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Efficient Clearance of Early Apoptotic Cells by Human Macrophages Requires M2c Polarization and MerTK Induction

Gaetano Zizzo,*,† Brendan A. Hilliard,*,† Marc Monestier,†,‡ and Philip L. Cohen*,†,‡

Mer tyrosine kinase (MerTK) is a major macrophage apoptotic cell (AC) receptor. Its functional impairment promotes autoimmunity and atherosclerosis, whereas overexpression correlates with poor prognosis in cancer. However, little is known about mechanisms regulating MerTK expression, in humans. We found that MerTK expression is heterogeneous among macrophage subsets, being mostly restricted to anti-inflammatory M2c (CD14+CD163+CD204+CD206+CD209+) cells, differentiated by M-CSF or glucocorticoids. Small numbers of MerTK+ “M2c-like” cells are also detectable among circulating CD14+CD16+ monocytes. MerTK expression levels adapt to changing immunologic environment, being suppressed in M1 and M2a macrophages and in glucocorticoids. Small numbers of MerTK+ “M2c-like” cells are also detectable among circulating CD14brightCD16+ monocytes.

Induction of regulatory T cell–mediated induction of anti-inflammatory monocytes-macrophages. MerTK enables M2c macrophages to clear early ACs more efficiently than other macrophage subsets, and it mediates AC clearance by CD14+CD16+ monocytes. Moreover, M2c cells release Gas6, which in turn amplifies IL-10 secretion via MerTK. IL-10–dependent induction of the Gas6/MerTK pathway may, therefore, constitute a positive loop for M2c macrophage homeostasis and a critical checkpoint for maintenance of anti-inflammatory conditions. Our findings give new insight into human macrophage polarization and favor a central role for MerTK in regulation of macrophage functions. Eliciting M2c polarization can have therapeutic utility for diseases such as lupus, in which a defective AC clearance contributes to initiate and perpetuate the pathological process. The Journal of Immunology, 2012, 189: 000–000.

The prompt recognition and removal of dead and dying cells is critical for maintenance of immunologic tolerance and resolution of inflammation. Physiologic mechanisms of apoptotic cell (AC) clearance typically associate with induction of regulatory pathways in phagocytes and release of anti-inflammatory cytokines (1, 2). Such mechanisms have attracted increasing interest over the last decade, and many distinct molecular pathways have been identified. Although there seems to be conspicuous redundancy among these pathways, they differ from each other in regard to several features. These features include dependence on specific nuclear transduction factors (3–6), expression under basal conditions versus inducibility by excess numbers of ACs (5–9), recognition of unmodified versus modified phosphatidylserine on ACs, direct AC recognition versus use of bridging molecules, and, importantly, clearance of early versus late (secondarily necrotic) ACs (9, 10).

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Abbreviations used in this article: AC, apoptotic cell; DC, dendritic cell; MerTK, Mer tyrosine kinase; MFI, mean fluorescence intensity; Treg, regulatory T cell.

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detrimental in cancer (26), whereby the ligand Gas6, secreted by tumor-associated macrophages, promotes tumor growth and metastasis (27).

Therefore, modulating MerTK activity could be a promising therapeutic approach to various pathologic conditions. Surprisingly, little is known about the mechanisms promoting MerTK expression in humans. Both MerTK and protein S can be upregulated by steroids (28–30), and protein S/MerTK-mediated AC clearance is enhanced by these drugs (12). However, steroids induce many other molecules involved in AC clearance (29, 30). It is, therefore, unclear whether MerTK upregulation by glucocorticoids is due to intrinsic and unique properties of these drugs or to steroid-induced apoptosis, which in turn promotes LXRs activation, or to other unidentified mechanisms.

Our study examined how the immunologic microenvironment affects MerTK expression and aimed to identify the human macrophage subsets in which MerTK is clearly expressed and functionally relevant. Macrophage subsets include: M1, secreting IL-12 and promoting Th1 differentiation; M2a, secreting IL-4 and related to Th2 polarization; M2b and M2c, both secreting IL-10 and associated with regulatory T cell (Treg) expansion (31). We found that, in the presence of IL-10, M-CSF differentiates macrophages expressing high levels of MerTK; such macrophages are characterized by an M2c phenotype. Glucocorticoids have analogous effects, with MerTK upregulation occurring as a consequence of steroid-induced M2c polarization; however, this process is IL-10 independent. Remarkably, neither M-CSF alone nor IL-10 alone is able to drive full M2c differentiation and upregulate MerTK significantly. MerTK is also expressed among M2c-like CD14<sup>high</sup>CD16<sup>dim</sup> circulating monocytes and in minor populations of CD14<sup>high</sup>CD16<sup>dim</sup>CD163<sup>+</sup> cells differentiated among M1 and M2a macrophages. MerTK makes M2c (M-CSF plus IL-10) macrophages and M2c-like cells highly capable of clearing ACs, and it significantly enhances IL-10 secretion by M-CSF–driven macrophages following Gas6 ligation. Our data support a broad role for MerTK in M2c macropage homeostasis and highlight the potential usefulness of polarizing human macrophages with both M-CSF and IL-10 to ensure an optimal, MerTK-mediated clearance of ACs.

Materials and Methods

Cell cultures

Human macrophages from buffy coats of healthy blood donors were isolated by Ficoll-Paque Plus gradient (GE Healthcare) and magnetic separation, using a kit for human monocyte enrichment by negative selection (EasySep, StemCell), according to the manufacturers’ instructions. Purity of CD14<sup>+</sup> cells was 89–94%. CD14<sup>+</sup> cells were cultured in 24-well plates at 0.8 × 10<sup>5</sup> cells/ml for 7–8 d at 37°C in 5% CO<sub>2</sub> in complete RPMI 1640 medium containing 10% human AB serum, 1-t-glutamine, penicillin, and streptomycin. To prevent formation of clumps induced by autologous serum and experiments in serum-free conditions, cells were cultured at 0.8 × 10<sup>5</sup> cells/ml for 7–8 d at 37°C in 5% CO<sub>2</sub> in complete RPMI 1640 medium supplemented with 10% heat-inactivated FBS, 10% NaCl, 1.0% Triton X-100, and freshly added cocktails of n-pentanol and phosphatase inhibitors (Sigma–Aldrich) (Sigma–Aldrich) or coated onto a 5% polyacrylamide gel. Proteins, transferred to PVDF membranes (Millipore), were probed with biotinylated goat polyclonal anti-human MerTK (R&D Systems) and rabbit anti–β-actin Abs (Santa Cruz Biotechnology), followed by HRP-conjugated streptavidin (BioLegend) and secondary goat anti-rabbit Ab (Santa Cruz Biotechnology), respectively. Immunoblots were developed and visualized by ECL using Amer sham ECL reagents (GE Healthcare). Densitometry of bands, normalized to β-actin expression, was calculated using Image J software.

