengths of 340 and 380 nm and an emission wavelength of 510 nm.

Chemotaxis assay

Cell migration was evaluated in 10-mm tissue culture inserts (Life Technologies). Cells were plated in the upper chamber, separated by a polycarbonate filter (8-μm pore size). After incubation at 37°C for 2 h, the filter was washed, fixed, and stained with 4',6-diamidino-2-phenylindole (DAPI) (30). The number...
of migrating cells in five high powered fields was counted using a fluorescence microscope.

**Bioassay of eosinophil accumulation in guinea pig skin**

Accumulation of endogenous peripheral blood eosinophils. Sedated guinea pigs were pretreated for 1 h with human IL-5 (18 pmol/kg; i.v. injection) to induce blood eosinophilia (31). Guinea pig eotaxin, human C5a, or LTB₄, with or without purified 35K. His protein, was then injected intradermally (50 μl/site) into individual sites in the shaved dorsal skin of the animals. After 2 h, the animals were killed, and the skin sites were excised (11 mm in diameter) for quantification of eosinophil accumulation by measurement of eosinophil peroxidase as previously described (31).
Accumulation of $^{111}$In-labeled eosinophils

Eosinophils were purified from the peritoneal cavity of horse serum-treated donor animals and radiolabeled with $^{111}$InCl$_3$ as previously described (28). $^{111}$In-labeled eosinophils ($5 \times 10^6$ cells/animal) were injected i.v. into recipient animals, and after 10 min, guinea pig eotaxin and human C5a, with or without purified 35K.His protein, were injected intradermally (100 $\mu$l/skin site; two sites per treatment) into individual sites in the shaved dorsal skin. After 2 h, the animals were killed, and the skin sites were excised (17 mm in diameter) for quantification of eosinophil accumulation by measurement of $^{111}$In counts using a gamma counter (28).

Results

VV, cowpox, and camelpox viruses encode soluble chemokine binding activity

Although no CKR homologues were found in the VV genome sequence (32), we searched for the expression of soluble chemokine binding proteins by orthopoxviruses. Binding assays with $[^{125}\text{I}]$RANTES were performed with media from cultures infected with 13 strains of VV, rabbitpox, or buffalopox viruses (considered VV strains), cowpox virus (Brighton Red strain), elephantpox virus (considered a cowpox virus strain), or camelpox virus, followed by chemical cross-linking with EDC. A $[^{125}\text{I}]$RANTES-vCKBP complex was observed with several viruses, including VV Evans (Fig. 1c), but not with the best characterized VV strains WR and Copenhagen (Fig. 1a). The size of the ligand-vCKBP complex ranged from 33 to 37 kDa, suggesting a vCKBP size of 25 to 29 kDa after subtraction of the 8-kDa monomeric RANTES. The variation in vCKBP size may reflect different polypeptide lengths or degrees of glycosylation among viruses. The different size predicted for vCKBP from orthopoxviruses by Graham et al. (19) in similar experiments (41 kDa) might be due to the use of higher resolution gels in this study. The soluble vCKBP was expressed before (early) and after (late) viral DNA synthesis by VV rabbit-

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The VV Lister 35-kDa secreted protein encoded vCKBP

Although the predicted vCKBP size was 25 to 29 kDa, its expression profile in orthopoxviruses (Fig. 1, a and c) correlated with the expression of a 35-kDa major secretory protein in VV Lister, Evans, and rabbitpox, but not in WR, Tian-Tan, or Wyeth (21, 33). The 35-kDa gene is present in cowpox (34), but is truncated by a frameshift mutation within the signal peptide in VV Copenhagen, where it is called B29R or C23L (32), and in VV WR, where it produces a 7.5-kDa protein (21, 35). The expression of vCKBP before (early) and after (late) viral DNA synthesis (Fig. 1b) was also consistent with the transcriptional regulation of the 35-kDa gene from the strong early/late promoter p7.5, widely used in VV expression vectors (36).