Induction of apoptosis and phagocytosis assays

Human neutrophils were isolated from Ficoll-Hypaque pellets through dextran erythrocyte sedimentation and lysis of contaminating erythrocytes by incubation with ice-cold ammonium chloride (0.15 M) and potassium bicarbonate (0.01 M) solution. Neutrophils were resuspended at 1 × 10<sup>6</sup> cell/ml in 10% FBS–RPMI, labeled with 2.5 μM CFSE (CFSE–RPMI) (Sigma–Aldrich), and incubated for 20 h at 37°C in 5% CO<sub>2</sub>. The composition of neutrophils routinely obtained after incubation, according to annexin V and propidium iodide (PI) staining, was 66.0 ± 10.2% early ACs (annexin V<sup>+</sup> PI<sup>+</sup>), 3.8 ± 2.2% late ACs (annexin V<sup>+</sup> PI<sup>-</sup>), 0.3 ± 0.2% necrotic cells (annexin V<sup>-</sup> PI<sup>-</sup>). Apoptotic neutrophils were added for 60 min to cultured monocyte/macrophages at a 5:1 ratio. Flow cytometry was used to quantify percentage of CD14<sup>+</sup> circulating monocytes that phagocytosed CFSE-labeled ACs, and to calculate phagocytosis index. In some experiments, as specified in the text, phagocytosis activity was assessed separately on CD14<sup>dim</sup>CD163<sup>+</sup> and CD14<sup>high</sup>CD163<sup>dim</sup> macrophage subsets.

For phagocytosis assays on circulating monocytes, PBMCs were mixed with apoptotic neutrophils at a 1:1 ratio, and incubated in BDF Falcon tubes for 4 h at 37°C in 5% CO<sub>2</sub>. Phagocytosis activity was measured on CD14<sup>+</sup>CD163<sup>+</sup>CD14<sup>high</sup>CD16<sup>+</sup>CD16<sup>high</sup>CD163<sup>+</sup> monocyte subsets by flow cytometry.

For inhibition studies, macrophages and circulating monocytes were preincubated with a goat anti-human MerTK Ab (R&D Systems) or goat control IgG (Southern Biotech) for 30 min before adding apoptotic neutrophils. The effects of MerTK block during phagocytosis assay was additionally observed using a Leica TCS SP5 confocal laser scanning microscope, 40×/1.25 numerical aperture (NA) oil objective, labeled apoptotic neutrophils with Hoechst 33342 (0.5 μg/ml; Invitrogen) and staining macrophages with mouse anti–CD14 (BioLegend) and goat biotinylated anti–CD63 (R&D Systems) Abs, followed by secondary APC-conjugated goat anti-mouse Ab and FITC-conjugated streptavidin (BioLegend), respectively.

Gas6, IL-10, and TNF-α detection by ELISA

Gas6 levels were measured in supernatants of cell cultures treated with various cytokines for 4 d in serum-free conditions, using sandwich ELISA according to standard procedure (23). Standard curves were prepared with rhGas6 (R&D Systems). Purified goat anti-human Gas6 Ab (R&D Systems) was used for capture. Biotinylated goat polyclonal anti-human Gas6 Ab (R&D Systems), followed by HRP-conjugated streptavidin (BioLegend), was used for detection.

Secretion of IL-10 and TNF-α was induced by LPS (50 ng/ml; Sigma–Aldrich) or recombinant MerTK (R&D Systems) for 24 h in cultured cells after washing in buffer containing 2% BSA. The following mouse mAbs were used: anti-CD14 (PE-Cy7), anti–HLA-DR (APC) (anti–CD163 (APC or PerCP-Cy5.5), anti-CD206 (APC-Cy7), anti–CD209 (PerCP-Cy5.5), anti–CD1a (APC), anti–CD210 (PE; BioLegend); anti–CD16 (APC-Cy7; BD Biosciences); anti–CD204 (APC), and anti–MerTK (clone 125518; PE; R&D Systems). MerTK expression was evaluated using appropriate PE-labeled isotype control (BioLegend). Cells were analyzed using FACSCalibur (BD Biosciences) and FlowJo software.

Detection of MerTK expression by Western blot

Cell lysates were obtained in buffer containing 50 mM HEPES, 150 mM NaCl, 10% glycerol, 1% Triton X-100, and freshly added cocktails of n-pentanol and phosphatase inhibitors (Sigma–Aldrich) (Sigma–Aldrich) or coated onto a 5% polyacrylamide gel. Proteins, transferred to PVDF membranes (Millipore), were probed with biotinylated goat polyclonal anti-human MerTK (R&D Systems) and rabbit anti–β-actin Abs (Santa Cruz Biotechnology), followed by HRP-conjugated streptavidin (BioLegend) and secondary goat anti-rabbit Ab (Santa Cruz Biotechnology), respectively. Immunoblots were developed and visualized by ECL using Amer sham ECL reagents (GE Healthcare). Densitometry of bands, normalized to β-actin expression, was calculated using Image J software.

Analysis of cell-surface molecules by FACS

Phenotype analysis by flow cytometry was performed in freshly isolated and in cultured cells after washing in buffer containing 2% BSA. The following mouse mAbs were used: anti–CD14 (PE-Cy7), anti–HLA-DR (APC) (anti–CD163 (APC or PerCP-Cy5.5), anti-CD206 (APC-Cy7), anti–CD209 (PerCP-Cy5.5), anti–CD1a (APC), anti–CD210 (PE; BioLegend); anti–CD16 (APC-Cy7; BD Biosciences); anti–CD204 (APC), and anti–MerTK (clone 125518; PE; R&D Systems). MerTK expression was evaluated using appropriate PE-labeled isotype control (BioLegend). Cells were analyzed using FACSCalibur (BD Biosciences) and FlowJo software.
a goat anti-human Gas6 Ab (5 μg/ml; R&D Systems) to block Gas6 activity prior to LPS stimulation. Potential variations in TNF-α levels were assessed by ELISA.

Statistical analysis

Data are expressed as mean ± SEM. Statistical significance among different cell treatments was assessed with Student paired t test, or one-way repeated measures ANOVA if treatment groups were more than two. Statistical significance was defined as p < 0.05. Analysis and graphing were performed using GraphPad Prism software.

Results

MerTK is upregulated during monocyte-to-macrophage differentiation and is further enhanced by M-CSF

Healthy monocytes were sorted from PBMCs through negative selection using magnetic beads and analyzed for MerTK expression at days 0, 1, and 3 by flow cytometry. Freshly isolated monocytes tended to aggregate with platelets, in variable proportions according to individuals; however, platelet-monocyte conjugates were poorly detectable after plating cells (Fig. 1A). Because platelets can also express MerTK (32), we had to rule out their potential contribution to MerTK detection. Therefore, we used the platelet marker CD42b along with CD14 for analyzing MerTK in monocytes, either isolated or conjugated with platelets. Surprisingly, no MerTK expression was clearly found at day 0 in either case, except for rare cells located in the conjugated fraction (Fig. 1B). In cultured monocytes, MerTK was gradually acquired during monocyte-to-macrophage differentiation, and was evident at intermediate stages (day 3) (Fig. 1C, 1D). However, in the absence of growth factors (i.e., CSFs), MerTK could be detected on the cell surface at variable levels, depending on individual experiments. In contrast, in the presence of M-CSF, MerTK upregulation was reproducibly enhanced. GM-CSF, instead, decreased MerTK expression levels (Fig. 1E–G).

Macrophage MerTK expression promptly adapts to changes of immunologic environment

Terminally differentiated macrophages, obtained after 8 d of culture, also upregulated MerTK in the presence of M-CSF, but not GM-CSF (Fig. 2A). We tested whether M-CSF and GM-CSF effects on MerTK expression were reversible. The addition of GM-CSF at day 5 to M-CSF–differentiated macrophages resulted in significant MerTK downregulation at day 8 (Fig. 2A, 2B), while the addition of M-CSF to GM-CSF-driven macrophages tended to increase MerTK expression, yet not significantly (Fig. 2C).

Because GM-CSF and M-CSF are known to drive M1 or M2 macrophage differentiation, respectively (33, 34), we studied the effects of other conventional M1 and M2 stimuli on terminally differentiated macrophages. According to a recent classification, macrophages can be divided into M1 (driven by IFN-γ, LPS, or both), M2a (driven by IL-4 or IL-13), M2b (driven by immune complexes and LPS), and M2c (driven by IL-10, TGF-β, or glucocorticoids) (31). Although M2b and M2c macrophages are elicited by different conditions, both subsets are characterized by IL-10 production, and we focused only on the M2c population, better

FIGURE 1. MerTK is upregulated during monocyte-to-macrophage differentiation, and is further enhanced by M-CSF. (A–D) Monocytes were sorted from healthy human PBMCs through negative selection magnetic beads and analyzed by flow cytometry at day 0, and after 1 and 3 d of culture. Monocytes were gated using an anti-CD14 Ab and stained with anti-MerTK Ab. An anti-CD42b Ab was also used to distinguish platelet-monocyte conjugates from isolated monocytes. (E–G) Monocytes were cultured in complete medium in the presence of M-CSF (50 ng/ml) or GM-CSF (100 ng/ml), or in the absence of CSFs, and analyzed for MerTK expression on day 3. Data shown are representative of three independent experiments. *p < 0.05.
described in humans (31). In either GM-CSF– or M-CSF–differen-
tiated macrophages, stimulation at day 5 with either M1 or M2a
polarizing factors (IFN-γ (10 ng/ml) plus LPS (1 μg/ml), IL-4 (20 ng/ml), or dexamethasone (Dex; 100 nM), for an additional 3 d. MerTK expression was analyzed by Western blot (A), or measured by flow cytometry as mean fluorescence intensity (MFI) fold variation compared with levels obtained with culturing cells with M-CSF alone (B) or GM-CSF alone (C) for 8 d. Data shown are representative of three independent experiments. (D and E) Cells were incubated with Dex (1–1000 nM) in the presence or absence of M-CSF (50 ng/ml) for 3 d in serum-free medium. MerTK upregulation was measured by flow cytometry as MFI fold increase compared with expression levels in untreated cells. Data shown are representative of four independent experiments. (F and G) Cells were cultured in complete medium in the presence of M-CSF (50 ng/ml) or GM-CSF (100 ng/ml), or in the absence of CSFs. On day 5, cells were treated with IL-4 (20 ng/ml) or IL-10 (50 ng/ml), for an additional 3 d. DCs were differentiated in the presence of GM-CSF and IL-4 from day 0 for 8 d. MerTK expression was analyzed by Western blot (F). Densitometry of Western blots was performed to quantify MerTK expression following cell treatment with IL-10, M-CSF, or both (G). Densitometry values were normalized to β-actin and are reported as fold variation compared with MerTK expression levels in untreated cells. Data are representative of three independent experiments. (H) Cells were incubated with GM-CSF or GM-CSF plus IL-4 for 8 d in complete medium, to differentiate M1 macrophages or DCs, respectively. Cells were stained for CD209 (DC-SIGN), CD1a, and MerTK. MerTK+ cells were quantified as percentages of total cells by flow cytometry. *p < 0.05, **p < 0.01, ***p < 0.001.

FIGURE 2. Macrophage MerTK expression promptly adapts to changes in immunologic environment. (A–C) CD14+ cells were cultured in complete medium in the presence of M-CSF (50 ng/ml) or GM-CSF (100 ng/ml). On day 5, cells were treated with GM-CSF (100 ng/ml), M-CSF (50 ng/ml), IFN-γ (10 ng/ml) plus LPS (1 μg/ml), IL-4 (20 ng/ml), or dexamethasone (Dex; 100 nM), for an additional 3 d. MerTK expression was analyzed by Western blot (A), or measured by flow cytometry as mean fluorescence intensity (MFI) fold variation compared with levels obtained with culturing cells with M-CSF alone (B) or GM-CSF alone (C) for 8 d. Data shown are representative of three independent experiments. (D and E) Cells were incubated with Dex (1–1000 nM) in the presence or absence of M-CSF (50 ng/ml) for 3 d in serum-free medium. MerTK upregulation was measured by flow cytometry as MFI fold increase compared with expression levels in untreated cells. Data shown are representative of four independent experiments. (F and G) Cells were cultured in complete medium in the presence of M-CSF (50 ng/ml) or GM-CSF (100 ng/ml), or in the absence of CSFs. On day 5, cells were treated with IL-4 (20 ng/ml) or IL-10 (50 ng/ml), for an additional 3 d. DCs were differentiated in the presence of GM-CSF and IL-4 from day 0 for 8 d. MerTK expression was analyzed by Western blot (F). Densitometry of Western blots was performed to quantify MerTK expression following cell treatment with IL-10, M-CSF, or both (G). Densitometry values were normalized to β-actin and are reported as fold variation compared with MerTK expression levels in untreated cells. Data are representative of three independent experiments. (H) Cells were incubated with GM-CSF or GM-CSF plus IL-4 for 8 d in complete medium, to differentiate M1 macrophages or DCs, respectively. Cells were stained for CD209 (DC-SIGN), CD1a, and MerTK. MerTK+ cells were quantified as percentages of total cells by flow cytometry. *p < 0.05, **p < 0.01, ***p < 0.001.