The absence of RANTES-binding activity in supernatants from cultures infected with a VV Lister mutant lacking the 35-kDa gene (ListerΔ35K) (21) indicated that the 35-kDa protein is the vCKBP encoded by VV Lister (Fig. 2a). This was confirmed by showing specific binding of $[^{125}\text{I}]$RANTES to the 35-kDa protein produced from recombinant baculovirus (Ac35K) and when fused to the Fc region of human IgG1 (35K-Fc), but not with preimmune, anti-35K, or anti-B15R rabbit serum. Cross-linking of media from cultures infected with VV Lister or Ac35K in the presence of 5 $\mu$l of preimmune, anti-35K, or anti-B15R rabbit serum was negative or very weak, suggesting vCKBP specificity for CC, but not CXC, chemokines.

FIGURE 2. Identification of the VV Lister 35-kDa protein as the vCKBP. a, Cross-linking of $[^{125}\text{I}]$RANTES with EDC to supernatants from cultures infected with the indicated VVs, recombinant baculoviruses, or 20 ng of purified Fc fusion proteins in the absence or the presence of a 1000-fold excess of RANTES (RAN) or IFN-γ. b, Cross-linking of $[^{125}\text{I}]$RANTES with EDC to supernatants from cells infected with VV Lister or Ac35K in the presence of 5 $\mu$l of preimmune, anti-35K, or anti-B15R rabbit serum. c, Cross-linking of $[^{125}\text{I}]$IFN-γ with EDC to supernatants from cultures infected with the indicated viruses. The amount of supernatant used was equivalent to 2 $\times$ 10$^6$ cells (VV) or 4 $\times$ 10$^5$ cells (baculovirus). An autoradiograph of the SDS-PAGE analysis, with molecular masses in kilodaltons, is shown. The positions of RANTES (R), IFN-γ monomers (M), IFN-γ dimers (D), and ligand-receptor complexes (triangles) are indicated.
complex, as reported for the myxoma IFN-γ (20), or B8R-Fc (VV IFN-γR) (20), was demonstrated to bind a wide range of chemokines by interaction with their proteoglycan binding domain (19, 37). Here we demonstrate that VV expresses a 35-kDa vCKBP that does not bind IFN-γ and is not B8R by showing that 1) the binding of [125I]RANTES to natural (VV Lister) or recombinant (Ac35K and 35K-Fc) 35-kDa protein was inhibited by unlabeled RANTES, but not IFN-γ (Fig. 2a); 2) [125I]IFN-γ was cross-linked to the natural IFN-γR expressed from all three viruses, WR, Lister, and ListerΔ35K, or the recombinant IFN-γR (AcB8R), but not to the 35-kDa protein (Ac35K; Fig. 2c); 3) the molecular sizes of monomeric IFN-γ complexed to IFN-γR were 60 and 52 kDa for the natural and recombinant IFN-γ-R, respectively (20), which were higher than the 35-kDa size of the RANTES-vCKBP complex and could not be explained by the size difference between IFN-γ (17 kDa) and RANTES (8 kDa; Fig. 2, a and c); 4) the 35-kDa size of the RANTES-vCKBP complex was smaller than the 50-kDa size predicted for the RANTES-IFN-γR complex, as reported for the myxoma IFN-γR/MTT (37); and 5) the binding of [125I]IFN-γ to the natural (VV Lister) or recombinant (AcB8R) VV IFN-γR was inhibited by unlabeled IFN-γ but not by RANTES (Fig. 2c). It is also demonstrated that VV IFN-γR does not bind RANTES by the inability of RANTES to bind rIFN-γR (AcB8R and B8R-Fc) (Fig. 2a) and the lack of binding of RANTES to supernatants from cells infected with Lister/35K, which still express IFN-γ-Rs (Fig. 2, a and c). In addition, a higher molecular size of the soluble IFN-γR, known to be encoded by all the orthopoxviruses included in this study (20), complexed with [125I]RANTES, [125I]GRO-α, or [125I]IL-8 was not observed (Fig. 1, a and c, and data not shown), indicating that the IFN-γ-R from other strains of VV, cowpox, or camelpox viruses does not bind CC or CXC chemokines.