Remarkably, M-CSF positively synergized with dexamethasone and, in the presence of M-CSF, a 10-fold lower dose of dexamethasone (10 nM instead than 100 nM) was sufficient to reach maximal induction of MerTK; moreover, the addition of M-CSF to dexamethasone 100 nM further enhanced MerTK expression, whereas increasing the dose of dexamethasone per se (1 μM) was no longer effective (Fig. 2D, 2E). IL-10 also synergized with M-CSF in upregulating MerTK (Fig. 2F, 2G). In contrast, GM-CSF
and IL-4 combined to abrogate MerTK expression in human peripheral monocyte-derived DCs (Fig. 2F, 2H). Therefore, MerTK is not only inducible by AC exposure (6), but is also finely regulated by cytokines, growth factors, or both. Loss of MerTK in DCs indicates that MerTK is dispensable for AC clearance in these cells, similar to what has been previously reported in mice (35, 36).

MerTK expression is restricted to (CD163+CD16+CD206+) M2 macrophages

Because MerTK expression was enhanced under M2 conditions, we examined MerTK expression along with several M2 surface receptors to identify potential markers that could predict the presence of MerTK in human macrophages.

Although CD206 (i.e., mannose receptor) was among the first identified markers for conventional (IL-4–driven) M2 macrophages (37), it is a nonspecific M2 marker in humans, being also upregulated by M-CSF, glucocorticoids (Fig. 3), TGF-β (not shown), and even by GM-CSF, a non-M2 factor. We noted instead that CD209 (earlier known as DC-SIGN) was a reliable and specific marker of exposure to IL-4 (M2a polarization), whereas CD163 (hemoglobin-haptoglobin scavenger receptor) or CD16 (FcγRIIIa), in agreement with a recent report (38), were specific markers of exposure to M-CSF, glucocorticoids, IL-10, or TGFβ (M2c polarization). Remarkably, MerTK showed the same expression pattern as CD163. Macrophages that had low levels of CD163 (M1 and M2a) also showed significant MerTK downregulation. However, CD163 detection was not sufficient to identify MerTK+ cells, as in the case of IL-10–treated cells. What best discriminated the macrophage subset that highly expressed MerTK was the coexpression of CD163 and CD16; alternatively, the same population could be identified by the coexpression of CD163 and CD206. Both M-CSF and dexamethasone were able to induce this specific phenotype (CD163+CD16+CD206+; Fig. 3). Therefore, MerTK upregulation by steroids is due to pharmacologic induction of M2c polarization, which reproduces the physiologic effects of M-CSF differentiation.

IL-10 is required for M-CSF to induce M2c differentiation and upregulate MerTK

It was noteworthy that IL-10 could enhance M-CSF effects, although IL-10 per se could not promote further MerTK upregulation compared with medium (Fig. 2F, 2G). We then wondered whether serum-derived bioactive IL-10 present in culture medium (39, 40) might account for the basal M-CSF effects observed. Consistent with this hypothesis, M-CSF upregulated MerTK only in the presence of serum (Fig. 4A). In serum-free medium, the addition of low doses of IL-10 was crucial for M-CSF induction of MerTK expression by the first days of differentiation (Fig. 4B). Conversely, blocking IL-10 in serum-containing medium, through a neutralizing anti–IL-10 Ab, prevented spontaneous increase as well as M-CSF enhancement of MerTK and CD163 expression.

**FIGURE 3.** MerTK expression is restricted to (CD163+CD16+CD206+) M2 macrophages. Macrophages were differentiated from peripheral monocytes for 7–8 d in complete medium, in the presence of GM-CSF (100 ng/ml), IFN-γ (10 ng/ml), IL-4 (20 ng/ml), M-CSF (50 ng/ml), dexamethasone (100 nM), or IL-10 (50 ng/ml). Cells were stained for MerTK, CD14, CD163, CD204/SR-A1, CD16, CD206, and CD209 and analyzed by flow cytometry. Histograms show MFI fold variations compared with levels in untreated cells (basal). Data shown are representative of 8 to 12 independent experiments. *p < 0.05, **p < 0.01, ***p < 0.001.
during differentiation (Fig. 4C). In serum-free conditions, neither M-CSF nor IL-10 alone, but only the combination M-CSF plus IL-10 was able to induce MerTK in macrophages, at levels comparable to those reached with dexamethasone. Interestingly, M-CSF plus IL-10 gave a modest but significant increase in IL-10 receptor (CD210) expression compared with GM-CSF or IL-4. More broadly, we found that the combination M-CSF plus IL-10 was critically required for complete M2c differentiation (e.g., CD163 and CD16 upregulation) in serum-free conditions (Fig. 4D).

Instead, dexamethasone upregulated MerTK and induced M2c macrophages even in serum-free medium (Fig. 4D) or, as previously reported for CD163 (41), in the presence of anti–IL-10 Ab in serum-containing medium (not shown). M2c differentiation was, therefore, IL-10 dependent for M-CSF and IL-10 independent for steroids. M2c (M-CSF plus IL-10) macrophages also differed from dexamethasone-treated cells in having significantly higher levels of CD14 (Fig. 4D), a receptor known to be involved not only in LPS recognition, but also in tethering ACs (9).

Although currently classified as an M2c stimulus (31), TGF-β gave a different phenotype, characterized by CD16 induction, but inhibition of MerTK, CD163, and CD14 expression (Fig. 4D). Contrary to IL-10, TGF-β did not upregulate MerTK even in combination with M-CSF and tended to reduce the upregulating effect of M-CSF plus IL-10 (not shown).

MerTK confers to M2c (M-CSF plus IL-10) macrophages enhanced ability to clear early ACs

We aimed to assess the functional importance of MerTK upregulation in M2c macrophages in regard to AC clearance efficiency. For this purpose, we cultured cells for 7 d in complete medium, adding IFN-γ (M1 stimulus), IL-4 (M2a stimulus), M-CSF, and IL-10 (M2c stimuli) or fresh medium only (M0) from day 4. At day 7, we coincubated macrophages with early apoptotic neutrophils (Fig. 5A), at a 1:5 ratio.

We observed a remarkably higher capacity of AC clearance in M2c (M-CSF plus IL-10) macrophages, compared with the other

**FIGURE 4.** IL-10 is required for M-CSF to induce M2c differentiation and upregulate MerTK. (A) CD14+ cells were cultured in the presence or absence of M-CSF (50 ng/ml), in either serum-containing or serum-free medium, for 3 d. (B) Cells were incubated with IL-10 (1 ng/ml), increasing doses of M-CSF (0.05 to 50 ng/ml), or both. MerTK upregulation was measured by flow cytometry as MFI fold increase compared with expression levels in untreated cells. Data shown are representative of three independent experiments. (C) Cells were cultured in serum-containing medium with or without M-CSF (50 ng/ml), in the presence or absence of a neutralizing mouse monoclonal anti-human IL-10 Ab (5 μg/ml; Biolegend, clone JES3-9D7), for 3 d. Cells were stained for CD163 and MerTK, and they were quantified by flow cytometry as percentages of total cells. (D) Cells were cultured in serum-free medium in the presence of M-CSF (50 ng/ml), IL-10 (50 ng/ml), M-CSF plus IL-10, TGF-β (20 ng/ml), and dexamethasone (Dex; 100 nM) for 4 d. Cells were stained for MerTK, CD163, CD16, and CD163. MerTK expression is shown as MFI fold variation compared with levels in untreated cells, as well as MerTK+ cell percentages; CD163, CD16, and CD14 MFI fold variations are also reported. Data shown are representative of four independent experiments. The expression of IL-10 receptor (IL-10R, or CD210) was measured after 3-d cytokine stimulations. Data shown are representative of three independent experiments. *p < 0.05, **p < 0.01, ***p < 0.001.
FIGURE 5. MerTK confers to M2c (M-CSF plus IL-10) macrophages an enhanced ability to clear early ACs. (A) Early ACs were obtained from incubating human neutrophils, isolated from peripheral blood healthy donors, in 10% FCS-RPMI for 20 h. According to annexin V and propidium iodide (PI) staining, early ACs constituted 65–70% of total neutrophils. (B and C) CFSE-labeled apoptotic neutrophils were added for 60 min to 7-d differentiated M0 (untreated), M1 (IFN-γ, 2.5 ng/ml), M2a (IL-4, 20 ng/ml) and M2c (M-CSF, 50 ng/ml plus IL-10, 50 ng/ml) macrophages, labeled with a fluorochrome-conjugated anti-CD14 Ab at a 5:1 ratio. M2c macrophages showed significantly enhanced ability to clear early ACs, expressed as higher percentages of phagocytic (CFSE+) macrophages. Preincubation of M2c macrophages with a goat polyclonal anti-human MerTK Ab (5 μg/ml; R&D Systems) for 30 min before addition of apoptotic neutrophils abolished such superiority of M2c cells to phagocytose ACs compared with other macrophage subsets. Blocking MerTK diminished not only the number of CFSE+ macrophages, but also the mean phagocytosis activity per single cell, depicted as CFSE MFI. Altogether, it resulted in a significant decrease of the phagocytosis index, determined by multiplying the percentage of CFSE+ macrophages by the CFSE MFI of phagocytic macrophages. Data shown are representative of three independent experiments. (D) By fluorescence microscopy (Leica TCS SP5 confocal laser scanning microscope, 40×/1.25 NA oil objective), M2c macrophages stained for CD14 (red) and CD163 (green) were shown to engulf Hoechst 33342-labeled apoptotic neutrophils (blue; left panel, yellow arrows). Preincubation of M2c macrophages with an anti-MerTK blocking Ab inhibited engulfment, but not the physical interaction between macrophages and apoptotic neutrophils (right panel, white arrows). *p < 0.05, **p < 0.01.

Subsets. Importantly, such superiority of M2c cells was abrogated in the presence of a blocking anti-MerTK Ab (Fig. 5B, 5C). Using immunofluorescence microscopy, we could observe that blocking MerTK inhibited AC engulfment by M2c macrophages, but not the physical interactions between macrophages and ACs (Fig. 5D) that typically precede phagocytosis of early ACs (10).

M2c markers universally define MerTK+ macrophages prone to AC clearance

We showed above that IFN-γ and IL-4 downregulate MerTK protein expression, which is consistent with previous microarray data (42). Nevertheless, after culturing monocytes for 4 d in the presence of IFN-γ or IL-4, we were able to detect rare MerTK+ cells even among M1 and M2a macrophages. Looking at the phenotype of these cells, we could observe that they were clearly distinguishable from the other macrophages by the selective expression of M2c markers (CD163, CD16, CD14, CD204; Fig. 6A, B). In particular, CD204 expression ratio (i.e., percentage of CD204+ cells among MerTK+ macrophages to percentage of CD204+ cells among total macrophages in culture) was significantly higher in M1 conditions, whereas CD163 and CD16 expression ratios were significantly higher in both M1 and M2a conditions (Fig. 6B). These minor populations of M2c macrophages did not occur as a consequence of IFN-γ or IL-4 inefficacy on some cells, because they were less clearly distinguishable among untreated cells; they were, rather, actively induced by treatment. In contrast to MerTK− M2a cells, the MerTK+ M2c macrophages occurring among IL-4–treated cells were CD209 negative and brighter for CD206 (Fig. 6C). Thus, MerTK localizes in macrophage subsets that share the phenotype CD14brightCD16+CD163+CD204+CD206brightCD209null. MerTK+ M2c cells occur even in non-M2c conditions, as small endogenous populations.

We tested whether such minor populations of M2c cells were more capable of performing AC clearance compared with the major populations of M1 and M2a macrophages. As predicted, CD14bright CD163+ cells showed significantly higher ability to clear apoptotic neutrophils compared with CD14dimCD163− cells. Moreover, phagocytosis of ACs by CD14brightCD163+ cells, but not by CD14dimCD163− cells, was significantly inhibited by blocking MerTK (Fig. 6D, 6E).