Binding specificity and affinity of vCKBP for chemokines

The binding specificity of vCKBP was further investigated in cross-linking experiments (Fig. 3). The vCKBP expressed from recombinant baculovirus, VV Lister, cowpox virus, or camelpox virus bound human [125I]RANTES, and this binding was effectively competed in a dose-dependent manner by all human CC chemokines tested, including eotaxin, a specific chemoattractant of eosinophils. In contrast, human CXC chemokines (not shown). These results demonstrate that vCKBP from VV, cowpox, and camelpox viruses bind CC, but not CXC, chemokines.

The affinity of vCKBP for CC chemokines was determined in binding assays of 35K-Fc to [125I]MIP-1α in the presence of increasing doses of unlabeled chemokine competitors (Fig. 4c). These data showed that vCKBP binds CC chemokines with different affinities: MIP-1α > eotaxin > RANTES > MCP-1 > I309.

35K-Fc was saturable and of high affinity: in two experiments Kd values of 103 ± 4 pM (Fig. 4a) and 128 ± 9 pM (not shown) were obtained. The high affinity interaction of vCKBP for CC chemokines (Kd = 115 pM) was similar or 100-fold higher than the affinity of chemokines for cellular CKRs (1, 4, 5). Scatchard analysis of the binding data predicted 4.2 binding sites for MIP-1α in the 35K-Fc protein in both experiments, suggesting 2.1 binding sites for chemokines in the 35-kDa protein. This was consistent with the binding of two chemokine molecules to vCKBP, as suggested by cross-linking studies (Fig. 1c). The interaction of [125I]MIP-1α with 16.5 pM 35K-Fc was inhibited by a similar dose of purified recombinant 35-kDa protein, which was produced in baculovirus and containing a C-terminal six-histidine tag (35K.His), suggesting that both proteins bind MIP-1α with similar affinities (Fig. 4b).

The affinity of vCKBP for other chemokines was determined in binding assays of 35K-Fc to [125I]MIP-1α in the presence of increasing doses of unlabeled chemokine competitors (Fig. 4c). These data showed that vCKBP binds CC chemokines with different affinities: MIP-1α > eotaxin > RANTES > MCP-1 > I309.
The lower affinity for I309 probably reflects the amino acid sequence of this chemokine being the most divergent of the CC chemokine group (4). Competition experiments with other chemokines corroborated, in a different quantitative binding assay, the specificity of vCKBP for CC, but not CXC or C, chemokines and extended these studies to 19 chemokines (Fig. 4).

The vCKBP does not bind to the proteoglycan binding site of chemokines

Chemokines interact with glucosaminoglycans such as heparin or heparan sulfate through their carboxyl terminus (6, 9). This interaction is thought to facilitate chemokine localization to the endothelial cells and does not interfere with chemokines binding to their receptors. To ascertain whether vCKBP bound to the proteoglycan binding site of chemokines, [125I]MIP-1α was preincubated with various doses of heparin or heparan sulfate before incubation with the 35K-Fc protein. Figure 4e shows that heparin or heparan sulfate did not interfere with vCKBP binding to MIP-1α at doses up to 10 μg/ml and had a minor effect at higher doses up to 200 μg/ml, equivalent to a 6 × 10^5-fold molar excess of heparin over MIP-1α.

The vCKBP blocks binding of CC chemokines to cell surface receptors

Biologic activity of vCKBP for CC chemokines was shown by the ability of supernatants from orthopoxviruses and recombinant baculovirus Ac35K to inhibit the binding of radiiodinated RANTES and MIP-1α, but not GRO-α, to cellular receptors (Fig. 5a). This again demonstrated the specificity of vCKBP for CC chemokines. As previously reported (38, 39), excess unlabeled RANTES failed to block the binding of [125I]RANTES to cells (Fig. 5a). The binding of [125I]MIP-1α, but not [125I]GRO-α, to U937 cells was inhibited in a dose-dependent manner by supernatants containing the 35-kDa protein (Fig. 5b). The potent inhibitory activity of vCKBP was illustrated by the complete blockade of MIP-1α binding to cells in the presence of 0.1 to 1 μl of medium, equivalent to the amount of vCKBP synthesized by only 200 to 2000 cells. Furthermore, 10 or 50 pM purified recombinant 35K.His protein produced in baculovirus blocked the binding of 100 pM [125I]MIP-1α to U937 cells by 50 and 95%, respectively (Fig. 5c). These results showed that the soluble vCKBP blocks the binding of CC chemokines to their cellular receptors and were consistent with the high
affinity interaction of MIP-1α and other CC chemokines with recombinant 35K-Fc.