M2c-like CD14brightCD16+ circulating monocytes use MerTK to phagocytose ACs

We wanted to determine whether the M2c markers were able to predict MerTK positivity and identify efferocytotic cells even among circulating monocytes. We reported above that freshly isolated monocytes analyzed after negative selection did not express MerTK. Nevertheless, we reasoned that negative selection eliminated CD16+ monocytes along with lymphocytes, and that this minor population of more mature (HLA-DR+) monocytes could represent a counterpart in the circulation of M2c macrophages. Thus, in this set of experiments, we looked at fresh monocytes directly from PBMCs, without magnetic sorting. According to CD14 and CD16 expression, three populations of monocytes were distinguishable: a major subset of CD14+CD16+ cells, and small numbers of CD14bright
CD16<sup>+</sup> and CD14<sup>dim</sup>CD16<sup>+</sup> cells (Fig. 7A, 7B). We could detect MerTK<sup>+</sup> monocytes within the CD14<sup>bright</sup>CD16<sup>+</sup> subset, representing approximately one third of this population (Fig. 7C). Detection of MerTK was not associated with a higher rate of platelet-monocyte conjugates in this subset (Fig. 7D); rather, MerTK positivity was associated with the expression of the M2c scavenger receptors CD163 and CD204 (Fig. 7E). CD204 is, in fact, known to be a negative regulator, as shown by its inhibitory effect on IL-4–induced Gas6 secretion (Fig. 8A).

Gas6 levels in the supernatants of serum-free cultures to investigate how MerTK regulation was related to Gas6 production. We found that Gas6 was released upon cell stimulation with IL-10, dexamethasone, or IL-4, but not with GM-CSF or IFN-γ, indicating that both M2c and M2a, but not M1, macrophages were Gas6 producers. Similarly to what we observed with MerTK expression, M-CSF alone had no effect on Gas6 production in serum-free conditions, yet enhanced dexamethasone effects. TGF-β was a negative regulator, as shown by its inhibitory effect on IL-4–induced Gas6 secretion (Fig. 8A).

The Gas6/MerTK pathway is known to inhibit production of proinflammatory cytokines (17, 18) and this function can be independent from AC clearance (19). Consistent with previous results (17, 18), we found that in monocyte-macrophages cultured in the absence of CSFs, Gas6 significantly reduced LPS-induced production of TNF-α (Fig. 8B) and tended to increase IL-10 production (Fig. 8C).

Because IL-10 production represents a central functional property of M2c macrophages (31), we hypothesized that Gas6 effects on IL-10 secretion could be more pronounced in cells cultured in the presence of M-CSF. In these conditions, it was possible that Gas6, inducible by IL-10, could signal through MerTK, inducible by M-CSF in an IL-10–dependent manner, to stimulate additional IL-10 secretion. We indeed found that rhGas6 significantly increased IL-10 levels in supernatants of M-CSF–cultured cells stimulated with low doses of LPS, and this effect was prevented by blocking MerTK activation (Fig. 8D). In the

**FIGURE 6.** M2c markers universally define MerTK<sup>+</sup> macrophages prone to AC clearance. (A–C) CD14<sup>+</sup> cells were cultured in serum-free medium in the presence of IFN-γ (10 ng/ml; M1), IL-4 (20 ng/ml; M2a), M-CSF (50 ng/ml) plus IL-10 (50 ng/ml; M2c), or in the absence of cytokines (M0), for 4 d. Cells were stained for MerTK, CD163, CD14, CD16, and CD204. Coexpression of MerTK and M2c surface markers was studied by flow cytometry (A). For each M2c receptor (CD14, CD163, CD16, CD204), an expression ratio was obtained by dividing “percentage of macrophages expressing a given M2c receptor among total macrophages in culture”, after differentiation in M0, M1, M2a, or M2c conditions. Frequencies of each M2c receptor (percentages of positive cells) among MerTK<sup>+</sup> macrophages and among total macrophages were analyzed for potential significant differences between the two sets of data (B). IL-4–treated cells were also stained for CD209 and CD206 (C). Data shown are representative of four independent experiments. (D and E) CD14<sup>+</sup> cells treated with IFN-γ or IL-4 for 3 d in serum-free medium were coincubated with CFSE<sup>+</sup> apoptotic neutrophils. Percentages of CFSE<sup>+</sup> phagocytic macrophages were determined among the major populations of M1 or M2a cells (CD14<sup>dim</sup>CD163<sup>+</sup>) and the minor populations of M2c-like cells (CD14<sup>bright</sup>CD163<sup>+</sup>). Data from one representative experiment (D) and three independent experiments (E) are reported. *p < 0.05, **p < 0.01.

Gas6 is a major ligand of MerTK; it is produced by several cell types, including macrophages themselves (36, 42). We examined Gas6 levels in the supernatants of serum-free cultures to investigate how MerTK regulation was related to Gas6 production. We found that Gas6 was released upon cell stimulation with IL-10, dexamethasone, or IL-4, but not with GM-CSF or IFN-γ, indicating that both M2c and M2a, but not M1, macrophages were Gas6 producers. Similarly to what we observed with MerTK expression, M-CSF alone had no effect on Gas6 production in serum-free conditions, yet enhanced dexamethasone effects. TGF-β was a negative regulator, as shown by its inhibitory effect on IL-4–induced Gas6 secretion (Fig. 8A).

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The presence of M-CSF, IL-10, and Gas6 were, therefore, able to reciprocally stimulate one another’s production. Gas6 release from MerTK+ M2c cells can, then, be part of an autocrine loop that amplifies IL-10 secretion and positively regulates homeostasis of M2c macrophages.

In the presence of M-CSF, cell secretion of TNF-α was instead negligible. We tested whether the endogenous production of Gas6 by M2c (M-CSF plus IL-10) cells played a role in inhibiting TNF-α production. However, the addition of either recombinant MerFc or a blocking anti-Gas6 Ab to neutralize Gas6 failed to increase TNF-α levels in supernatants (Fig. 8E).

**Discussion**

In this study, we examine the expression of the key AC receptor MerTK in human populations of monocytes and macrophages, determining for each subset the ability to clear ACs and the functional relevance of MerTK in this process. Furthermore, we make a novel contribution to characterization of human anti-inflammatory macrophages, in regard to their phenotype, the immunologic factors promoting their differentiation, and the favoring role of the Gas6/MerTK pathway in mediating IL-10 secretion from these cells.

It has been shown previously that MerTK is regulated by metabolic pathways through LXRs, nuclear sensors activated following macrophage exposure to ACs or to other sources of cholesterol, and through PPARs and RXRs, transcription factors activated during macrophage differentiation (4–6). In this study, we show that this molecule is also importantly regulated by specific immunologic factors. In fact, MerTK is not homogenously distributed among human macrophage populations, but is mostly restricted to a discrete subset of IL-10–secreting anti-inflammatory M2 macrophages.

Recently, Galvan et al. (15) reported that prolonged macrophage exposure to the complement component C1q, another molecule primarily involved in AC clearance, stimulates expression of MerTK.

**FIGURE 7.** M2c-like CD14brightCD16+ circulating monocytes use MerTK to phagocitose ACs. (A–E) Freshly isolated monocytes were analyzed by flow cytometry directly from PBMCs, without magnetic sorting, to include also CD16+ (HLA-DR+) monocytes. On the basis of CD14 and CD16 expression levels, monocytes were divided into three categories: CD14brightCD16− (red histograms and peaks), CD14brightCD16+ (blue) and CD14dimCD16+ (green). Platelet-monocyte conjugates were depicted by flow cytometry as events also positive for the platelet marker CD42b in each monocyte subset (D). Percentages in E refer to positivity of CD14brightCD16+ cells for the receptors indicated. Data shown are representative of four independent experiments. (F–H) Freshly isolated PBMCs were coincubated with CFSE-labeled apoptotic neutrophils at a 1:1 ratio for 4 h. For inhibition studies, cells were preincubated with a goat polyclonal anti-human MerTK Ab (2 μg/ml; R&D Systems) or a goat control IgG (2 μg/ml; Southern Biotech) for 30 min before addition of apoptotic neutrophils. Percentages of CFSE+ phagocytic monocytes were determined within each monocyte subset. Data from three independent experiments (F, G) and one representative experiment (H) are reported. *p < 0.05, **p < 0.01, ***p < 0.001.
and production of Gas6 and C1q itself. Moreover, MerTK was recognized to be essential for C1q-dependent efferocytosis. However, the mechanism accounting for C1q induction of MerTK was not defined. In this regard, it will be interesting to determine whether stimulation with C1q is able to elicit the M2c phenotype. Alternatively, M2c polarization might promote release of C1q, which in its turn could mediate or amplify MerTK upregulation.

M2 differentiation is a key process regulating inflammation and fibrosis (33, 34, 37). The factors controlling this process are just becoming understood. We performed the first systematic study on the effects of cytokines and growth factors on human macrophage phenotype, MerTK expression, and Gas6 secretion. The effects observed are summarized in Table I. Remarkably, MerTK and Gas6 levels follow the expression pattern of CD16 and CD163—molecules identifiable as specific M2c markers—in agreement with a recent study (38). CD206 is highly expressed in these macrophages; however, CD206 is also upregulated in M2a (IL-4–treated) and in GM-CSF-differentiated M1 macrophages. M2a

Table I. Effects of immunologic environment on macrophage phenotype markers, MerTK expression, and Gas6 production

<table>
<thead>
<tr>
<th>Macrophage Differentiation Factors</th>
<th>CD206</th>
<th>CD209</th>
<th>CD16</th>
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<th>CD14</th>
<th>MerTK</th>
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† indicates increase, † indicates decrease, = indicates “no change” in macrophage expression of phenotype markers, MerTK or Gas6 compared to expression in macrophages obtained in the absence of differentiation factors.
cells are selectively characterized by CD209 expression and downregulate surface expression of MerTK. Nevertheless, contrary to what was reported by others (27), we found that M2a macrophages secrete Gas6, consistent with IL-4 effects on Gas6 gene induction (42); this suggests that IL-4 might induce other membrane or soluble TAM receptors.

Our study provides novel insights into mechanisms of M2c polarization in humans. IL-10, TGF-β, and glucocorticoids are the factors currently classified as M2c stimuli (31). We show that M2c macrophages are differentiated in the presence of steroids or in the presence of M-CSF and IL-10. These two types of stimulation have comparable effects on macrophage polarization. Steroids are known to increase a broad range of molecules involved in AC clearance, including MerTK, MFG-E8, C1q, Axl, ADORA3, and thrombospondin-1 (29, 30). We show in this study that MerTK upregulation by these drugs is closely related to induction of M2c polarization. We report for the first time that steroids stimulate macrophages to secrete Gas6, possibly resulting in enhanced MerTK-dependent clearance independent from Protein S (12, 28).

In contrast, we observed that TGF-β gives a different phenotype (CD163+CD16*MerTK*Gas6*), whereas IL-10 gives only a partial phenotype (CD163*CD16+MerTK*Gas6*), that requires M-CSF for its complete expression (CD163*CD16+MerTK*Gas6*).

Incubation of monocytes with M-CSF promotes M2 macrophage differentiation (33, 34). We noted that M-CSF synergizes with either IL-10 or glucocorticoids, thereby identifying M-CSF as a specific M2c factor or cofactor rather than a general M2 cytokine. Additive effects between M-CSF and IL-10 were previously reported. Specifically, IL-10 increased CD16 (FcγRII) and CD32 (FcγRI) expression in M-CSF–driven macrophages, thus enhancing FcγR-mediated phagocytosis in these cells (44); IL-10 alone, instead, was only able to upregulate CD64 (FcγRI) (45). Similarly, we found that MerTK expression and MerTK-mediated AC phagocytosis rely on a combined action of M-CSF and IL-10, whereas IL-10 alone is able to induce only Gas6, in agreement with a recent report (27).

Importantly, we demonstrated that IL-10 is not only an enhancer of M-CSF effects, but is essential for M-CSF to act as an M2c factor and upregulate MerTK. We examined the effects of M2c factors in the presence and absence of serum in cell culture medium, as well as in IL-10–blocked serum-containing and IL-10–supplemented serum-free medium. We concluded that M-CSF–driven M2c differentiation is dependent on IL-10 bioactivity, present in serum-containing medium (39, 40) or exogenously added in serum-free conditions, whereas glucocorticoid-induced polarization is IL-10 independent.