The vCKBP blocks the in vitro biologic activity of CC chemokines

The blockade of CC chemokine binding to cell receptors suggested that vCKBP would inhibit their biologic activity. This was demonstrated by the ability of vCKBP to inhibit the rapid and transient increase in intracellular calcium in eosinophils by the CC chemokines MCP-4 and eotaxin (Fig. 6a). This inhibitory effect was selective for these agonists, since vCKBP had no effect on the calcium response induced by C5a in eosinophils (not shown). In contrast, vCKBP had no effect on the elevation of intracellular calcium levels in human neutrophils in response to the CXC chemokines IL-8, GRO-α, and NAP-2 (Fig. 6b). This demonstrated specificity of vCKBP for CC, but not CXC, chemokines in a biologic assay.

The vCKBP from VV Lister or recombinant baculovirus Ac35K also blocked the migration of U937 cells in vitro in response to the CC chemokine MIP-1α (Fig. 6c). In contrast, ListerD35K and AcB8R supernatants did not.

Blockade of eotaxin chemotactic activity by vCKBP in vivo

The blockade of chemokine activity by vCKBP was demonstrated in vivo in a guinea pig skin model of eotaxin-induced eosinophil infiltration (31). Purified recombinant 35K.His bound guinea pig eosinophil peroxidase as an index of eosinophil numbers (Fig. 7a). The inhibitory effect of various doses of vCKBP on eosinophil accumulation in the skin in response to eotaxin was determined in guinea pigs injected i.v. with 111In-labeled eosinophils (Fig. 7b). The potency of this inhibitor in vivo was illustrated by the complete blockade of eotaxin activity by a threefold molar excess of vCKBP and 50% inhibition by an equal molar concentration (Fig. 7, a and b). This emphasizes the potential therapeutic application of vCKBP in inflammatory diseases.

Discussion

In this study we have identified and characterized a soluble vCKBP from poxviruses. We demonstrate that vCKBP is the VV Lister 35-kDa secreted protein, and similar proteins are expressed by other orthopoxviruses. This protein is encoded by gene B29R or C23L in the VV strain Copenhagen (32), where it is truncated and inactive, and is transcribed from the strong, early-late promoter p7.5, frequently used in VV expression vectors (36).

In contrast to known cellular CKRs, which are hydrophobic proteins with seven transmembrane domains, the 35-kDa protein is a cysteine-rich, acidic (pI 4.2), soluble protein with no sequence similarity to cellular counterparts. Many of the poxvirus-encoded cytokine receptors mimic the extracellular binding domain of known cellular counterparts; thus, vCKBP may represent a viral version of an unidentified cellular molecule, as has been proposed for the VV IFN-α/βR (18).

Chemokine binding proteins closely related to the VV Lister 35-kDa protein are found in other VV strains, the leporipoxviruses Shope fibroma virus (S-T1) and myxoma virus (M-T1), racoonpox
virus, cowpox virus, and camelpox virus (Ref. 19 and this report), and are predicted to be produced by strains of variola virus, the cause of smallpox (G3R) (40, 41). Interestingly, VV WR gene A41L is predicted to encode a secretory protein more distantly related to the VV Lister 35-kDa protein (33, 42) and may represent another vCKBP that binds chemokines not included in this study.