In the absence of M-CSF, IL-10 did not upregulate MerTK, which was consistent with previous microarray studies of IL-10 effects on gene regulation in monocytes-macrophages (46, 47). Only Jung et al. (48) observed Mer gene upregulation by IL-10. However, these authors incubated PBMCs rather than purified monocytes in vitro with IL-10, and only afterward sorted monocytes for microarray analysis. This may have underestimated the contribution of T lymphocytes to MerTK upregulation; in particular, T cells are a well-known source of M-CSF (49). Indeed, Jung et al. (48) also reported IL-10 induction of CD16 and CD32, for which M-CSF is required (44). Of interest, Tiemessen et al. (50) reported that Tregs promote anti-inflammatory monocytes-macrophages expressing CD163 and CD206; however, although CD163 expression was dependent on IL-10 released by Tregs, CD206 induction was suggested to be cytokine independent. In this study, we showed that macrophage coexpression of CD163, CD206, and CD16, is inducible by M-CSF, strongly supporting a central role for T cell–derived M-CSF, together with IL-10, in the induction of anti-inflammatory M2c macrophages by Tregs. From this perspective, production of M-CSF by Tregs might counterbalance the well-known production of GM-CSF by proinflammatory Th1 and Th17 lymphocytes (51), with strong and direct repercussions of the Treg versus Th17 balance on M-CSF versus GM-CSF macrophage differentiation.

Interestingly, MerTK+ M2c-like cells could be found even among circulating CD16+ monocytes. CD16+ monocytes represent more mature monocytes and can phagocytose ACs via CD36 (43). However, only CD14brightCD16+, but not CD14dimCD16+, monocytes express MerTK and the M2c receptors CD163 and CD204; accordingly, their ability to clear ACs is in part MerTK dependent. This finding is consistent with previous findings that CD14bright CD16+ monocytes are predominantly anti-inflammatory and secrete IL-10, whereas CD14dimCD16+ are proinflammatory and produce TNF-α (52, 53). A role for TGF-β in the development of CD16+ monocytes was also suggested (54). We reported that M-CSF and IL-10 trigger CD16 expression along with CD14, CD163, and MerTK, whereas TGF-β upregulates CD16, but not CD14, CD163, or MerTK; therefore, we propose that CD14bright CD16+ and CD14dimCD16+ subsets are selectively elicited by different cytokines.

M-CSF–driven macrophages have already been reported by Xu et al. (55) to have augmented capability of clearing early ACs; however, these authors attributed such ability to enhanced macropinocytosis activity, independent from IL-10. This conclusion was soon challenged by Krysko et al. (10), who demonstrated a major role for macroinocytosis in removing late, rather than early, ACs. In fact, Xu et al. (55) initially showed enhanced AC clearance by M-CSF–driven macrophages, even when late ACs were preponderant compared with early ACs, and suggested the involvement of macroinocytosis in clearance of both early ACs and blebs (i.e., late apoptotic debris). Krysko et al. (10) also argued that Xu et al. did not provide evidence of colocalization of fluid phase tracers with endosomes at microscopy, which is crucial to definitively prove macroinocytosis. Although we do not exclude enhanced macroinocytosis by M-CSF–driven macrophages, we identified MerTK as the major driver of early AC clearance in these cells, dependent on IL-10. In fact, whereas the majority of AC clearance receptors preferentially binds to late ACs (10), MerTK specifically clears early membrane-intact ACs (7, 12). Therapeutic induction of M2c polarization may, therefore, ameliorate the defective clearance of early ACs in diseases such as lupus, in which secondary necrotic cells are believed to be responsible for development of autoimmune and inflammation (21, 22). This may also be beneficial in atherosclerosis, in which secondary necrosis of cholesterol-laden apoptotic macrophages is associated with plaque instability (24).

In addition, we showed that Gas6 is able to significantly enhance IL-10 secretion via MerTK from M-CSF–cultured cells stimulated with LPS. This finding suggests that M-CSF–driven, IL-10–dependent, M2c macrophages may use this pathway as a positive feedback loop to strengthen and prolongate secretion of IL-10 in the microenvironment, facilitating in this way the reestablishment or the persistence of anti-inflammatory conditions. There are striking parallels with the heme oxygenase 1 (HO-1) pathway, inducible by IL-10 and promoting IL-10 secretion in M-CSF–driven CD163+ (i.e., M2c) macrophages (56). Stimulating Gas6 and MerTK activity in CD163+ macrophages to increase IL-10 production may represent a promising strategy to treat some inflammatory diseases in which CD163+ macrophages and IL-10 production are reduced, such as in atherosclerotic lesions (57, 58). Conversely, blocking the Gas6/MerTK pathway in tumor-associated CD163+ macrophages may be critical to reduce IL-10
production and immune tolerance to cancer (59). The prognostic relevance of TAM receptors in cancer is so far uniquely attributed to the aberrant expression of Axl and MerTK in tumor cells, which may promote cell survival and proliferation in response to Gas6 released by tissue macrophages (26, 27). Because tumor-associated macrophages are characterized by an M2c phenotype (59), it is tempting to speculate that MerTK is overexpressed also in these nonmalignant cells that infiltrate and surround the tumor. In this view, Gas6 might foster tumor growth by also acting on macrophages in an autocrine/paracrine manner; macrophage secretion of anti-inflammatory cytokines mediated by MerTK would ultimately result in suppression of antitumor immune responses.

In conclusion, we provide evidence that M2c polarization is desirable to ensure efficient clearance of early ACs by human macrophages and monocytes, owing to intense MerTK up-regulation. The presence of both M-CSF and IL-10 is needed to differentiate M2c macrophages, implying that T cell release of M-CSF, along with IL-10, can be crucial for induction of anti-inflammatory monocytes/macrophages by Tregs. M-CSF and IL-10 may have therapeutic utility in the treatment of autoimmune diseases and atherosclerosis, in which AC clearance is impaired. In addition, modulating MerTK activity in vivo (e.g., through recombinant Gas6 or activating and inhibitory mAbs) may constitute an effective strategy to attenuate or promote innate inflammation with IL-10, can be crucial for induction of anti-inflammatory monocytes/macrophages by Tregs. M-CSF and IL-10 may have therapeutic utility in the treatment of autoimmune diseases and atherosclerosis, in which AC clearance is impaired. In addition, modulating MerTK activity in vivo (e.g., through recombinant Gas6 or activating and inhibitory mAbs) may constitute an effective strategy to attenuate or promote innate inflammation related to autoimmunity, metabolic, and tumoral diseases.

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Disclosures

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