Concerning the specificity of vCKBP for different chemokines, we demonstrate unequivocally that it is a specific inhibitor of CC, but not CXC or C, chemokines. This is shown by cross-linking experiments with cold competitors, in more quantitative binding experiments, and by determination of affinity constants in studies in which 19 chemokines were analyzed. Most importantly, the CC specificity was confirmed in biologic assays showing that vCKBP does not block the interaction of CXC to cellular receptors or their

FIGURE 6. Blockade of chemokine biologic activity by vCKBP in vitro. a, Elevation of intracellular calcium levels in human eosinophils by CC chemokines. MCP-4 or eosinin was preincubated with the indicated doses of purified 35K.His or control supernatant for 5 min before its addition to human eosinophils and measurement of calcium mobilization. The maximum elevation of intracellular calcium (mean ± SEM) in three experiments with eosinophils from different donors is shown. Representative traces from one experiment are represented in the insets. b, Elevation of intracellular calcium levels in human neutrophils by CXC chemokines. IL-8, GRO-α, or NAP-2 was preincubated with purified 35K.His or control supernatant for 5 min before its addition to human neutrophils and measurement of calcium mobilization. The maximum elevation of intracellular calcium (mean ± SEM) in four experiments with neutrophils from different donors is shown. c, Chemotactic activity of MIP-1α on U937 cells. a MIP-1α alone, + Lister, + Lister+35K, + Ac35K, + AcB8R. b Number of Migrating Cells

FIGURE 7. Inhibition of eotaxin-induced eosinophil accumulation in guinea pig skin by vCKBP. a, Guinea pigs were pretreated for 1 h with human IL-5 to induce blood eosinophilia. Guinea pig eotaxin, human C5a, or LTB₄ was injected intradermally in the presence of purified 35K.His (30 pmol/skin site) or control medium, and eosinophil accumulation was determined after 2 h. Results represent the number of eosinophils per skin site ± SEM from four or five animals. A statistically significant difference between groups that did or did not receive purified vCKBP is indicated by an asterisk (p < 0.02). The detection limit of the assay is indicated with a dashed line. No eosinophil accumulation was detected in response to the injection vehicle (HBSS containing 0.1% low endotoxin BSA), purified 35K.His, or control medium alone. b, ¹¹¹In-labeled eosinophils were injected i.v. into guinea pigs, and after 10 min, guinea pig eotaxin or human C5a, with or without 35K.His, was injected intradermally into individual sites. Accumulation of ¹¹¹In-labeled eosinophils was determined after 2 h. Results represent the mean ± SEM from four animals. Eosinophil accumulation in response to the injection vehicle is indicated with a dashed line and was similar to that obtained with purified 35K.His or control medium alone.
capacity to transduce signals. This contradicts the conclusion of Graham et al. (19), who reported that it had a broad specificity for CC and CXC chemokines. However, consistent with our results, these researchers found that excess IL-8 could not compete the cross-linking of RANTES to vCKBP, but the effect on the biologic activity of CXC chemokines was not investigated.

The mechanism by which vCKBP inhibits chemokine activity might be its interaction with the proteoglycan binding domain of chemokines, which is thought to facilitate chemokine localization to endothelial cells, as suggested by Graham et al. (19). Alternatively, vCKBP might interact with the receptor binding domain of chemokines and thus block binding to cellular receptors, as reported for poxvirus soluble receptors (13, 14). The data in this paper show that vCKBP blocks CC chemokine activity by binding chemokines with high affinity, at a site different from the proteoglycan binding domain, and preventing their interaction with cellular receptors. Consistent with this, the affinity of vCKBP for human MIP-1α (K_d = 115 pM) is similar to 10- to 100-fold higher than the affinity of most chemokines for cellular CKRs, which normally ranges from 1 to 10 nM (1, 4, 5). The vCKBP also binds eotaxin, RANTES, and MCP-1 with high affinity (K_d = 1–15 nM). The lower affinity of RANTES for myxoma M-T1 protein reported by Graham et al. (19) (K_d = 73 nM) might be due to the use of human chemokines with the M-T1 protein from myxoma virus, whose natural host is the rabbit. There is precedent for myxoma virus-soluble cytokine receptors binding human cytokines with much lower affinity than rabbit cytokines (43, 44).

Evidence for the binding of vCKBP to two chemokine molecules was found in cross-linking experiments using EGS and in Scatchard analysis of the binding of MIP-1α to 35K-Fc. MIP-1α and IL-8 can dimerize, but there is controversy about binding to cellular CKRs and the biologic activity of chemokines as monomers or dimers (4). If vCKBP does bind to dimeric chemokines, this might enhance the inhibitory activity of vCKBP.

Poxviruses have evolved two different mechanisms to block chemokine activity. First, the soluble IFN-γR from myxoma virus (M-T7), but not VV strain WR, binds the heparin-binding domain of a wide range of chemokines (19, 37), and this additional functional of M-T7 is consistent with an increased leukocyte infiltration to sites of myxoma virus replication in rabbits infected with a deletion mutant lacking this protein (45). The myxoma M-T7 protein may prevent the correct localization of chemokines in vivo, which form a gradient by interacting with proteoglycans at the surface of endothelial cells. We show here that the soluble IFN-γR from 19 orthopoxviruses (20), including strains of VV, cowpox, and camelpox viruses, does not bind CC or CXC chemokines. Second, we show that orthopoxviruses encode a vCKBP that binds chemokines with high affinity at a site different from the proteoglycan binding domain and blocks their interaction with cellular CKRs.

The expression of a potent, soluble, CC-specific vCKBP by poxviruses suggests an important role for CC chemokines in anti-viral defense. Nonetheless, deletion of the 35-kDa protein in VV rabitpox did not greatly affect the outcome of infection in mice and rabbits (33). Graham et al. (19) now report an increased leukocyte influx in the skin of rabbits infected with VV rabitpox mutant lacking the 35-kDa gene, compared with wild-type VV rabbitpox. However, construction of a revertant virus in which the 35-kDa gene is reintroduced into the VV rabbitpox mutant genome is required to support this conclusion. This VV rabbitpox mutant was constructed by transfecting DNA from VV Lister into VV rabbitpox-infected cells (33), and other mutations might have been introduced in the regions flanking the 35-kDa gene, which includes a gene encoding a TNFR homologue (32). Additional studies with this or alternative models of poxvirus infection are required to fully understand the role of vCKBP in poxvirus pathogenesis.

The important role of chemokines and CKRs in the pathogenesis of virus infections is emphasized by the expression of 1) chemokine-like proteins by molluscum contagiosum virus (46), Kaposi’s sarcoma-associated herpes virus (47, 48), and murine CMV (49), representing potential alternative ways to modulate chemokine activity; and 2) seven-transmembrane domain CKRs by large DNA viruses such as herpesviruses and poxviruses (14). Moreover, the MIP-1α knockout mouse is less able to clear influenza virus infection (50), and RANTES, MIP-1α, and MIP-1β increase resistance to HIV infection (2, 3).

Poxvirus proteins that counteract the immune system have been optimized during the evolution of viruses with their hosts; are providing insights into the physiologic role of immune regulatory molecules such as cytokines (51) or chemokines; and represent potential sources of immunomodulatory proteins and new strategies of immune modulation. In this report a novel soluble chemokine inhibitor from poxviruses with therapeutic potential is described. Eosinophil accumulation in response to eotaxin is a feature of allergic inflammatory reactions, such as those occurring in allergic asthma. The potent eosinophil inhibitory effect of vCKBP in vivo emphasizes its potential therapeutic application in inflammatory diseases. The vCKBP has binding specificity for CC chemokines, but the soluble inhibitor or variants from other poxviruses may provide the structural scaffolding needed to design specific soluble inhibitors of other chemokines.

The finding of a soluble vCKBP also has implications for infectious agents such as HIV, in which CKRs play an important role in determining transmission and disease progression (2, 3). Inhibition of the interaction of the HIV envelope with the chemokine binding domain of CKRs has recently emerged as a new target for intervention. The vCKBP might be engineered to create soluble molecules that bind to the domain of gp120, which interacts with the CKR to block HIV infection at an early stage (2, 3). A similar approach might yield therapeutic agents to block attachment of the malarial parasite to the Duffy Ag on erythrocytes and initiation of the infection (52).

The poxvirus chemokine inhibitor reported here represents a new and exciting virus immune evasion mechanism that provides insights into virus pathogenesis, the function of key immune regulatory molecules, and new strategies for therapeutic intervention in immune responses and disease.

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References
